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Spatial analyses of Ediacaran communities at Mistaken Point

Emily G. Mitchell and Nicholas J. Butterfield

Abstract.—Bedding-plane assemblages of Ediacaran fossils from Mistaken Point, Newfoundland, are among the oldest known records of complex multicellular life on Earth (dated to ~565 Ma). The in situ preservation of these sessile but otherwise deeply enigmatic organisms means that statistical analyses of specimen positions can be used to illuminate their underlying ecological dynamics, including the interactions between taxa.

Fossil assemblages on Mistaken Point D and E surfaces were mapped to millimeter accuracy using differentiated GPS. Spatial correlations between 10 well-defined taxa (*Bradgatia*, Charniid, *Charniodiscus*, *Fractofusus*, Ivesheadiomorphs, Lobate Discs, *Pectinifrons*, *Plumeropriscum*, *Hiemalora*, and *Thectardis*) were identified using Bayesian network inference (BNI), and then described and analyzed using spatial point-process analysis. BNI found that the E-surface community had a complex web of interactions and associations between taxa, with all but one taxon (*Thectardis*) interacting with at least one other. The unique spatial distribution of *Thectardis* supports previous, morphology-based arguments for its fundamentally distinct nature. BNI revealed that the D-surface community showed no interspecific interactions or associations, a pattern consistent with a homogeneous environment.

On the E surface, all six of the abundant taxonomic groups (*Fractofusus*, *Bradgatia*, Charniid, *Charniodiscus*, *Thectardis*, and *Plumeropriscum*) were found to have a unique set of interactions with other taxa, reflecting a broad range of underlying ecological responses. Four instances of habitat associations were detected between taxa, of which two (*Charniodiscus–Plumeropriscum* and *Plumeropriscum–Fractofusus*) led to weak competition for resources. One case of preemptive competition between Charniid and Lobate Discs was detected. There were no instances of interspecific facilitation. Ivesheadiomorph interactions mirror those of *Fractofusus* and *Charniodiscus*, identifying them as a form-taxonomic grouping of degradationally homogenized taphomorphs. The absence of increased fossil abundance in proximity to these taphomorphs argues against scavenging or saprophytic behaviors dominating the E-surface community.

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Introduction

Ediacaran organisms occupy a key place in the evolution of life on Earth, straddling the transition between the microbe-dominated world of the Proterozoic and the animal-dominated world of the Phanerozoic. Ediacaran macrofossils are represented by three broadly delineated assemblages (Waggoner 2003), of which the oldest, the Avalonian assemblage, consists of a diverse range of deep-water, sessile organisms (Narbonne 2004). Avalonian organisms share few features with living forms, making their biology, phylogenetic relationships, and ecological interactions difficult to assess (Brasier et al. 2012; Hoyal Cuthill and Conway Morris 2014; Liu et al. 2015; Dufour and McIlroy 2017). Even so, almost all of these Avalonian macroorganisms were sessile (Seilacher et al.

2005; Liu et al. 2011), so their typically in situ preservation provides a direct account of their biological and ecological processes. Significant insights into Avalonian community ecology have been gained from the statistical analysis of specimen positions using spatial point-process analyses (SPPA), with Clapham et al. (2003) demonstrating their nonrandom distribution on the seven principal surfaces at Mistaken Point, SE Newfoundland. More recent work has used computational and mathematical advances in SPPA to tease out the underlying biological processes, including distinctive modes of reproduction (Mitchell et al. 2015).

Almost all spatial analyses of Avalonian communities to date have focused on interactions within a single taxon. Real organisms, however, rarely act in isolation, so any useful

resolution of Avalonian ecology will also need to assess interactions *between* co-occurring taxa. The extent to which these interactions impact community structure depends on a multitude of interlinked factors, including resource availability and the response of constituent taxa to local conditions. Such interactions can be positive, wherein one taxon facilitates the survival of another, or negative, whereby one taxon inhibits another through competition, predation, or chemical exclusion. Combinations of positive and negative interspecific interactions also occur, acting over different temporal and/or spatial scales.

Recent advances of SPPA have demonstrated rich potential to resolve such relationships for communities of sessile organisms (e.g., Wiegand et al. 2007a; Muko et al. 2014). Community-scale spatial distributions depend on the interplay of a number of different factors, most importantly physical environment (Wiegand et al. 2007b), organism dispersal/reproduction (Seidler and Plotkin 2006), competition for resources (Getzin et al. 2006), facilitation between taxa (Lingua et al. 2008), and differential mortality (Getzin et al. 2008). The emergent spatial patterns are rarely discernible to the naked eye (Illian et al. 2008) but can be readily resolved statistically. The distance metric used by Clapham et al. (2003) to analyze Avalonian community structure was "nearest-neighbor analysis," in which the distance from one specimen to another is measured and plotted on a cumulative frequency curve. Such calculations, however, only capture local associations, and overlook more complex or larger-scale spatial patterns: if all specimens occur within 10 cm of each other, for example, then no patterns larger than 10 cm will be detected. Nearest-neighbor analyses also fail to distinguish different types of aggregation, such as those due to vegetative reproduction (e.g., stolon-like clustering) versus small-scale habitat preference (Mitchell et al. 2015). More generally, model comparisons using nearest-neighbor distances are challenged by the difficulty of visualizing the distribution shape or magnitude.

In contrast to nearest-neighbor analysis, pair-correlation functions (PCFs) describe complex spatial distributions over large distances, providing an "organism's point of view" of the surrounding community by quantifying how density changes with increasing distance from the average specimen (Law et al. 2009). In addition to within-taxon or "univariate" analyses (e.g., Mitchell et al. 2015), PCFs can be used to describe how the spatial density of one taxon changes relative to another—"bivariate" PCFs. Distinct modes of interspecific interaction can be detected as nonrandom distributions between taxa either as aggregation/clustering (closer together than complete spatial randomness [CSR]), segregation (further apart than CSR), or some combination of these patterns (e.g., segregated clusters). This property means that bivariate PCFs provide a description of the scale, magnitude, and shape of interspecific spatial distributions, capturing complex patterns across a wide range of spatial scales. Comparison of different spatial models (model-fitting analyses) can be performed using Monte Carlo simulations and goodness-of-fit tests to compare how well they fit observed data (Diggle 2003; Illian et al. 2008). If the model fits the data well (i.e., if the goodness-of-fit test p-value $p_d < 0.05$), the hypothesized process is strongly supported. Further information can be teased from spatial patterns using random-labeling analyses (RLAs) to assess the relative differences of density-dependent behavior between taxon pairs. Because interspecific interactions collectively define community structure, they can also be used to address how a community develops from initial colonization to mature community (succession) and how different taxa affect the community as a whole.

Not every correlation is causal of course, so demonstration of interspecific associations does not in itself translate to (direct) ecological interaction. In modern systems, indirect correlations can be distinguished from direct interactions by analyzing the dynamics of the whole system using Bayesian network inference (BNI) (e.g., Milns et al. 2010), in which the joint probability distributions between variables (here the densities of each taxon population) is represented graphically (see Milns et al. 2010: Appendix B for a Bayesian network overview). Once direct links between taxa have been established using BNI, the nature of those links

can then be investigated using bivariate SPPA. Deployed in tandem, BNI and SPPA offer a powerful means of reconstructing ecological structure from spatial data. In this study, we apply it to the question of Avalonian community ecology.

Materials and Methods

To assess the interspecific dynamics of Ediacaran Avalonian communities, we carried out a detailed BNI and SPPA of the D and E surfaces of the Mistaken Point Formation in SE Newfoundland (Fig. 1). These two surfaces host the most abundant and best preserved communities of Avalonian macrofossils on record (Liu et al. 2015), dated to 565 ± 3 Ma (Benus 1988). The fossils are preserved as external molds in siltstone hemipelagites, cast from above by volcaniclastic deposits (Wood et al. 2003; Ichaso et al. 2007). This study uses the data set of Mitchell et al. (2015), comprising 2977 fossil specimens from the E surface and 1402 specimens from the D surface (Supplementary Fig. S1). The mean accuracy of the GPS coordinates of the data was 0.460 ± 0.06 cm horizontally and 0.8260 ± 0.11 cm vertically. All fossils with a consistently recognizable form were assigned to 10 taxonomic groupings (cf. Clapham et al. 2003): *Bradgatia*, Charniid, *Charniodiscus*, *Fractofusus*, *Hiemalora*, Ivesheadiomorphs, Lobate Discs, *Pectinifrons*, *Plumeropriscum*, and *Thectardis*. For completeness, all remaining specimens were placed in one of two "bin groups": Holdfast Discs and Other Species (see Supplementary Fig. S2 and Section S1 for taxonomic definitions).

Spatial Analyses Performed on the D and E Surfaces at Mistaken Point

Two types of spatial analyses were performed on the spatial data of the D and E surfaces: BNI followed by SPPA (see Supplementary Section S2 for extended methods). Performing BNI prior to SPPA analyses ensures that bivariate correlations are not incorrectly linked to an interspecific ecological process, by distinguishing a connected series of indirect correlations between several taxa from one direct correlation between two taxa. Such BNI screening is not required with univariate PCF analyses, or bivariate analyses between differing size classes, because there is no possibility of indirect correlations. In principle, careful bivariate SPPA should be able to distinguish direct from indirect correlations; however, when dealing with problematic fossils, such as those of the Ediacaran Avalon, BNI provides a key check on whether the signals found are indeed genuine direct correlations.

Bayesian Network Inference.—The set of correlations or "edges" between taxon pairs (networks) for both D and E surfaces was identified using BNI (Heckerman et al. 1995). To find the best-fit network for each surface, the mapped areas were first divided into quadrats and discretized into zero, low, and high densities. Then 100 samples were created by bootstrapping (subsampling) these quadrat data sets at the 95% level (Magurran 2013), and the Bayesian network was calculated using the software Banjo (Smith et al. 2006). The resulting set of edges formed a bimodal distribution, represented by either rare–low occurrences or high occurrences. High-occurrence edges (determined using 'mclust';
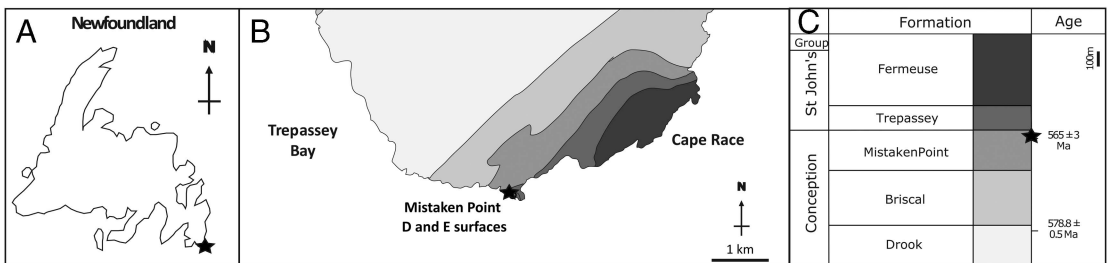
FIGURE 1. Locality map showing: A, the location of Mistaken Point Ecological Reserve in Newfoundland, Canada; and B, the locations of D and E surfaces within the reserve, with the formations shown in grayscale, as given in C. C, Stratigraphic column showing radiometric dates and the locations of D and E surfaces (star). (Modified from Liu 2016: Fig. 1E,F.)

Fraley et al. 2012) are the constituent edges of the underlying network, while the low-occurrence edges are random "noise" and do not correspond to actual correlations (cf. Yu et al. 2004); the results are depicted in the form of network diagrams. The interaction strength is defined as the relative weight or strength of each high-occurrence edge output by Banjo, where 1 represents a strong positive correlation, −1 is a very strong negative correlation, and 0 is a nonmonotonic correlation (i.e., positive and negative at different spatial scales). Where a correlation between taxa was asymmetric (i.e., the strength and/or nature of one taxon on the other was not identical), this correlation was indicated on the network diagram by an arrow. Mann-Whitney tests were used to compare the effects of taxon, edge removal, and directionality and to assess the relative importance of each to the network.

Spatial Point-Process Analyses.—Unlike the BNI assessment of whole-community structure, PCF analyses can only consider an individual taxon (univariate) or pairs of taxa (bivariate) during one set of analyses. To describe an entire community structure, all individuals within the community need to be accounted for, hence the inclusion of obvious organ–taxa (*Hiemalora*) and bin groups (Holdfast Discs and Other Species) in the BNI analyses. At the same time, however, the taxonomic indeterminacy of such groups is likely to obscure any paleoecological signal based on their spatial distributions. As such, these groups have been omitted from follow-up PCF analyses and discussion. Putative taphomorph taxa, which may include carcasses and associated decay-induced microbial colonies (cf. Darroch et al. 2013) such as Ivesheadiomorphs and Lobate Discs, are included in the PCF analyses because they are both morphologically and taphonomically distinct from all of the other taxonomic groups. Form-taxonomic groups that are likely to include a range of "natural" taxa (e.g., Holdfast Discs, Other Species, *Hiemalora*) do not have equivalent extant ecological models with which to interpret the correlations, so they are excluded from the bivariate PCF analyses.

Initial data exploration and heterogeneous Poisson modeling were performed in R using the package 'spatstat' (Baddeley et al. 2015;

Supplementary Methods S2). Three types of SPPA were used to determine the most likely underlying process behind each interspecific interaction found using BNI: (1) PCFs, (2) model fitting to the PCFs, and (3) RLAs. Programita was used to find PCFs and to perform aggregation model fitting (Wiegand and Moloney 2004, 2013; Wiegand et al. 2006, 2009). Monte Carlo simulations and Diggle's goodness-of-fit test were used to compare the fit of different spatial model PCFs to the observed PCFs (the p-value p_d, in which $p_d = 1$ indicates a complete model fit, and $p_d = 0$ indicates no fit [Diggle 2003; Illian et al. 2008]) and to determine the occurrence of habitat associations, competition, and facilitation between taxon pairs. Note that the p_d is not a probability, but more akin to the coefficient of determination (R^2) in a linear regression, wherein the value represents the percentage of the data described by the model. Finally, RLAs were used to detect density-dependent mortality processes (Jacquemyn et al. 2010; Raventós et al. 2010) using Monte Carlo simulations and calculating the difference in the extent to which each taxon departs from random labeling; that is, whether the two populations exhibit density dependence. Because this measure is a difference between two PCF quotients, $p_d{}^{RLA} = 0$ corresponds to a random RLA pattern between the two taxa, indicating no density dependence; by contrast, $p_d{}^{RLA} = 1$ indicates no distribution overlap and spatial patterning that is fully density dependent. These spatial analyses and their relationships to ecological processes are outlined below; the underlying mathematics is described in detail by Wiegand and Moloney (2004) and Wiegand et al. (2006).

Using Spatial Point-Process Analyses to Detect and Describe Interspecific Interactions

In the case of sessile communities, there are just four principal interspecific processes that influence interspecific distributions: (1) habitat associations (colocation of two taxa on the same habitat), (2) competition (one or both taxa limit a mutual resource), (3) facilitation (one taxon enhances the survival of another), and (4) differential mortality/density-dependent

effects (Wiegand et al. 2007a). Each of these processes is best described by a distinct spatial point-process model, so comparison of observed bivariate spatial distributions to these models can resolve the underlying process(es) (Diggle 2003; Wiegand and Moloney 2004). Although these models have been developed primarily in the context of terrestrial forest ecology, the underlying principles are equally applicable to other communities of sessile organisms, including fungal sporocarps (e.g., Liang et al. 2007) and nonmotile animals (e.g., Muko et al. 2014).

Bivariate or pairwise PCFs describe how the densities between specimens belonging to two different taxa change with spatial scale, thus capturing interspecific patterns. A bivariate PCF = 1 corresponds to two populations that exhibit CSR with respect to one another; that is, they have no pairwise spatial structure and their bivariate distribution can be modeled by a homogeneous Poisson process (Illian et al. 2008). Accordingly, if a homogeneous Poisson model is the best fit to the data (i.e., exhibits CSR), then the constituent organisms do not exhibit any significant interactions. Insofar as most co-occurring taxa in most ecosystems are likely to exhibit some degree of interaction, sessile communities with no spatial structure are unusual, typically seen only during early establishment and/or when resources are unlimited (e.g., Greig-Smith 1979; Lin et al. 2011; Wiegand et al. 2012).

By contrast, a bivariate PCF ≠ 1 indicates statistically significant aggregation (PCF > 1) or segregation (PCF < 1) between taxa. The magnitude of the PCF reflects the intensity of underlying biological and physical processes: two taxon populations with a PCF = 4, for example, are four times more aggregated than if they exhibited CSR; thus, the relative magnitudes of the PCFs can be used to compare relative strengths of interactions and associations.

Habitat Associations.—Habitat associations occur when two (or more) taxa have the same environmental preferences, such as the aggregation of alpine tree species at a common altitude (Wang et al. 2011) or aggregations around a patchy distribution of soil nutrients (John et al. 2007). Where interspecific interactions derive from such habitat associations, they can

be modeled by shared-source models (also called shared-parent models) in which the two sets of taxa aggregate around the same set of mutually exclusive points; that is, the focus of the taxon clusters are points that are not biological taxa but some other "environmental" factor (Wiegand et al. 2007a). Where habitat associations are shared between more than two taxa, they can be described by heterogeneous Poisson models, whereby specimen density is modeled by a random process in which density varies across the sample area depending on the given variable (e.g., altitude). Differing sensitivities to habitat heterogeneities can be encapsulated by using differing radii to form the heterogeneous Poisson model that describes the habitat (Supplementary Appendix S2). Use of smaller radii to form the heterogeneous background corresponds to a stronger (more sensitive) reaction to the habitat, while use of larger radii corresponds to a weaker (more diffuse) reaction. Heterogeneous habitats formed from different substrate variations are quantified by different parameters of the bivariate shared-source model, such as the mean patch/cluster radius, the total number of patches/clusters, and the mean number of specimens within each patch/cluster. When two bivariate models have different parameterizations, two distinct underlying substrate variations will have been identified, a reflection of ecological differences between the associated taxa.

Competition between Taxa.—Among sessile organisms, competition over limited resources is typically expressed in the form of reduced specimen density, a process known as thinning. In forests, thinning of fast-growing but shade-intolerant pioneer species occurs as a consequence of progressive light competition (Getzin et al. 2006). In spatial terms, such interspecific competition is detected as segregation between taxa, wherein the segregation is demonstrably not due to an association with a habitat that is itself patchy (in which case the pattern would fit a bivariate shared-source or heterogeneous Poisson model; see Wiegand et al. 2007a).

Interspecific competition is scale-dependent, with preemptive and interference competition acting on smaller spatial scales, and resource competition acting over larger spatial scales. In modern corals, for example, segregation often

occurs when one taxon excludes another by preemptively occupying substrate space, yielding a system in which the size of individual coral heads corresponds to local levels of (preemptive) competition (McCook et al. 2001). In this case the pattern can be recognized spatially by small-scale thinning or segregation, which occurs at similar magnitudes to the occupying taxon. At larger scales, however, such direct interspecific competition may derive from the active release of inhibitory biochemicals (allelopathy), disabling the settlement and/or survival of their neighbors and yielding a quantitatively distinct spatial distribution (Willis 2007). By assessing small-scale segregation in the context of body size, it is possible to distinguish these distinct types of competition. Large-scale segregation can be distinguished from mutual association over a habitat of segregated patches by assessing whether heterogeneous Poisson models (i.e., local habitat heterogeneities) can be used to describe the large-scale segregations (Wiegand et al. 2007b). Thus, comparison of the spatial scales of segregations and of segregated distributions using heterogeneous Poisson and shared-source models (to detect habitat associations) can be used to detect competition and infer the most likely underlying process.

Facilitation.—Interspecific facilitation is a process whereby the presence of one taxon benefits the survival of another (e.g., Brooker et al. 2008). Facilitation can be physical, such as protection from harsh conditions or the creation of suitable habitat (Jones et al. 1997), or intimately biotic, as encountered in mutualistic endosymbioses (e.g., Bruno et al. 2003). In spatial terms, facilitation is indicated when the best-fit bivariate is a linked-cluster or double-cluster model (also known as bivariate Neymann-Scott/Thomas cluster models). In linked-cluster models, the spatial pattern is described by a series of clusters with normally distributed densities (i.e., Thomas clusters) centered on the positions of the facilitating taxa (Dickie et al. 2005; Dale and Fortin 2014). A linked Thomas double-cluster pattern is induced when the facilitated taxon (which forms Thomas clusters around the facilitating taxon) reproduces to form secondary Thomas clusters, resulting in Thomas double clusters,

each centered on specimens of the facilitating taxon. Mutualisms are best modeled by a heterogeneous Poisson process with a density created from the joint density of the two taxa, which indicates mutual clustering with no external driving factor.

Density-dependent Mortality Processes.—Mortality rates within a population can vary due to a variety of density-dependent processes, potentially influencing community spatial distributions and structure. For example, the high biodiversity of tropical forests appears to be maintained through Janzen-Connell effects, whereby taxon-specific consumers such as herbivores or pathogens generate (positive) density-dependent mortality of their prey taxa. Janzen-Connell effects prevent any single taxon from dominating the community, thus maintaining high community diversity (Velázquez et al. 2016). Mortality processes can be investigated using RLAs (Goreaud and Pélissier 2003; Raventós et al. 2010), wherein the spatial positions of the organisms are held constant, but a specimen property such as size class or preservation detail is randomly shuffled among the observed spatial positions using Monte Carlo simulations. If the observed pattern falls outside this generated simulation envelope, then the specimen property (subgroup) is nonrandomly distributed within the community population and a density-dependent process is detected. RLAs have been used to interrogate the effects of fire and canopy gaps on forest ecosystems and postfire succession (De Luis et al. 2008; Getzin et al. 2008). They can also be employed to compare the spatial distributions of superficially distinct forms, such as juveniles or differentially degraded carcasses.

Application of Spatial Point-Process Analyses to Identify Taphomorphs

Identification of taphomorphs is crucial to ensure an accurate ecological understanding of Avalonian communities, ensuring that any impact of time-averaging can be identified and the relative importance of decay-based processes assessed. Unlike extant communities, in which living and dead specimens are

easily distinguished, identifying preserved taphomorphs is impeded within Avalonian communities, because the distinction between true morphological characters and taphonomically degraded characters is not well defined (e.g., Antcliffe et al. 2015); and more generally Avalonian morphological characters are limited in number compared with Phanerozoic organisms (e.g., Dececchi et al. 2017). However, comparison of the bivariate spatial distributions of putative taphomorphs with those of non-taphomorph taxa can be used to establish both the likelihood of taphomorphic affinity and, for taphomorphs, their precursor taxa: whereby, the spatial distributions of taphomorph taxa are expected to mirror those of their precursor taxa, that is, to be nonunique. Note that these analyses assume that there is no patchy small-scale degradation (reflected by intraspecific segregation), a pattern that is not encountered on the D or E surfaces (Mitchell et al. 2015). In this context, taphomorphs are expected to exhibit a random distribution within their precursor taxon populations and to have similar bivariate correlations. Ecological interpretations of taphomorph distributions need to take their affinity into account, because when taphomorphs mirror the spatial distributions of their precursors, a competitive-type spatial distribution can be detected, for example, not because the taphomorphs are directly competing with another taxon, but because the precursor taxon to the taphomorph did compete with that taxon.

Ivesheadiomorphs and Lobate Discs (Supplementary Fig. S2J, F) are the two dominant putative taphomorphs found within Mistaken Point communities (Clapham et al. 2003). Ivesheadiomorphs are characterized by a lack of consistent internal or external form, low preservation detail, and rarity of symmetrical features (Liu et al. 2011). Lobate Discs are also relatively irregular compared with other taxonomic groups such as the rangeomorphs, but they are characterized by a circular shape with high relief, and approximately radially symmetrical, irregular lobes. Both putative taphomorphs have a high morphological variability within identified specimens that, when coupled with differences in spatial distributions due to the differing impact of

local habitat heterogeneities and/or differing times of community development, means that their affinities should be assessed on a surface-by-surface basis. In other words, identification of any taphomorph and its precursor taxa on one surface may not apply to another surface (Kenchington and Wilby 2014).

Results and Discussion

Our analyses focused on the D- and E-surface communities of Mistaken Point, SE Newfoundland. The E surface is the most diverse and abundant assemblage of Avalonian fossils on record, preserving 14 distinct species (with 8 abundant taxa) and 2977 identifiable specimens within its 63.5 m^2 area (Clapham et al. 2003; Mitchell et al. 2015). The Mistaken Point D surface is also taxonomically diverse, preserving 8 identifiable species (with 3 abundant taxa) within its 62.0 m^2 area and 1402 individual fossils. Combined BNI and advanced SPPA provided an unprecedented level of ecological insight into the two principle surfaces, and revealed ecological distinctions between them beyond differences in community composition (cf. Clapham et al. 2003).

BNI

E Surface.—On the E surface, BNI analysis revealed a complex network of interspecific interaction and associations between the 11 taxonomic groups identified in this study (Fig. 2). Out of a possible 66 correlations, we identified 14 interspecific correlations involving all but one of the constituent taxa, *Thectardis*. On average, each taxon interacted with at least two other taxa (2.41 mean per taxon correlation); though interaction strengths were skewed toward low values (0.17 mean interaction strength). There were four nonmonotonic correlations (different positive and negative correlations at different spatial scales), one negative correlation, and nine positive correlations (Table 1). *Fractofusus* and *Plumeropriscum* were the most connected taxa, with four correlations each: *Fractofusus* correlated with *Plumeropriscum*, Ivesheadiomorphs, Lobate Discs, and Holdfast Discs; *Plumeropriscum* correlated with *Fractofusus*, *Charniodiscus*,
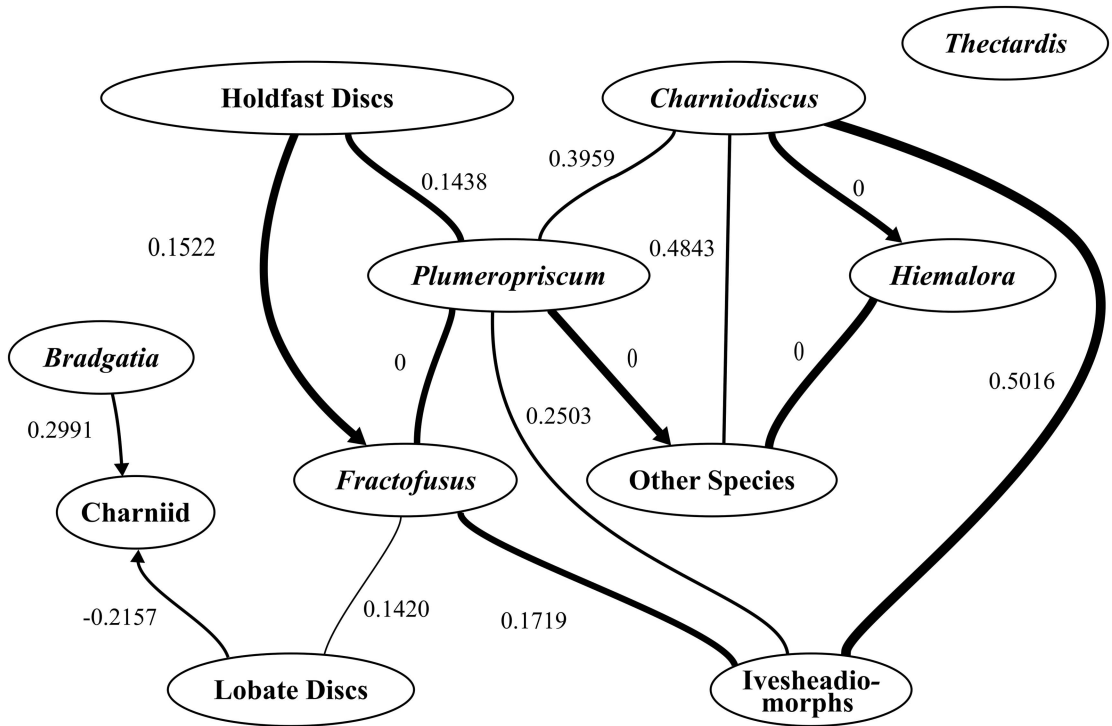
FIGURE 2. Bayesian network for the E surface. Correlations are indicated by lines connecting the taxon pairs. The occurrence rate is indicated by the width of the edge (the line depicting correlation between two taxa); the wider the line, the higher the occurrence rate. Arrows indicate nonmutual dependence between two taxa; for example taxon 1 is aggregated with respect to taxon 2, but taxon 2 is not aggregated with respect to taxon 1. Numbers by the lines are the mean interaction strengths of the correlations, with positive interaction strengths indicating aggregation, negative interaction strengths indicating segregation, and zero indicating different aggregation and segregation behaviors at different densities.

Holdfast Discs, and Other Species. *Bradgatia* was the least connected taxon within the network, only correlating with the Charniid group. Excluding *Hiemalora* and the two bin groups resulted in eight interspecific correlations among the eight remaining taxa—with a mean per taxon correlation of 2.28 and a mean interaction strength of 0.19.

The only taxon on the E surface to show no interspecific interactions or associations was *Thectardis* (based on 10,000,000 networks from 100 95% bootstrap samples; Fig. 2), which could be selectively excluded from the analyses without changing the network ($p = 0.54$; Fig. 2; Table 1). This result is not due to low abundance of *Thectardis* (<1% of E-surface specimens): the BNI data are input using discrete values, so the absolute abundance does not influence the network output. Moreover, taxa with similar abundances (*Hiemalora* and *Bradgatia*) interact with other taxa even at the lower sampling of 80% BNI (Table 1). As such,

the absence of *Thectardis* from the recovered network is most likely to be a real ecological signal, demonstrating its ecological isolation from other E-surface taxa. In addition to its conspicuously unconnected spatial distribution, *Thectardis* also stands out morphologically, distinguished by a simple triangular outline (Supplementary Fig. S3D) and absence of the fractal or frondose differentiation seen in most co-occurring forms. Sperling et al. (2011) have speculated on its possible sponge affiliations, but with little corroborating evidence (Antcliffe et al. 2014). The ecological disparity of *Thectardis* is consistent with *Thectardis* having a different feeding mode than the other E-surface taxa, for example, the active filter feeding that sponges employ, versus the passive osmotrophic feeding of the Rangeomorphs. As such, our spatial analyses are consistent, but not definitive, in the support of a sponge affinity for *Thectardis*.

TABLE 1. Edge properties for the Bayesian network given in Fig. 4. The column labels are defined as follows: Removal *p*-value: The *p*-value for whether the network score (the fit of the network onto the data) is significantly different if that correlation is removed. Direction: The *p*-value for whether the network score is significantly different if the direction of that correlation is reversed. Mean IS: The mean interaction strength of all the networks bootstrapped at 95%. Size seen (m): The smallest quadrat size at which the correlation is detected. Bootstrapping 95%: The occurrence rate for the 95% bootstrapped networks that indicates consistency of correlation over different quadrats of the bedding plane. Bootstrapping 90–50%: The occurrence rates for the bootstrapped data when subsampled at the 50%, 60%, 70%, 80%, and 90% levels. See SI Methods for bootstrapping details. NA indicates mutual correlations. The following taxon notation is used: Brad, *Bradgatia*; Char, Charniid: Chard, *Charniodiscus*: Plum, *Plumeropriscum*: Fract, *Fractofusus*: Hiem, *Hiemalora*: Ives, Ivesheadiomorph: Lob, Lobate Discs.

Correlation	Removal *p*-value	Direction	Mean IS	Size seen (m)	Bootstrapping					
					95%	90%	80%	70%	60%	50%
Brad–Char	0.0497	0.10	0.2990	1.50	60	54	56	48	28	29
Lob–Char	0.0002	0.06	−0.2157	0.50	57	56	59	54	51	29
Fract–Lob	0.0006	NA	0.1420	1.00	68	67	60	54	50	54
Fract–Ives	0.0046	NA	0.1719	1.75	83	69	44	42	36	32
Plum–Fract	0.0008	NA	0.0000	1.75	66	45	35	31	30	13
Disc–Fract	0.0000	0.07	0.1522	0.50	96	88	86	74	36	48
Plum–Ives	0.0000	NA	0.0021	1.00	52	49	46	42	38	19
Disc–Plum	0.0171	NA	0.1432	1.50	68	52	36	29	32	19
Chard–Plum	0.0000	NA	0.3959	0.50	63	68	71	62	61	69
Chard–Hiem	0.0011	0.01	0.0000	1.00	75	61	54	15	17	22
Plum–Other	0.0010	0.06	0.0000	1.50	94	77	69	48	42	42
Hiem–Other	0.0063	NA	0.0000	1.50	99	83	75	56	38	16
Chard–Other	0.0016	NA	0.4843	1.75	59	51	53	47	37	32
Ives–Chard	0.0001	NA	0.5016	1.75	100	71	70	50	60	67

D Surface.—Despite the comparably abundant and diverse assemblage of the D surface, BNI analysis found no bivariate correlations between the three abundant taxa (*Fractofusus*, *Bradgatia*, and *Pectinifrons*), while the remaining five (*Plumeropriscum*, *Charniodiscus*, Ivesheadiomorphs, Charniid, and Lobate Discs) occur in too low an abundance (<30 specimens/taxon) to yield statistically significant results. As such, there are no reportable correlations or interaction strengths for the D surface and no potential for follow-up SPPA analysis. The only nonrandom PCF distribution on the D surface is the univariate distribution of *Fractofusus* (Mitchell et al. 2015).

The marked disparity in bivariate correlations between the D and E surfaces points to fundamental differences in their respective community structures, though only if the respective data sets are taphonomically comparable. It is notable, for example, that the overall density of fossils on the D surface is ~50% that of the E surface (Clapham et al. 2003; Mitchell et al. 2015), which could potentially derive from differential modern erosion (cf. Matthews et al. 2017). Our subsampling of the E-surface community to a similar density as the D surface (50%), however, found that 3 of its 14 correlations were still readily detectable (Table 1), demonstrating significant signal retention. Moreover, the three abundant D-surface taxa have a total density comparable to that of their counterparts on the E surface (2.27–22.40 specimens/m^2 vs. 0.62–20.70 specimens/m^2, respectively), while the E-surface pairwise densities of both *Bradgatia* + Charniid and Charniid + Lobate Discs are greater than the pairwise densities of D-surface taxon pairs. At least in the case of these taxa and pairwise densities, it is clear that the absence of correlations on the D-surface is not an artifact of differential preservation or sampling. The differences are ecological.

Another possible interpretation of the disparity in BNI results between E and D is that the lower specimen density of D represents an early stage of community succession, yet to put pressure on local resources (Townsend et al. 2003); that is, the organisms have not yet reached sufficient densities to be impacted by each other or their habitat. The presence of at least two generations of *Fractofusus* (Mitchell et al. 2015), however, demonstrates substantial community development. As such, the absence

of ecological interactions, or even interspecific correlation, on the D surface is more likely to reflect a more homogeneous background environment than that of E surface; in the absence of habitat/resource patchiness, there was minimal competition. Habitat heterogeneity is generally correlated to taxonomic diversity (e.g., Agarwal 2008), so the reduced diversity of the D surface might be a simple consequence of the homogeneous environment.

Spatial Point-Process Analyses of the E Surface

Using bivariate PCF analyses, we found seven of the eight E-surface correlations to be habitat associations and one to be the result of preemptive competition (Fig. 3; Supplementary Table S1). Within the habitat associations, two were devoid of any discernible interaction beyond colocation, three exhibited nonunique or mirrored behaviors (Supplementary Table S2), and two showed clear evidence of subsequent behavior. There was no evidence of facilitation between any of the taxa on the E surface.

Preemptive Competition.—Lobate Discs segregated from Charniid on spatial scales smaller than 0.5 m at a PCF of 0.75, reducing Charniid density by 21% ($PCF_{min} = 0.79$; Fig. 3A; Supplementary Table S1). This small-scale segregation of Lobate Discs and Charniid is of the same spatial scale as the mean radius of Lobate Discs (Clapham et al. 2003), suggesting that the segregation is due to preemptive competition of Charniid not settling directly on the Lobate Discs. An alternative taphonomic explanation, in which Charniid is preferentially degraded in association with Lobate Discs, is unlikely given the absence of such effects with co-occurring rangeomorphs such as *Bradgatia* or *Plumeropriscum*. Allelopathy also causes small-scale segregation, but in this case the Lobate Discs–Charniid correlation exhibits the same spatial scale as the univariate Lobate Discs (albeit nonsignificant) aggregation (Fig. 3A), suggesting that the underlying phenomenon did not extend more broadly, the expected pattern with chemical exclusion. That said, other chemical processes could result in such segregation if the inhibitory compound(s) remained localized within the Lobate Discs–occupied substrate, one possibility being the hydrogen sulfide scenario proposed by Dufour and McIlroy (2017).

Interpreting the mechanisms behind the Lobate Discs–Charniid segregation is substantially hampered by the problematic nature of Lobate Discs, which lack a formal taxonomical definition or basic biological resolution; current interpretations range from a distinct macroscopic taxon to microbial colonies, taphomorphs, or even sedimentary intrusions (e.g., Laflamme et al. 2011; Liu et al. 2011). Nonetheless, the statistical evidence for segregation is unambiguous and demonstrates that the Lobate Discs were largely in place before Charniid establishment.

The absence of allelopathy within the E-surface community contrasts with extant marine sessile communities, in which such chemical-based exclusion and other direct interference competition such as overgrowth competition is pervasive (e.g., Jackson and Buss 1975; Engel and Pawlik 2000). Modern communities of sessile deep-sea organisms also tend to be densely packed, with corals and sponges commonly occupying 90% or more of available substrate (e.g., Calle 2010). By contrast, even the relatively dense E-surface community has conspicuously lower substrate occupation rates (<13%; Clapham et al. 2003), reflecting correspondingly lower levels of substrate competition. Even so, the univariate Charniid PCF shows strong environmentally mediated spatial distributions (Supplementary Fig. S3; Mitchell et al. 2015), suggesting that they may be more susceptible to the local substrate differences induced by the presence of Lobate Discs.

Unique Habitat Associations.—PCF analyses found that seven bivariate correlations were most likely due to habitat associations, wherein both the constituent taxa experienced enhanced survival related to some (otherwise unseen) aspect of the background environment (Fig. 4A). Two of these seven unique habitat associations resulted in large-scale aggregations (*Bradgatia*–Charniid and *Fractofusus*–Lobate Discs; Fig. 3B,C), but without reducing the longer-term survival of either taxon (as would have been detected by segregation in the bivariate PCFs). The *Bradgatia*–Charniid
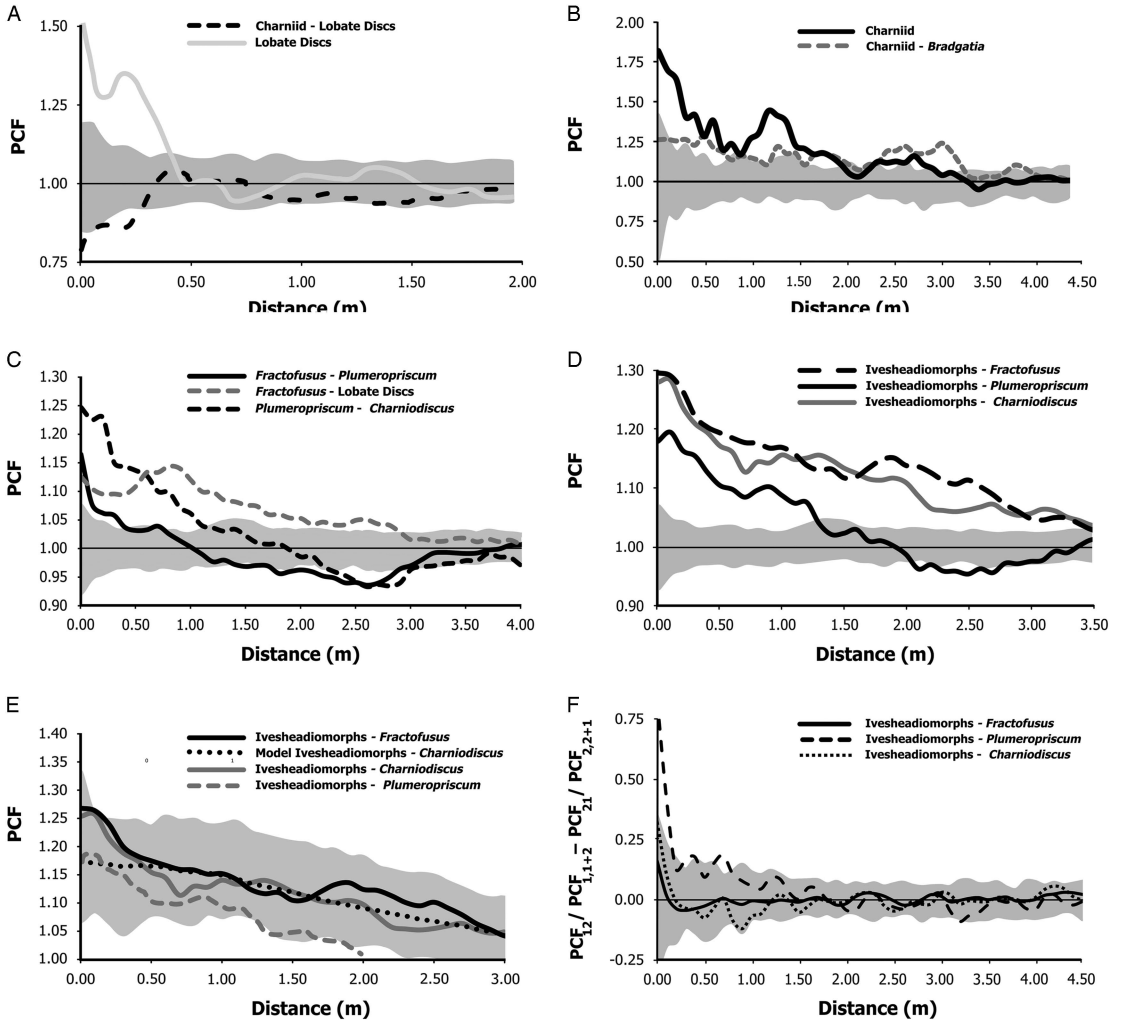
FIGURE 3. Pair-correlation functions (PCFs) for the E surface. The x-axis is the interpoint distance between organisms in meters. On the y-axis, PCF = 1 indicates complete spatial randomness (CSR), < 1 indicates segregation, and >1 indicates aggregation. A, Charniid–Lobate Discs bivariate distribution and Lobate Discs univariate distribution. Gray shaded area is the boundaries of 99 Monte Carlo simulations of the CSR bivariate distribution. B, Charniid–*Bradgatia* bivariate distribution and Charniid univariate distribution. Gray shaded area is the boundaries of 99 Monte Carlo simulations of the CSR bivariate distribution. C, PCFs of non-Ivesheadiomorph bivariate distributions of *Fractofusus* and *Plumeropriscum*. Gray shaded area is the boundaries of 99 Monte Carlo simulations of the CSR bivariate distribution of *Fractofusus*–*Plumeropriscum*. D, Bivariate PCFs of Ivesheadiomorphs interactions. Gray shaded area is the boundaries of 99 Monte Carlo simulations of the CSR bivariate distribution of Ivesheadiormorphs–*Plumeropriscum.* E, Bivariate PCFs of Ivesheadiomorphs interactions showing the best-fit shared-source model of Ivesheadiomorphs–*Charniodiscus.* Gray shaded area shows the boundaries of 99 Monte Carlo simulations for the best-fit shared-source model of Ivesheadiomorphs–*Charniodiscus.* F, Random-labeling analysis results for the Ivesheadiomorphs distributions. On the y-axis, PCF = 0 indicates CSR. Gray shaded area is the boundaries of 99 Monte Carlo simulations of the CSR randomly labeled distributions.

correlation is a relatively consistent aggregation that corresponds to a mutual density increase of *Bradgatia* and Charniid by 15% above CSR up to a 3.4-m radius ($PCF_{max} = 1.15$; Fig. 3B). The greatest aggregation of *Bradgatia* and Charniid occurs under an 0.8-m radius ($PCF_{max} = 1.3$) and

is best modeled by a shared-source model ($p_d = 0.66$; Fig. 3B). This habitat heterogeneity also impacts Charniid, which is inferred by the similar spatial scales of the aggregated part of the univariate Charniid and bivariate Charniid–*Bradgatia* spatial distributions, but only very
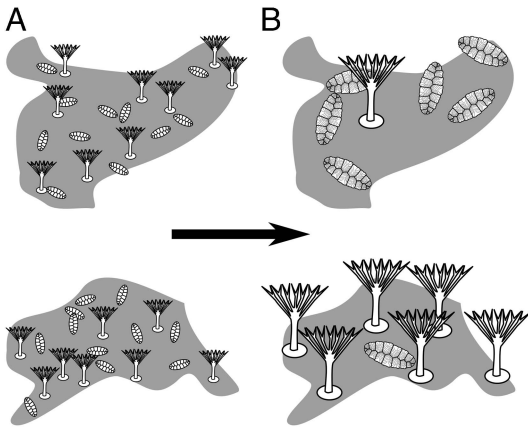
FIGURE 4. A, Reconstruction of the habitat association between *Fractofusus–Plumeropriscum* (SI Fig. 2I and 2B, respectively) resulting in mutual aggregation on top of the inferred heterogeneous habitat (gray shown in column A). In the case of *Plumeropriscum–Fractofusus* (and *Plumeropriscum–Charniodiscus,* not figured), these habitat associations result in competition for resources between mature specimens over time (depicted by the arrow), leading to thinning (segregation) of these larger individuals shown in column B. Note that only single generations are depicted to ensure the clarity of the process. More realistically, there would be multiple generations and different size classes (since these taxa reproduce continuously [Darroch et. al. 2013]).

weakly affects *Bradgatia*, as reflected in its random univariate PCF (Fig 3A–C). The Charniid–*Bradgatia* spatial pattern is much weaker than that of the univariate Charniid aggregation, suggesting that this habitat heterogeneity strongly promotes Charniid establishment and only weakly promotes *Bradgatia* establishment. A similar pattern is observed in corals, where settlement is dependent on the type of previously established algae (cf. Carlon and Olson 1993). Univariate Charniid and the bivariate Charniid–*Bradgatia* are the only PCFs that correlate at this spatial scale (Fig. 3A–C), suggesting that this habitat heterogeneity did not affect other taxa on a meter scale.

The E-surface *Fractofusus*–Lobate Discs correlation is consistent but weak ($PCF_{max} = 1.15$) up to a 2-m radius between these two taxa; it is best modeled by a shared-source model ($p_d = 0.70$; Fig. 3C). The association between *Fractofusus* and Lobate Discs does not exhibit the same spatial scaling as other univariate or bivariate spatial distributions (Fig. 3A–C) and could not be modeled by another best-fit bivariate model (Supplementary Table S2): as

with *Bradgatia* and Charniid, it is a unique association. Notably, this pattern is a rare instance in which spatial pattern is evident in the field, with *Fractofusus* commonly observed to overlie Lobate Discs (cf. Dufour and McIlroy 2017).

Habitat Associations Leading to Competition.— Both *Plumeropriscum–Fractofusus* and *Plumeropriscum–Charniodiscus* exhibit small-scale aggregation coupled with large-scale segregation (Fig. 3C). In the case of *Fractofusus* and *Plumeropriscum*, both taxa exhibit increased densities over a shared paleoenvironmental heterogeneity (as revealed by being best modeled by a shared-source $p_d = 0.79$; Fig. 3C; Supplementary Table S1), coupled with segregation ($PCF_{min} = 0.92$ between 1.5 m and 3.5 m, reducing the established specimen density by 8.4%; Fig. 3C). Notably, this segregation could not be modeled by a heterogeneous Poisson model (Supplementary Tables S1 and S4). Such patterns most likely derive from interspecific resource competition, as growth leads to progressive thinning and spatial segregation of the most mature specimens (cf. Mason et al. 2003; Fig. 4A,B). This same style of habitat association and resource competition is seen between *Plumeropriscum* and *Charniodiscus* and is similarly best modeled by a shared source ($p_d = 0.97$; Fig. 3C; Supplementary Table S1). The habitat associations between *Plumeropriscum–Fractofusus* and *Plumeropriscum–Charniodiscus* were significantly different, pointing to two distinct underlying aggregation sources ($p_d = 0.04$ and $p_d = 0.01$; Supplementary Tables S1 and S2), for example, variations in microbial mat coverage or depth.

Out of a possible 13 taxon pairs on the E surface, it is notable that interspecific resource competition is limited to just these two instances. The limited instances of interspecific competition stand in stark contrast to extant marine benthic communities and other sessile communities (such as terrestrial forests), where competition is ubiquitous (Bertness and Leonard 1997; Wiegand et al. 2007b). These spatial differences also undermine the suggestion that Avalonian communities were similar in structure to extant benthic communities (Clapham et al. 2003). At the same time,

however, it is these three competing taxa—*Fractofusus*, *Plumeropriscum*, and *Charniodiscus*—that form the majority (58.6%) of the specimens on the E surface. In other words, although the majority of *taxa* do not exhibit interspecific resource competition, the majority of *individuals* appear to be competing for limited resources.

Importance of Using BNI Combined with Spatial Point-Process Analyses.—SPPA applied to all possible pairwise combinations on the E surface revealed six nonrandom correlations not found by BNI. These derive from the combined effects of two unrelated correlations rather than direct interaction. Lobate Discs, for example, have a nonrandom PCF with respect to *Plumeropriscum*, but because this correlation was not found by BNI, the signal should be interpreted as a consequence of each taxon being separately correlated with *Fractofusus*. The presence of indirect, nonrandom PCFs that have a clear connection via an intermediate taxon highlights the importance of using BNI in combination with SPPA to minimize type I errors (false positives) when drawing paleoecological inferences.

Using Spatial Analyses to Identify Putative Taphomorphs

The D- and E-surface communities contain abundant populations of Ivesheadiomorphs and Lobate Discs, both of which have been widely viewed as the degraded taphomorphs of more "biological" taxa (Liu et al. 2011). Such interpretation clearly bears critically on any spatial analysis of Avalonian ecology, particularly on the E surface, where these two problematic forms represent 13.2% of all individuals and 46.7% of the total fossil area, a proxy for biomass (Clapham et al. 2003). Our PCF and RLA analyses strongly support the identification of Ivesheadiomorphs as taphomorphs and identify the likely precursor taxa, but find that spatial distributions of Lobate Discs are inconsistent with a taphomorph affinity.

Ivesheadiomorphs' Affinity and Precursor Taxa.—PCF analyses found that *Fractofusus*–Ivesheadiomorphs was best modeled by a shared-source model ($p_d = 0.65$; Fig. 3D,E; Supplementary Table S1) as was *Charniodiscus*–

Ivesheadiomorphs ($p_d = 0.82$). Neither the *Fractofusus*–Ivesheadiomorphs nor the *Charniodiscus*–Ivesheadiomorphs best-fit models were unique, and further analyses strongly suggest that *Charniodiscus* and *Fractofusus* are the precursor taxa of Ivesheadiomorphs on the E surface (cf. Liu et al. 2011) as follows: First, the *Fractofusus*–Ivesheadiomorphs and *Charniodiscus*–Ivesheadiomorphs correlations closely follow the univariate *Fractofusus* and *Charniodiscus* clustering (they exhibit the same small-scale high aggregation under 0.4 m (Fig. 3E). Second, the best-fit models for both correlations are heterogeneous Poisson models based on *Fractofusus* (for the Ivesheadiomorphs–*Fractofusus* correlation, $p_d = 0.55$) or *Charniodiscus* (for the Ivesheadiomorphs–*Charniodiscus* correlation, $p_d = 0.75$) densities. Likewise, the *Fractofusus*–Ivesheadiomorphs distribution could be modeled by the *Plumeropriscum*–*Fractofusus* correlation ($p_d = 0.56$) and vice versa ($p_d = 0.71$). And finally, the RLAs show that Ivesheadiomorphs are randomly distributed within both the *Fractofusus* and *Charniodiscus* populations ($p_d^{RLA} < 0.05$ for both; Fig. 3F; Supplementary Table S3).

BNI of the E surface revealed a correlation between Ivesheadiomorphs and *Plumeropriscum*, which is best modeled by a shared-source model ($p_d = 0.69$; Fig. 3D,E; Supplementary Table S1), with a large-scale segregation of 95% CSR occurring between 2.0 and 3.5 m. However, corresponding SPPA rules out any precursor–taphomorph correspondence: the PCF of the Ivesheadiomorphs–*Plumeropriscum* distribution differs significantly from the univariate *Plumeropriscum*; the bivariate correlation of the two "taxa" cannot be modeled as a heterogeneous Poisson model using *Plumeropriscum*, and RLA also shows significantly different density-dependent behavior of *Plumeropriscum* relative to Ivesheadiomorphs, and there is further overlap with other bivariate models (Supplementary Tables S2 and S4): the Ivesheadiomorphs–*Plumeropriscum* correlation can be modeled by the same best-fit model as *Fractofusus*–*Plumeropriscum* ($p_d = 0.58$ for non-CSR PCF) and by *Charniodiscus*–*Plumeropriscum* ($p_d = 0.55$ for non-CSR PCF), pointing to closely comparable bivariate habits between Ivesheadiomorphs and both *Fractofusus* and

Charniodiscus (that the combined 58% and 55% of these two spatial distributions exceed 100% is explained by a modest 25% overlap in the two models).

These three Ivesheadiomorphs correlations (Ivesheadiomorphs–*Fractofusus*, Ivesheadio-morphs–*Charniodiscus*, and Ivesheadiomorphs–*Plumeropriscum*) are notable in being the only observed correlations on the D and E surfaces that were nonunique. In other words, the spatial distributions of these pairwise correlations are statistically indistinguishable from the bivariate distributions of other pairs—in marked contrast to all other non-Ivesheadiomorphs correlations (Supplementary Table S2). The similarity of these Ivesheadiomorphs correlations with other E-surface correlations makes a microbial affinity (cf. Laflamme et al. 2011) unlikely. Further, the inclusion of non-frondose *Fractofusus* within the Ivesheadiomorphs correlations is inconsistent with the interpretation of Ivesheadiomorphs as sediment intrusions beneath fronds (cf. Wilby et al. 2011), since there was no space between recumbent *Fractofusus* and the substrate that could be infilled by sediment.

Putative Non-Ivesheadiomorphs Taphomorphs.—Apart from Ivesheadiomorphs, it has been argued that Lobate Discs (Liu et al. 2011), *Thectardis*, and *Charniodiscus* (Antcliffe et al. 2015) may also represent preservational variants of other (mostly unspecified) entities (as opposed to organ taxa, such as *Hiemalora*). Possible precursor taxa for Lobate Discs on the E surface are (the holdfasts of) Charniid, *Charniodiscus*, *Bradgatia*, and *Plumeropriscum*. None of these exhibit statistically similar bivariate spatial correlations, however, substantially undermining any taxonomic connection (Fig. 3D,E). Indeed, there are no frondose taxa on the E surface that could have been plausible precursors of Lobate Discs. By the same token, neither *Charniodiscus* nor *Thectardis* have univariate and bivariate spatial patterns that are statistically comparable to any co-occurring taxa, so they are unlikely to be taphomorphic variants (contra Antcliffe et al. 2015) (Mitchell et al. 2015; Fig. 2; Supplementary Fig. S3; Supplementary Table S2).

Multiple Successions.—On a more general level, it is possible that the fossils preserved on any particular bedding surface belong to a succession of discrete communities; that is, not all were alive at the same time (cf. McIlroy and Garton 2010; Liu et al. 2012; Antcliffe et al. 2015). The BNI and SPPA results, however, are inconsistent with multiple colonization of the E surface, as are the univariate population analyses of Darroch et al. (2013) and the SPPA of Mitchell et al. (2015). Significantly, size-distribution analyses of the E surface demonstrate single (not multiple) populations of abundant taxa (with the exception of *Thectardis*; Darroch et al. 2013), and univariate spatial analyses find that there is no evidence of strong univariate environmental (habitat) influences in any E-surface taxa except Charniid (which eliminates bivariate correlations due to repeated successions being subject to the same environmental influence; Mitchell et al. 2015). As such, seven of the eight E-surface taxa can be recognized as contemporaneous. This reasoning cannot be applied to *Thectardis*, since it is not spatially correlated with any other taxon; however, its shared current alignment with frondose taxa on the same bedding surface suggests that it too was part of this community (Clapham et al. 2004).

Taphomorph Ecology.—Within Ediacaran communities, the habitat heterogeneity produced by decaying organisms has been proposed as the determining factor in community structure (Antcliffe et al. 2015; Budd and Jensen 2017; Liu et al. 2015; Dulfour and McIlroy 2017). In this "Ediacaran-fall" model, localized concentrations of resources represented by macro-carcasses are likened to more recent whale-fall or wood-fall ecosystems (c.f. Smith et al. 2015), whereby decaying organisms form the focus of for colonizers to settle upon and form communities. When organisms feed directly on this carbon (or indirectly on the sulfides) produced by specimen decay, then the population densities of scavenger/saprophytic species tend to increase around the local resource, resulting in a spatial pattern best modeled as a linked cluster (or double-cluster) model or a heterogeneous Poisson model based on carcass densities (Wiegand and Moloney 2013). Notably, no such spatial patterns are observed with Ivesheadiomorphs on the Mistaken Point surfaces. Instead there is a mutual clustering around two different shared sources for Ivesheadiomorphs, one with
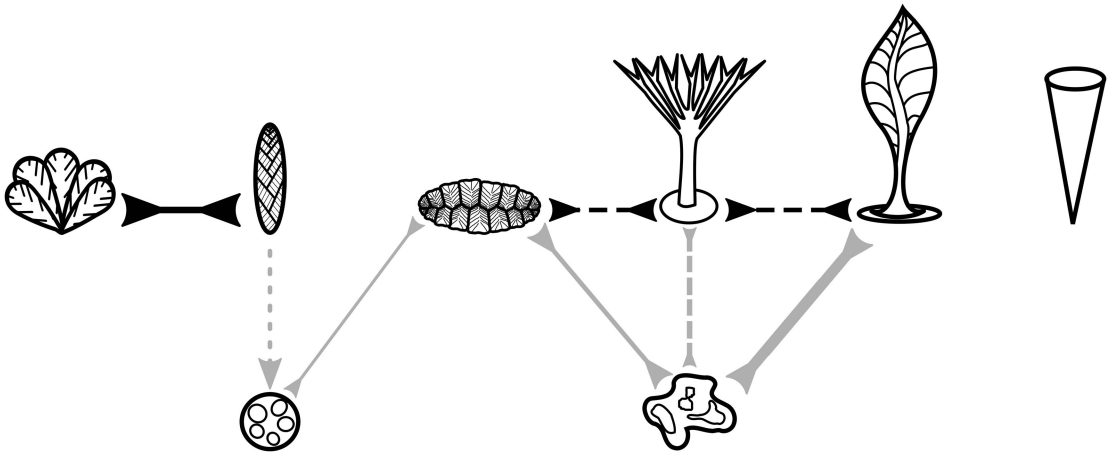
FIGURE 5. Summary diagram of the E-surface bivariate interactions and associations. Top row from left: *Bradgatia*, Charniid, *Fractofusus*, *Plumeropriscum*, *Charniodiscus*, and *Thectardis*. Bottom row from left: Lobate Discs and Ivesheadiomorphs. Gray lines denote taxa with unknown or disputed affinities, while black indicates interactions and associations between living-at-time-of-burial organisms. Two inward arrows indicate mutual habitat associations; solid lines indicate only these habitat associations, while long dashes indicate that these associations result in resource competition. Arrow with dotted line depicts preemptive competition.

Fractofusus and the other with *Charniodiscus* (Fig. 3D; Supplementary Table S1); that is, there is no increased survival for taxa that settle near Ivesheadiomorphs. The correlations between Ivesheadiomorphs–*Fractofusus* and Ivesheadiomorphs–*Charniodiscus* are most likely a reflection of the single-taxon clustering that occurs for both these taxa, with dead specimens (i.e., Ivesheadiomorphs) randomly distributed among the living. Thus, our spatial analyses refute the hypothesis that any single taxon (such as *Fractofusus*, *Charniodiscus*, or *Plumeropriscum*) was a scavenger. Alternatively, a habitat heterogeneity formed from unpreserved decayed carcasses could be detected as a habitat association between taxa. However, if several taxon pairs were impacted by the same single-decay heterogeneity, then they would be expected to share the same shared-source model, which is inconsistent with the four distinct habitat associations present on the E surface and the absence of any such associations or interactions on the D surface. Taken together, SPPA analysis rules out saprophytic habits as a controlling factor in structuring these Avalonian communities.

Conclusions

Our spatial analyses of the E surface at Mistaken Point reveal a fundamentally more complex community structure than has been previously recognized (Fig. 2), showing clear ecological differentiation between taxa, with all six abundant non-taphomorph taxa developing different responses to a variety of habitat spatial variations (Fig. 5; Supplementary Table S2). The presence of multiple ecological responses demonstrates that individual taxa have distinct approaches for adapting to local habitat, leading to divergent selection, reproductive isolation, and ultimately biodiversification (Kawecki and Ebert 2004; Mitchell-Olds and Schmitt 2006; Hereford 2009; Sobel et al. 2010; Futuyma and Agrawal 2009). The fundamentally different degree of organismal interaction in these Avalonian communities demonstrates their fundamentally non-uniformitarian nature, highlighted by the complete lack of D-surface community interspecific interactions, and could well account for the conspicuously slow nature of their evolutionary turnover (cf. Grazhdankin 2004). At the same time, the low levels of competitive interactions (e.g., in comparison to modern forests or deep-sea reefs) established in these earliest communities of macroscopic organisms sets the stage for progressive escalation; first through elevated competition for resources, then leading to macroscopic movement and, ultimately, carnivory.

This study has demonstrated how new approaches to spatial analysis can resolve key aspects of Avalonian paleoecology. By combining BNI with PCFs, it is possible to describe the spatial variation of specimen densities between taxon pairs, establishing when taxa are responding to one another and/or their habitat. Model fitting takes these analyses beyond descriptive statistics, enabling verifiable predictions to be made and ecological hypotheses to be tested. RLA enables the comparison of density-dependent behavior within a given spatial pattern, thus assisting in the identification of taphomorphs. These techniques present a framework for further investigations, which can incorporate morphological details such as body size to investigate broader ecological themes such as the implications of tiering and/or ecological successions on Ediacaran community structure.

Acknowledgments

Literature Cited

Agarwal, S. K. 2008. Fundamentals of ecology. APH Publishing, New Delhi.

Antcliffe, J. B., R. H. T. Callow, and M. D. Brasier. 2014. Giving the early fossil record of sponges a squeeze. Biological Reviews 89:972–1004.

Antcliffe, J. B., A. D. Hancy, and M. D. Brasier. 2015. A new ecological model for the ~565 Ma Ediacaran biota of Mistaken Point, Newfoundland. Precambrian Research 268:227–242.

Baddeley, A., E. Rubak, and R. Turner. 2015. Spatial point patterns. methodology and applications with R. CRC, Boca Raton, Fla.

Benus, A. P. 1988. Sedimentological context of a deep-water Ediacaran fauna (Mistaken Point, Avalon Zone, eastern Newfoundland). In E. Landing, G. M. Narbonne, and P. Myrow, eds. Trace fossils, small shelly fossils and the Precambrian–Cambrian boundary. New York State Museum and Geological Survey Bulletin 463:8–9. Albany, N.Y.

Brasier, M. D., J. B. Antcliffe, and A. G. Liu. 2012. The architecture of Ediacaran fronds. Palaeontology 55:1105–1124.

Bertness, M.D., and G.H. Leonard. 1997. The role of positive interactions in communities: lessons from intertidal habitats. Ecology 78:1976–1989.

Brooker, R. W., et al. 2008. Facilitation in plant communities: the past, the present, and the future. Journal of Ecology 96:18–34.

Bruno, J. F., J. J. Stachowicz, and M. D. Bertness. 2003. Inclusion of facilitation into ecological theory. Trends in Ecology and Evolution 18:119–125.

Budd, G. E., and S. Jensen. 2017. The origin of the animals and a "Savannah" hypothesis for early bilaterian evolution. Biological Reviews 92:446–473.

Calle, S. R. 2010. Ecological aspects of sponges in mesophotic coral ecosystems. Ph.D. dissertation. University of Puerto Rico, Mayagüez. Masters Abstracts International 49(3).

Carlon, D. B., and R. R. Olson. 1993. Larval dispersal distance as an explanation for adult spatial pattern in two Caribbean reef corals. Journal of Experimental Marine Biology and Ecology 173:247–263.

Clapham, M. E., G. M. Narbonne, and J. G. Gehling. 2003. Paleoecology of the oldest known animal communities: Ediacaran assemblages at Mistaken Point, Newfoundland. Paleobiology 29:527–544.

Clapham, M. E., G. M. Narbonne, J. G. Gehling, C. Greentree, and M. M. Anderson. 2004. Thectardis avalonensis: a new Ediacaran fossil from the Mistaken Point biota, Newfoundland. Journal of Paleontology 78:1031–1036.

Dale, M.R. T., and M. J. Fortin. 2014. Spatial analysis: a guide for ecologists. Cambridge University Press, New York.

Darroch, S. A. F., M. Laflamme, and M. E. Clapham. 2013. Population structure of the oldest known macroscopic communities from Mistaken Point, Newfoundland. Paleobiology 39:591–608.

Dececchi, T. A., G. M. Narbonne, C. Greentree, and M. Laflamme. 2017. Relating Ediacaran fronds. Paleobiology 43:171–180.

De Luis, M., J. Raventós, T. Wiegand, and J. C. González-Hidalgo. 2008. Temporal and spatial differentiation in seedling emergence may promote species coexistence in Mediterranean fire-prone ecosystems. Ecography 31:620–629.

Dickie, I. A., S. A. Schnitzer, P. B. Reich, and S. E. Hobbie. 2005. Spatially disjunct effects of co-occurring competition and facilitation. Ecology Letters 8:1191–1200.

Diggle, P. 2003. Statistical analysis of spatial point patterns, 2nd ed. Arnold, London.

Dufour, S. C., and D. McIlroy. 2017. Ediacaran pre-placozoan diploblasts in the Avalonian biota: the role of chemosynthesis in the evolution of early animal life. Geological Society of London Special Publication 448:211–219.

Engel, S., and J. R. Pawlik. 2000. Allelopathic activities of sponge extracts. Marine Ecology Progress Series 207:273–282.

Fraley, C., A. E. Raftery, and L. Scrucca. 2012). mclust version 4 for R: normal mixture modeling for model-based clustering, classification, and density estimation. Technical Report No. 597. Department of Statistics, University of Washington, Seattle.

Futuyma, D. J., and A. A. Agrawal. 2009. Macroevolution and the biological diversity of plants and herbivores. Proceedings of the National Academy of Sciences USA 106:18054–18061.

Getzin, S., Ch. Dean, F. He, J. A Trofymow, K. Wiegand, and T. Wiegand. 2006. Spatial patterns and competition of tree species in a Douglas-fir chronosequence on Vancouver Island. Ecography 29:671–682.

Getzin, S., T. W., K. Wiegand, and F. He. 2008. Heterogeneity influences spatial patterns and demographics in forest stands. Journal of Ecology 96:807–820.

Goreaud, F., and R. Pélissier. 2003. Avoiding misinterpretation of biotic interactions with the intertype K12-function: population independence vs. random labelling hypotheses. Journal of Vegetation Science 14:681–692.

Grazhdankin, D. 2004. Patterns of distribution in the Ediacaran biotas: facies versus biogeography and evolution. Paleobiology 30:203–221.

Greig-Smith, P. 1979. Pattern in vegetation. Journal of Ecology 67:755–779.

Heckerman, D., D. Geiger, and D. M. Chickering. 1995. Learning Bayesian networks: the combination of knowledge and statistical data. Machine Learning 20:197–243.

Hereford, J. 2009. A quantitative survey of local adaptation and fitness trade-offs. American Naturalist 173:579–588.

Hoyal Cuthill, J. F., and S. Conway Morris. 2014). Fractal branching organizations of Ediacaran rangeomorph fronds reveal a lost Proterozoic body plan. Proceedings of the National Academy of Sciences USA 111:13122–13126.

Ichaso, A. A., R. W. Dalrymple, and G. M. Narbonne. 2007. Paleoenvironmental and basin analysis of the late Neoproterozoic (Ediacaran) upper Conception and St. John's groups, west Conception Bay, Newfoundland. Canadian Journal of Earth Sciences 44:25–41.

Illian, J., A. Penttinen, H. Stoyan, and D. Stoyan. 2008. Statistical analysis and modelling of spatial point patterns. Statistics in practice Vol. 70. Wiley, Chichester, U.K.

Jackson, J. B. C., and L. Buss. 1975). Allelopathy and spatial competition among coral reef invertebrates. Proceedings of the National Academy of Sciences USA 72:5160–5163.

Jacquemyn, H., P. Endels, O. Honnay, and T. Wiegand. 2010. Evaluating management interventions in small populations of a perennial herb *Primula vulgaris* using spatio-temporal analyses of point patterns. Journal of Applied Ecology 47:431–440.

John, R., et al. 2007. Soil nutrients influence spatial distributions of tropical tree species. Proceedings of the National Academy of Sciences USA 104:864–869.

Jones, C. G., J. H. Lawton, and M. Shachak. 1997. Positive and negative effects of organisms as physical ecosystem engineers. Ecology 78:1946–1957.

Kawecki, T. J., and D. Ebert. 2004. Conceptual issues in local adaptation. Ecology Letters 7:1225–1241.

Kenchington, C. G., and P. R Wilby. 2014). Of time and taphonomy: preservation in the Ediacaran. *In* M. Laflamme, J. D. Schiffbauer, and S. A. F. Darroch, eds. Reading and writing of the fossil record: preservational pathways to exceptional fossilization. Paleontological Society Papers 20:101–122.

Laflamme, M., J. D. Schiffbauer, and G. M. Narbonne. 2011. Deep-water microbially induced sedimentary structures (MISS) in deep time: the Ediacaran fossil Ivesheadia. Microbial mats in siliciclastic depositional systems through time. SEPM Special Publication 101:111–123.

Law, R., J. Illian, D. F. R. P. Burslem, G. Gratzer, C. V. S. Gunatilleke, and I. A. U. N. Gunatilleke. 2009. Ecological information from spatial patterns of plants: insights from point process theory. Journal of Ecology 97:616–628.

Lin, Y. C., L. W. Chang, K. C. Yang, H. H. Wang, and I. F. Sun. 2011. Point patterns of tree distribution determined by habitat heterogeneity and dispersal limitation. Oecologia 165:175–184.

Liang, Y., L. D. Guo, X. J. Du, and K. P. Ma. 2007. Spatial structure and diversity of woody plants and ectomycorrhizal fungus sporocarps in a natural subtropical forest. Mycorrhiza 17:271.

Lingua, E., P. Cherubini, R. Motta, and P. Nola. 2008. Spatial structure along an altitudinal gradient in the Italian central Alps

suggests competition and facilitation among coniferous species. Journal of Vegetation Science 19:425–436.

Liu, A. G. 2016. Framboidal pyrite shroud confirms the "death mask" model for moldic preservation of Ediacaran soft-bodied organisms. Palaios 31:259–274.

Liu, A. G., D. Mcilroy, J. B. Antcliffe, and M. D. Brasier. 2011. Effaced preservation in the Ediacara biota and its implications for the early macrofossil record. Palaeontology 54:607–630.

Liu, A. G., D McIlroy, J. J. Matthews, and M. D. Brasier. 2012. A new assemblage of juvenile Ediacaran fronds from the Drook Formation, Newfoundland. Journal of the Geological Society 169:395–403.

Liu, A. G., C. G. Kenchington, and E. G. Mitchell. 2015. Remarkable insights into the paleoecology of the Avalonian Ediacaran macrobiota. Gondwana Research 27:1355–1380.

Magurran, A. E. 2013. Measuring biological diversity. Wiley, Oxford, U.K.

Mason, B., et al. 2003. Continuous cover forestry in British conifer forests. Forest Research Annual Report and Accounts 2004:38–53.

Matthews, J. J., A. G. Liu, and D. McIlroy. 2017. Post-fossilization processes and their implications for understanding Ediacaran macrofossil assemblages. Geological Society of London Special Publication 448:251–269.

McCook, L., J. Jompa, and G. Diaz-Pulido. 2001. Competition between corals and algae on coral reefs: a review of evidence and mechanisms. Coral Reefs 19:400–417.

McIlroy, D., and M. Garton. 2010. Realistic interpretation of ichnofabrics and palaeoecology of the pipe-rock biotope. Lethaia 43:420–426.

Milns, I., C. M. Beale, and V. A. Smith. 2010. Revealing ecological networks using Bayesian network inference algorithms. Ecology 91:1892–1899.

Mitchell, E. G., C. G. Kenchington, A. G. Liu, J. J. Matthews, and N. J. Butterfield. 2015. Reconstructing the reproductive mode of an Ediacaran macro-organism. Nature 524:343.

Mitchell-Olds, T., and J. Schmitt. 2006. Genetic mechanisms and evolutionary significance of natural variation in *Arabidopsis*. Nature 441:947.

Muko, S., I. K. Shimatani, and Y. Nozawa. 2014. Spatial analyses for non-overlapping objects with size variations and their application to coral communities. Journal of Animal Ecology 83:980–990.

Narbonne, G. M. 2004. Modular construction of early Ediacaran complex life forms. Science 305:1141–1144.

Raventós, J., T. Wiegand, and M. De Luis. 2010. Evidence for the spatial segregation hypothesis: a test with nine year survivorship data in a Mediterranean shrubland. Ecology 91: 2110–2120.

Seidler, T. G., and J. B. Plotkin. 2006. Seed dispersal and spatial pattern in tropical trees. PLoS Biology 4:e344.

Seilacher, A., L. A. Buatois, and M. G. Mángano. 2005. Trace fossils in the Ediacaran–Cambrian transition: behavioural diversification, ecological turnover and environmental shift. Palaeogeography, Palaeoclimatology, Palaeoecology 227: 323–356.

Smith, C. R., A. G. Glover, T. Treude, N. D. Higgs, and D. J. Amon. 2015. Whale–fall ecosystems: recent insights into ecology, paleoecology, and evolution. Annual Review of Marine Science 7:571–596.

Smith, V. A., J. Yu, T. V. Smulders, A. J. Hartemink, and E. D. Jarvis. 2006. Computational inference of neural information flow networks. PLoS Computational Biology 2:e161.

Sobel, J. M., G. F. Chen, L. R. Watt, and D. W. Schemske. 2010. The biology of speciation. Evolution 64:295–315.

Sperling, E. A., K. J. Peterson, and M. Laflamme. 2011. Rangeomorphs, *Thectardis* (*Porifera*?) and dissolved organic carbon in the Ediacaran oceans. Geobiology 9:24–33.

Townsend, C. R., M. Begon, and J. L. Harper. 2003. Essentials of ecology, 2nd ed. Blackwell Scientific, Oxford, U.K.

Velázquez, E., M. Kazmierczak, and T. Wiegand. 2016. Spatial patterns of sapling mortality in a moist tropical forest: consistency with total density dependent effects. Oikos 125:872–882.

Waggoner, B. 2003. The Ediacaran biotas in space and time. Integrative and Comparative Biology 43:104–113.

Wang, X., et al. 2011. Spatial patterns of tree species richness in two temperate forests. Journal of Ecology 99:1382–1393.

Wiegand, T., and K. A. Moloney. 2004. Rings, circles, and null-models for point pattern analysis in ecology. Oikos 104:209–229.

——. 2013. Handbook of spatial point-pattern analysis in ecology. CRC, Boca Raton, Fla.

Wiegand, T., W. D. Kissling, P. A. Cipriotti, and M. R. Aguiar. 2006. Extending point pattern analysis for objects of finite size and irregular shape. Journal of Ecology 94:825–837.

Wiegand, T., S. Gunatilleke, and N. Gunatilleke. 2007a. Species associations in a heterogeneous Sri Lankan dipterocarp forest. American Naturalist 170:E77–E95.

Wiegand, T., S. Gunatilleke, N. Gunatilleke, and T. Okuda. 2007b. Analyzing the spatial structure of a Sri Lankan tree species with multiple scales of clustering. Ecology 88:3088–3102.

Wiegand, T., I. Martínez, and A. Huth. 2009. Recruitment in tropical tree species: revealing complex spatial patterns. American Naturalist 174:E106–E140.

Wiegand, T., A. Huth, S. Getzin, X. Wang, Z. Hao, C. V. S. Gunatilleke, and I. N. Gunatilleke. 2012). Testing the independent species' arrangement assertion made by theories of stochastic geometry of biodiversity. Proceedings of the Royal Society of London B 279. 10.1098/rspb.2012.0376.

Wilby, P. R., J. N. Carney, and M. P.A. Howe. 2011. A rich Ediacaran assemblage from eastern Avalonia: evidence of early widespread diversity in the deep ocean. Geology 39:655–658.

Willis, R. J. 2007. The history of allelopathy. Springer Science & Business Media, Dordrecht, Netherlands.

Wood, D. A., R. W. Dalrymple, G. M. Narbonne, J. G. Gehling, and M. E. Clapham. 2003. Paleoenvironmental analysis of the late Neoproterozoic Mistaken Point and Trepassey formations, south-eastern Newfoundland. Canadian Journal of Earth Sciences 40:1375–1391.

Yu, J., V. A. Smith, P. P. Wang, A. J. Hartemink, and E. D. Jarvis. 2004. Advances to Bayesian network inference for generating causal networks from observational biological data. Bioinformatics 20:3594–3603.