

Integrated records of environmental change and evolution challenge the Cambrian Explosion

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The ‘Cambrian Explosion’ describes the rapid increase in animal diversity and abundance, as manifest in the fossil record, between ~540 and 520 million years ago (Ma). This event, however, is nested within a far more ancient record of microfossils extending at least into the late Ediacaran at ~571 Ma. The evolutionary events documented during the Ediacaran–Cambrian interval coincide with geochemical evidence for the modernisation of Earth’s biogeochemical cycles. Holistic integration of fossil and geochemical records leads us to challenge the notion that the Ediacaran and Cambrian worlds were markedly distinct, and places biotic and environmental change within a longer-term narrative. We propose that the evolution of metazoans may have been facilitated by a series of dynamic and global changes in redox conditions and nutrient supply, which, potentially together with biotic feedbacks, enabled turnover events that sustained multiple phases of radiation. We argue that early metazoan diversification should be recast as a series of successive, transitional radiations that extended from the late Ediacaran and continued through the early Palaeozoic. We conclude that while the Cambrian Explosion represents a radiation of crown-group bilaterians, it was simply one phase amongst several metazoan radiations, some older and some younger.

The Cambrian Explosion, starting at ~540 Ma, is considered to mark the abrupt appearance in the fossil record of abundant and diverse metazoans. This was accompanied by an increase in complexity of morphologies and behaviours, an increase in morphological disparity, the appearance of the first representatives of most extant phyla, and the rise of metazoan-dominated marine ecosystems¹. Although most works acknowledge that metazoans originated considerably before the Cambrian^{2,3} (though see ref. ⁴), the ‘roots’ of the Cambrian Explosion are greatly contested. The time at which this event was initiated is debated; it may have been around the Ediacaran–Cambrian boundary⁵, in the terminal Ediacaran⁶, or even deeper at either the appearance of the Nama Assemblage (~550–541 Ma⁷) or the Avalon–White Sea assemblages boundary at ~561 Ma^{3,7,8}, or, on the basis of molecular phylogenies, in the Tonian or Cryogenian at ~720 Ma¹. Furthermore, independent faunal turnover events and metazoan radiations are recognized in the subsequent early Palaeozoic, and so the relationship of these to evolutionary and ecological innovations across the Ediacaran–Cambrian boundary must be understood^{9–11}.

Whilst knowledge of individual aspects of these evolutionary developments is advancing, our broader understanding of early animal evolution is often hampered by highly compartmentalised, rather than holistic, study. Insights from diverse palaeobiological records have only recently been integrated with high-resolution geochemical studies and models, which have revealed much about the operation of the Earth system during this interval, such as the interaction between ecosystem engineers and oceanic biogeochemistry^{12–16}. Global stratigraphic correlation between key sections, aided by accurate and precise radiometric dating, is a continuous work in progress. Yet, holistic integration of datasets across the broader Ediacaran–Cambrian interval may enable us to address profound uncertainties, such as how seemingly different biotas

might be related^{8,17}; whether one or more mass extinctions occurred during this interval⁷; the evolutionary response, if any, to changes in oceanic redox conditions and nutrient availability^{15,16,18–20}; and the resolution of conflicting evidence for the origin of major metazoan clades from molecular clock, biomarker and palaeontological data^{2,4}. Without comprehensive geobiological and temporal integration, we risk missing the bigger, more significant evolutionary picture.

We focus here on integrating the tractable fossil and environmental proxy records of the Ediacaran–Cambrian interval to document the diversification of animals and their behaviour. This integration reveals a record of interactions between environmental change and biological evolution, culminating in the establishment of the crown-group metazoan phyla. Our compilation enables a re-evaluation of the record and explores the potential drivers of early metazoan evolution.

Environmental change versus evolutionary innovation

The late Cryogenian–Cambrian interval shows evidence for dramatic changes in the carbon cycle (Fig. 1b) and ocean redox conditions (Fig. 1c and Box 1). These geochemical changes, potentially driven by tectonic readjustment²¹, coincide with a series of major biotic innovations (Fig. 1a), including the appearance of metazoan motility by ~565 Ma, biomineralisation by ~550 Ma and bilaterian crown groups and predators by ~535 Ma (Box 2). A causal relationship between these records has long been proposed²². In particular, there is a broad consensus that dissolved oxygen provision reached a threshold, or series of thresholds, during the Neoproterozoic, which allowed the diversification of metazoans and their increasing metabolic demands^{23–25}. However, there remains considerable debate as to whether oxygenation was the main driver of early metazoan evolution after this initial physiological requirement was met^{18,23,26}. Indeed, the relationship between oxygen availability and

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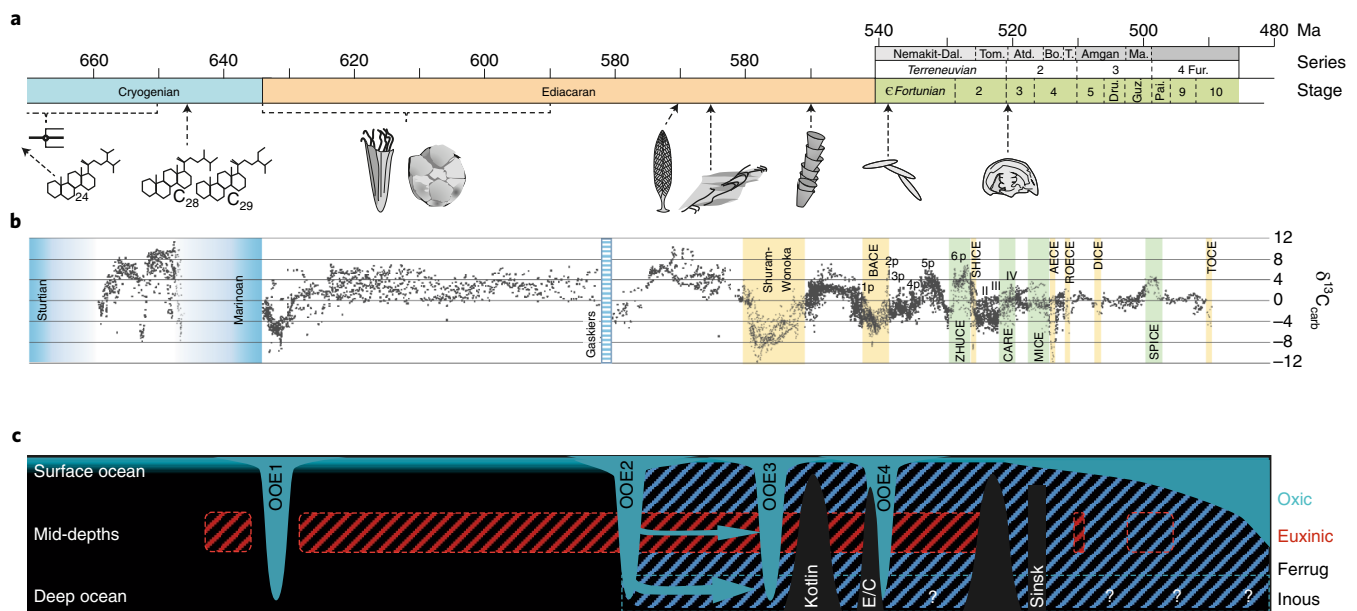


Fig. 1 | Integrated geochemical and biotic record between 670 and 480 million years ago. **a**, The time of the first appearances of major evolutionary milestones: 713–635 Ma, range of demosponge biomarker (24-isopropylcholestane)⁹²; 659–645 Ma⁹³, minimum age for marine planktonic algae (Archaeplastida); 635–590 Ma, possible stem-group cnidarian *Lantianella laevis*⁹⁴, phosphatised proposed animal embryos⁹⁵; >570.95 Ma, juvenile *Charnia masoni*⁹⁶; ~565 Ma, candidate for the earliest surface locomotion trace fossil⁹⁷; ~550 Ma, earliest skeletal animals, *Cloudina*⁹⁸ and earliest possible evidence for predation (borings in *Cloudina*⁹⁹); first appearance datum of the trace fossil *Treptichnus pedum*¹⁰⁰; earliest trilobite *Profallotaspis jakutensis*. **c**, Cambrian Period; Nemakit-Dal., Nemakit-Daldynian stage; Tom., Tommotian stage; Atd., Atdabanian stage; Bo., Botoman stage; T., Toyonian stage; Ma., Mayan stage; Fur., Furongian Epoch; Dru., Drumian Stage; Guz., Guzhangian Stage; Pai., Paibian Stage. **b**, Ediacaran C-isotope ($\delta^{13}\text{C}_{\text{carb}}$) compilation modified after refs. ^{101,102} and the references therein. Cambrian C-isotope profile conforms to composite curve of ref. ¹⁰³ (full details and data provided in Supplementary Information). Carbon isotope excursions are colour coded according to directionality (yellow, negative, green: positive) and abbreviations are as follows: ZHUCE, Zhujiqing; CARE, Cambrian arthropod radiation; SHICE, Shiyantou; MICE, Mingxinsi; AECE, Archaeocyath Extinction; ROECE, Redlichiiid-Olenellid Extinction; DICE, Drumian; TOCE, Top of Cambrian. Excursions 1p–6p and II to III correspond to carbon isotope peaks of Mongolia and Siberia. **c**, Schematic evolution of redox conditions based on compiled iron speciation data^{27,30,104} and proposed widespread anoxic intervals and 'oceanic oxygenation events'^{34,37,40,41,105}. Diagonal stripes indicate known regional differences in redox state. Paucity of truly basinal shales prevents determination of the redox state of the global oceanic deep basin (indicated by white question marks). E/C, Ediacaran–Cambrian; OOE, oceanic oxygenation event.

biotic response was likely to have been complicated by the operation of ecological and genetic factors, as well as poorly understood feedbacks between life and the broader Earth system.

Different geochemical proxies yield information with varying levels of spatial and temporal resolution. Local marine palaeo-redox reconstruction via iron speciation and rare earth element (REE) profiles in several key stratigraphic sequences indicates that open marine conditions in the Cryogenian to early Cambrian were typified by redox stratification^{14,23}. During this period, most, but not all, sampled basins record a shallow and highly dynamic chemocline above deeper ferruginous waters. However, redox proxy data are unavailable for many important successions, most notably in Morocco, Spain, Australia, India and Ukraine. In addition, the targeting of fine-grained facies has led to considerable bias in existing data²⁷, and the paucity of deep-water successions for key intervals during the Ediacaran has prevented unequivocal distinction between global layer-cake redox stratification of the oceans and highly dynamic oxygen minimum zones (OMZs) overlying potentially oxic basinal waters. There is growing evidence for the existence of OMZs in early Cambrian basins^{28,29,30}.

Many global proxies allow only for estimates of expanded seafloor anoxia and do not differentiate between deep and shallow marine settings. Consequently, it is not clear if such expansions restricted the habitable area of the shallow shelf, where most biodiversity resides. At least some biotas throughout this interval were subject to upwelling incursions of oxygen-deficient water that were controlled by local changes in relative sea level and productivity²⁸.

Therefore, throughout the entire Ediacaran–Cambrian radiation, and indeed beyond, oxygen levels may have been relatively low, but highly dynamic, fluctuating redox conditions were prevalent on local (that is, ecological), as well as global and evolutionary, temporal and spatial scales¹⁹.

The role of redox instability in biotic response. While all extant metazoans need oxygen, their demands are not equal. Modern low-oxygen regions are heterogeneous and dynamic habitats that support low-diversity communities of opportunistic and non-skeletal metazoans, many of which are meiofaunal; large, skeletal and motile metazoans, which form complex, biodiverse ecosystems, typically require higher oxygen levels²³. Experimental work has demonstrated that certain early-diverging clades (poriferans and ctenophores) may have very low oxygen demands since they lack hypoxia-inducible factor (HIF) pathways that maintain cellular oxygen homeostasis^{31,32}. This suggests that stem-group metazoans, and the metazoan Last Common Ancestor, may also have lacked the HIF pathway and so could have aerobically metabolized under very low concentrations of oxygen in their environments³². Animals most likely originated in a non-uniformitarian world with a low level of atmospheric oxygen, almost certainly before the permanent oxygenation of the deep ocean. However, it remains unclear whether animal diversification and increased ecosystem complexity were driven extrinsically by the expansion of permissive oxic niches or by genetic or developmental innovations that enabled animals to expand into the oxic realm.

Box 1 | Oxygen and biogeochemical cycles during the Ediacaran–Cambrian

The non-uniformitarian nature of the oceans across the Ediacaran–Cambrian interval, continuing into the Ordovician, is evidenced by considerable instability in the carbon isotope record in inorganic carbonates (Fig. 1b). The magnitude of the largest known negative carbonate carbon isotope ($\delta^{13}\text{C}_{\text{carb}}$) excursion in the geological record, the Ediacaran Shuram/Wonoka anomaly, has a nadir of -12% (ref. 22). Following recovery to positive values before 551–548 Ma, $\delta^{13}\text{C}_{\text{carb}}$ remained relatively unperturbed for the final ~ 10 million years of the Ediacaran (that is, the EPIP), before falling again to values indicative of the BACE^{103,110,111}. The BACE pre-dates the first appearance of *Treptichnus pedum*^{91,112}, while in South China it correlates with the *Asteridium–Heliosphaeridium–Comasphaeridium* (AHC) acritarch assemblage and the *Anabarites trisulcatus–Protohertzina anabarica* (small shelly fossils) Assemblage Zone^{113,114}. A number of additional short-lived $\delta^{13}\text{C}_{\text{carb}}$ excursions continued to punctuate the Cambrian^{43,103} and Ordovician chemostratigraphic records¹¹⁵, which exhibit a progressive decline in overall excursion magnitude into the Phanerozoic¹¹⁶.

Proposed explanations for $\delta^{13}\text{C}_{\text{carb}}$ anomalies throughout this interval are variable and remain contentious^{22,117}, but they may, at least in part, be related to dynamic change in redox and nutrient cycling. While selenium isotopes suggest that there was a generally progressive oxygenation through the Neoproterozoic¹¹⁸, compiled data show that oceanic redox conditions oscillated dramatically over million-year timescales before a permanent and stable oxygenated state was achieved³⁷. The exact timing of these events and confirmation of their global extent, as well as the drivers for such rapid and global changes, remain unclear. Molybdenum and uranium isotopes show that an increase in oxygenation was punctuated by intervals of expanded anoxic seafloor^{34,40,119}; low thorium/uranium ratios show a positive correlation with $\delta^{13}\text{C}$ values at multiple sites across the Ediacaran–Cambrian boundary, suggesting that the BACE was a response to the widespread development of shallow marine anoxia^{120,121}. A shift in the marine sulphur cycle, as recorded by $\delta^{34}\text{S}$, is possibly consistent with increasingly widespread sulphate reduction under anoxic conditions around ~ 550 Ma^{22,122}.

Others have argued for intervening late Ediacaran ‘oceanic oxygenation events’ (OOEs) (Fig. 1c) at around 575 Ma (OOE2), ~ 560 Ma (OOE3) — possibly coincident with the start of the Shuram — and at ~ 540 Ma (OOE4)^{37,105}, although differing redox proxies are not always consistent with the timing of these intervals. For example, on the basis of nitrogen isotopes, oxic intervals have been proposed ~ 551 – 543 Ma, ~ 544 – 529 Ma, and ~ 521 – 517 Ma³⁵. Diverse proxies suggest that the global ocean became progressively more oxygenated through the early Cambrian until ~ 520 Ma, after which time there was a return to more widespread anoxia^{36,41}. The Sinsk event, accompanied by a negative carbon isotope excursion (CIE), is another short-lived anoxic interval at ~ 513 Ma¹¹. Independent proxies suggest that full oxygenation of deep ocean was not reached until the Devonian^{27,106}.

When palaeoredox proxy data are integrated with biotic distribution, metazoans are usually restricted to localised oxygenated habitats, either above a shallow chemocline or potentially below an OMZ^{29,33}. The instability of the chemocline would therefore have provided strong anactualistic controls on the distribution of metazoans and potentially on the taphonomic windows for their preservation. Ediacaran and Cambrian sediments deposited below the chemocline (or within an OMZ) may therefore be expected to lack significant metazoan life assemblages.

Box 2 | The metazoan fossil record between ~ 571 – 520 Ma

The oldest macrofossils of the so-called ‘Ediacaran macrobiota’ are known from the Drook Formation, Newfoundland¹⁰⁷ and are dated at ~ 571 Ma³¹. Although their precise phylogenetic position remains unclear, at least some taxa are reasonably interpreted to have been total group metazoans¹⁰⁸. The wider macrobiota includes a range of complex organisms with tubular, frondose, modular and sheet-like morphologies that likely include multiple clades¹²³. These have been grouped within three proposed biotic ‘assemblages’ (recurrent community compositions) that appear to be predominately controlled by facies rather than age⁵⁵: the Avalon (~ 571 – 555 Ma), White Sea (~ 560 – 551 Ma) and Nama (~ 555 – 541 Ma) assemblages^{76,90}. Frondose taxa (for example, *Charnia* and *Arborea*) dominate the Avalon Assemblage and are accompanied by rare candidate cnidarians¹⁰⁹, but both the diversity and taxonomic disparity of macroscopic soft-bodied organisms increase significantly after ~ 560 Ma^{3,124}. The first identifiable motile, heterotrophic organisms are *Kimberella quadrata*, *Yorgia* and *Dickinsonia* from Russia at ~ 558 – 555 Ma; the latter is highly likely to have been a metazoan on the basis of ichnological, developmental and biomarker evidence^{108,125}. A widespread record of surface locomotory trace fossils exists from at least 560 Ma^{54,97}, and these diversify in form and complexity through the late Ediacaran^{47,48,126,127}. Latest Ediacaran assemblages show a marked reduction in soft-bodied macrofossil diversity, but also exhibit the appearance of organic-walled (for example, *Corumbella*) and skeletal tubular fossils, and a diversification of bilaterian trace fossils^{9,54}. The oldest skeletal macrofossil, *Cloudina*, which also shows the first possible evidence for predatory borings⁹⁹, is known globally from ~ 550 Ma⁹⁸. *Cloudina* could form reefs¹²⁸ (but also see ref. 129) and together with the late Ediacaran (~ 543 Ma) *Paraconularia*, is plausibly interpreted as a total-group cnidarian (and, therefore, a crown-eumetazoan).

The Ediacaran–Cambrian boundary is defined by the first appearance of the complex trace fossil *Treptichnus pedum*¹⁰⁰, presently dated to 541 Ma based on the inferred correlation of successions from Newfoundland, Namibia and Oman. The earliest Cambrian (Fortunian) fossil record shows a marked increase in ichnofossil abundance, size and complexity in shallow marine environments⁹. The first probable crown-group molluscs and brachiopods appear in the late Fortunian or early Stage 2 (ref. 42). There is also a notable rise of bilaterian predators¹³⁰. The early to middle Cambrian hosts a variety of lagerstätten that document crown-group representatives of disparate skeletal and non-biominalising animal phyla¹¹³, as well as increases in body size across many animal clades, skeletonisation, and the expansion of ecological networks¹.

These patterns in the fossil record are yet to be converted into a coherent understanding of the dynamics of how metazoan phyla appeared and evolved⁵, but quantitative analysis of lophotrochozoan skeletal species from the terminal Ediacaran to Cambrian Stage 5 (~ 545 – 505 Ma) on the Siberian Platform show a disjunct temporal distribution that suggests that the radiation of bilaterian metazoans occurred in two phases, separated by an extinction event. The first was dominated by lophophorate, brachiopod and mollusc stem groups from ~ 542 – 513 Ma, and the second was marked by radiating brachiopod and mollusc crown-group species from ~ 513 Ma, extending to the GOBE⁴².

At least regionally, stable ocean oxygenation following the Gaskiers glaciation broadly coincides with the appearance of soft-bodied Ediacaran macroscopic biota in Newfoundland. Extensive ocean oxygenation is argued to have happened together with the

end of the Shuram event, ca. 560–551 Ma (review ref. ³⁰). This event is broadly coincident with the first evidence for probable motile, heterotrophic and muscular bilaterians, although dating and global correlation are not well constrained, and integrated, local datasets are not available for establishment of cause and effect³⁰. It is also not clear whether the rise of mobile bilaterians and predators required additional ecological triggers²³. Records from the open oceanic Laurentian margin (632–540 Ma), the Nama Group (~550–538 Ma) and the variably restricted Yangtze Block (635–520 Ma) all show continued redox instability after the first fossil evidence for (probable) metazoans (review ref. ³⁰). The first skeletal metazoans appear at ~550 Ma, under local conditions of continued redox stratification³³ and before an interval of expanded anoxia³⁴. However, in the Nama Basin, integrated iron speciation and cerium anomaly data show that in-situ Ediacaran skeletal metazoans did not occupy low-oxygen waters¹⁴.

These dynamic carbon and redox records are also closely tied to probable changes in nutrient cycling, but mechanistic details are far from clear^{15,20,35,36}. For example, step changes in the burial of phosphorus and organic carbon, driven by evolutionary innovations, have been argued to have progressively lowered marine phosphate concentrations¹⁵. Each step change would result in a pulse of marine oxygenation, but, over long timescales, the decrease in organic carbon/phosphorus burial ratios would have driven atmospheric oxygen levels down and slowly deoxygenated the oceans¹⁵.

Others have noted that the low total organic carbon content in Neoproterozoic shales suggests that the ocean at this time may have had a lower flux of primary productivity to the seafloor than in the Phanerozoic²⁰. An expected consequence of this nutrient limitation is that biota would have small body sizes and sparse, heterogeneous global distributions²⁰. Analysis of nitrogen isotopes over the late Ediacaran–early Cambrian interval supports the hypothesis that pulses of oxygenation, punctuated by regional anoxic events of shoaling chemoclines, were closely associated with an increase in nutrients that boosted the primary productivity of large-celled, eukaryotic phytoplankton, perhaps in turn stimulating metazoan evolution³⁶.

Individual marine basins continued to record unstable redox conditions over short timescales well into the Cambrian, with dominantly ferruginous and even euxinic conditions, particularly in shelf and slope environments^{29,33,37,30}. Adjacent basins can show different redox histories, and the oxidic chemocline often shoaled during sea-level transgressions^{33,30}. While some studies propose regional deepening of the oxycline during Cambrian Stage 2, with an accompanying increase in biological diversity^{38,39}, others have shown that anoxic or low-oxygen concentrations may have prevailed for extended intervals that contain notably biodiverse faunas, such as those recorded by the Chengjiang biota²⁹. However, just as we know little about the spatial scales of Ediacaran–Cambrian redox heterogeneity, we also know little quantitatively about the lateral distribution and patchiness of benthic biotas.

In a world with lower atmospheric oxygen than today, the habitability of benthic marine environments may have been governed by oxygen demand, dependent on rates of primary production, rates of oxygen replenishment via atmospheric mixing and diffusion, and the sinking rates of dominant primary producers²⁹. The distribution of shallow marine habitable zones would therefore have evolved dynamically as a result of changes in nutrient availability as a consequence of changes in circulation, upwelling and sea level.

The processes driving biotic response to rising atmospheric oxygen levels, which may have manifest as a deepening of the chemocline, an increase in local redox stability or a reduction of global redox heterogeneity, are not clear. Increasing oxygen levels have been proposed to result in an increase in overall biodiversity, the rise of new traits that are more metabolically demanding, such as motility and skeletisation, or the formation of more complex food webs

and ecosystems²³. Increasing areal occupation of seafloor, or changes in biogeography as habitable settings expanded and connected, may also be expected. Globally expansive anoxia is locally expressed as heterogeneous, poorly ventilated basins, but whether habitable shallow shelf space was reduced or fragmented during past anoxic intervals is unknown. This emphasizes the need to integrate global and local redox proxies. Notably, integrated geochemical and biotic data show that diverse Cambrian communities were established under dynamic redox conditions in oxidic refugia²⁹, essentially under similar conditions to terminal Ediacaran ecosystems with far lower diversity. Hence, the relationship between the evolution of metazoan ecosystems and increasing atmospheric oxygen and redox stability appears far more complex than direct, linear cause-and-effect.

It is possible that fluctuations in redox may, in fact, reinforce rather than hinder evolutionary transitions, with variability in near-surface oceanic oxygenation promoting morphological evolution and novelty¹⁹. Several major radiations, including that across the Ediacaran–Cambrian boundary, the great Ordovician biodiversification event (GOBE) and that in the mid-late Triassic, follow intervals of protracted or dynamic shallow marine anoxia. These dynamic conditions may have created opportunities for the generation of evolutionary novelty in soft-bodied benthos, which then provided ancestral stock for subsequent skeletonised lineages once oxidic conditions became widespread, connected and stable¹⁹.

The period of globally expanded anoxia at ~551 Ma has been proposed to coincide with a reduction in diversity of the Ediacaran macrobiota as manifest in the depauperate Nama Assemblage⁴⁰. But data from the Nama Group, Namibia, show that the transition toward globally widespread anoxic conditions postdates the first appearance of both the skeletal and soft-bodied fauna of the Nama Assemblage, demonstrating that this expansion did not coincide with the decline of the Ediacaran biota³⁴. This expansion may rather reflect a geochemical response to an ecological innovation or change^{15,34}. We note that the diversification of ichnofossils in Namibia and the appearance of organic-walled tubular taxa broadly coincide with the subsequent reduction of global anoxic seafloor conditions.

There is evidence to suggest that new lineages of crown-group bilaterians appear after anoxia at the Ediacaran–Cambrian boundary at ~540 Ma and again after an interval of anoxia at ~520 Ma⁴¹. Following this, we see two phases of radiation separated by the Sinsk Event extinction⁴². The first is dominated by non-bilaterian and bilaterian stem groups from ~542–513 Ma, and the second is marked by radiating non-bilaterian and bilaterian crown-group species from ~513 Ma through the Ordovician radiation. This second radiation may have been interrupted by the late Cambrian Steptoean positive carbon isotope excursion (SPICE) anoxic event⁴³, coincident with a negative carbon isotope excursion, which marked a further minor extinction, but also ushered in the GOBE (Fig. 1). We thus postulate that waves of metazoan innovation immediately followed intervals of dynamic redox conditions (following the model of ref. ¹⁹) throughout the Ediacaran to Ordovician periods.

Understanding redox and ecology at the local scale. The apparent drop in biodiversity between the comparatively diverse White Sea Assemblage and the Nama Assemblage and the disappearance of Ediacaran soft-bodied macrobiota at the end of the Ediacaran have both been suggested to have been mediated by increasing competition and predation following the rise of bilaterian or crown-group animals. The rise of bioturbation, predation, biomineralisation and grazing by bilaterians may also have perturbed sediment stability and reduced the availability of dissolved organic carbon (DOC)—this is the so-called ‘biotic replacement’ model, which involves ecosystem engineering^{44–46}.

Multiple bedding surfaces in late Ediacaran successions from Australia, China and Russia (Fig. 2b,g) demonstrate the co-occurrence of prominent horizontal burrows, such as *Lamonte* and

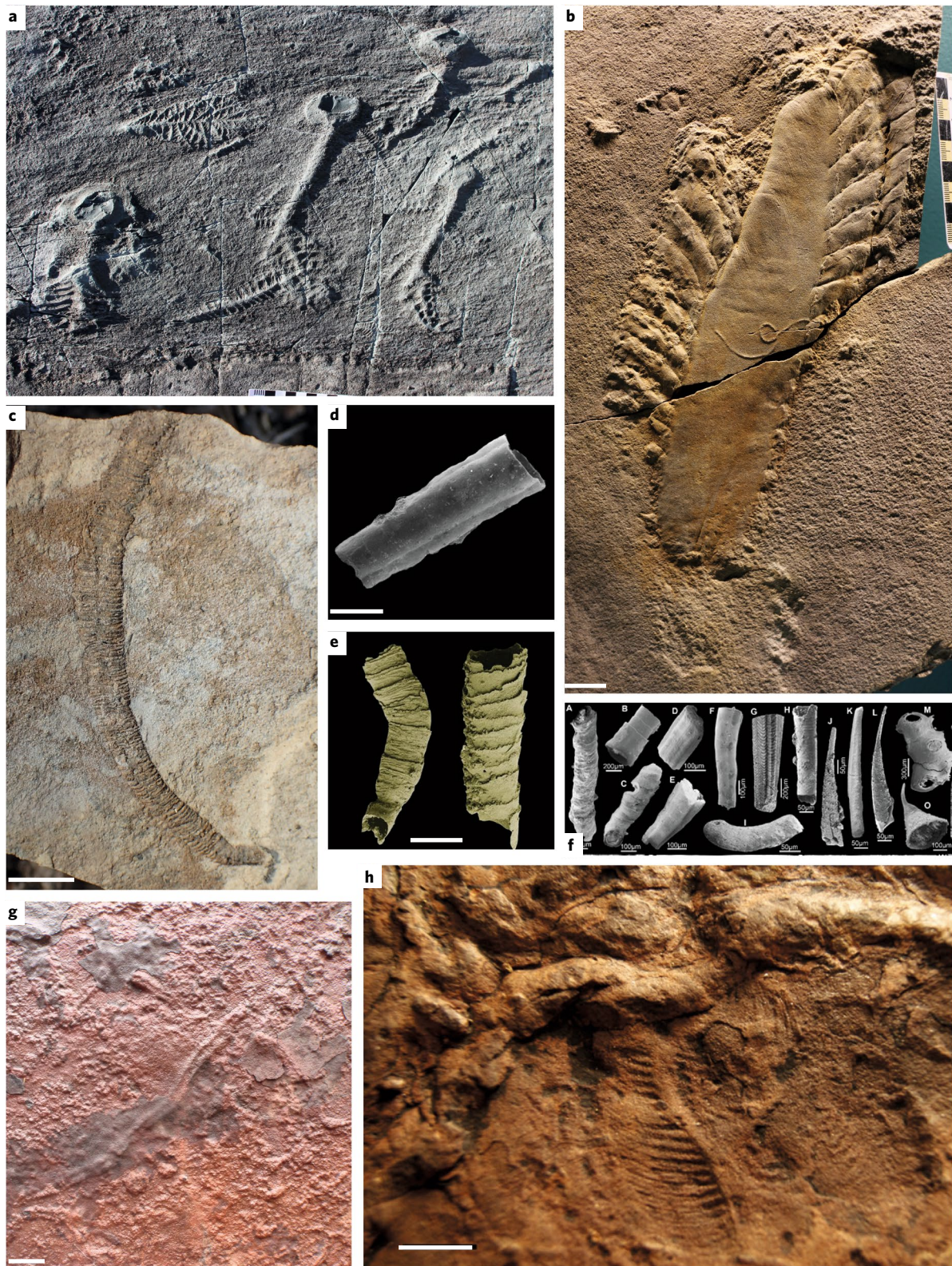


Fig. 2 | Key transitional Ediacaran and Cambrian taxa. **a**, Representative taxa of the Avalon biota of Newfoundland (-566 Ma) almost uniquely comprised of soft-bodied, frondose members of the Ediacaran biota. **b**, Ediacaran *Arborea* with associated trace fossil; Flinders Ranges, South Australia. South Australia Museum (SAM) specimen SAM P49393. **c**, *Corumbella*, organic-walled tubular fossil; latest Ediacaran Corumbá Group, Brazil. **d**, *Anabarites trisulcatus*; Ust'-Yudoma Formation, Kyra-Ytyga River, Siberia. **e**, *Cloudina*, South China. **f**, Transitional small shelly fossil biota; Siberia (adapted from ref. ⁶). **g**, Bilobed trace fossils from the Ediacaran Dengying Formation; Wuhe, South China, occurring on beds adjacent to surfaces bearing non-mineralized tubular organisms (for example, *Wutubus*) and soft-bodied Ediacaran macrobiota (for example, *Pteridinium* and *Charniodiscus*). **h**, Cambrian *Swartpuntia*-like moulds alongside trace-fossils, including *Treptichnus*; Uratanna Formation, Flinders Ranges, South Australia. SAM P36399/36403. Scale bars, 10 mm (**b**), 5 mm (**c**), 300 μm (**d**), 1 mm (**e**), 20 mm (**g**), 10 mm (**h**). Credit: A. Fedorov (**d**); S. Xiao (**e**)

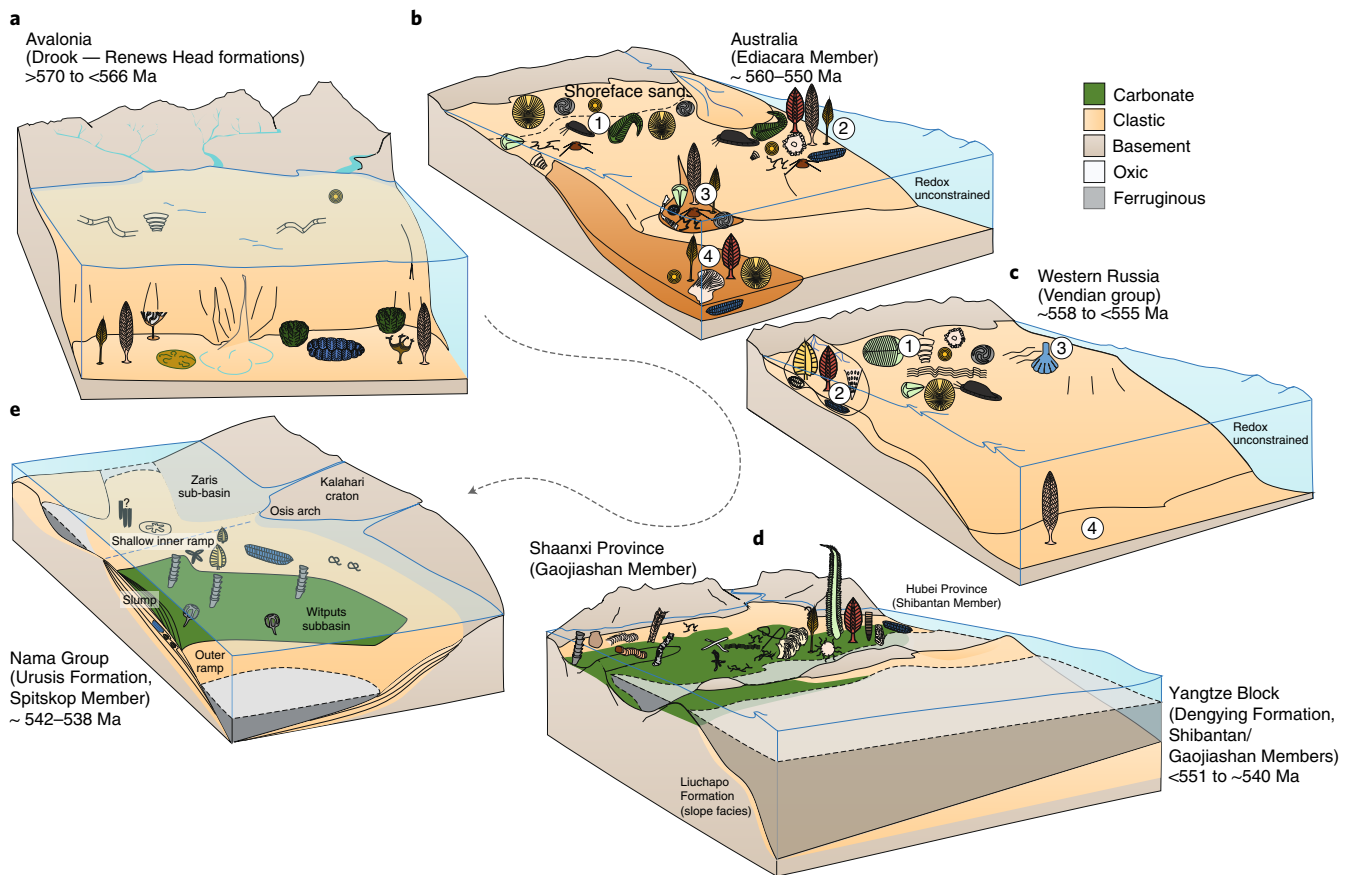


Fig. 3 | Ediacaran ecosystem dioramas. a–e, Single, conformable sequences from (a) Avalonia, (b) Australia, (c) Western Russia (White Sea Region), (d) the Yangtze Block, China, and (e) Namibia. Each shows the distribution of contemporary biota within the local sedimentological and redox setting and the relative water depth. The dashed grey arrow indicates relative ages of each palaeoenvironment, from oldest to youngest. See the Supplementary Information for the key to stylised biota and numbering.

Helminthoidichnites, with mobile soft-bodied taxa such as *Yorgia* and *Dickinsonia*, and tubular and frondose macro-organisms^{47–49}. Many such occurrences can be reasonably assumed to record contemporary communities. Such co-existence continues into the earliest Cambrian, where relatively large trace fossils, such as *Treptichnus*, are found alongside putative *Swartpuntia*-like impressions in the earliest Cambrian Stage 3 (Fig. 2h). We see no evidence of direct competitive replacement, but rather of probable sustained co-existence in both shallow marine carbonate and siliciclastic settings.

For biotic replacement to occur, taxa must be both spatially co-located and have similar resource requirements, yet spatial analyses of contemporary communities find only very limited instances of resource competition. Integrated sedimentological and redox models for key fossiliferous Ediacaran successions in Avalonia (Newfoundland), south Australia, western Russia, the Yangtze Block (South China) and Namibia reveal the diversity of settings occupied by early metazoans (Fig. 3).

The Drook to Renewes Head formations of eastern Newfoundland (~571–566 Ma) represent the Avalon Assemblage (Fig. 3a) and were deposited in deep marine environments⁵⁰. Benthic colonisation, mainly by sessile, frondose rangeomorphs, occurred after the appearance of stable oxic conditions^{51,52}. The biota from the Ediacara Member, South Australia (~560–550 Ma), represents the shallow marine White Sea Assemblage (Fig. 3b; based on data in ref. 53), which hosts rangeomorphs, soft-bodied motile taxa such as *Kimberella*⁵³ and *Dickinsonia* and trace fossils⁵⁴. The broadly coeval White Sea region of western Russia (Fig. 3c) shows distinct facies-based soft-bodied assemblages, with *Charnia* communities

in deeper settings and shallower, pro-deltaic White Sea and Nama-like Assemblages⁵⁵. Both assemblages suggest occupation of at least intermittently oxic settings, permissible for life habits potentially involving relatively high metabolic oxygen demands⁵⁶. The Dengying Formation, China (~551–541 Ma), shows persistent and long-lasting redox stratification, with deep ferruginous waters commonly encroaching onto the shallow platform¹². The highly fossiliferous shallow marine bituminous Shibantan Member (Fig. 3d) has soft-bodied frond-like taxa, tubular forms, vendotaenids and trace fossils as well as biota characteristic of the Avalon and Nama Assemblages^{47,48,57} and shows intermittent ventilation during storm events^{58,59}. By contrast, the time-equivalent mixed carbonate and siliciclastic deposits of the Gaojiashan Member of the Dengying Formation record a very different assemblage dominated by tubular skeletonising forms, including *Cloudina* and *Sinotubulites*, and non-biomineralised tubular *Gaojiashania*, *Conotubus* and *Shaanxilithes*^{60–63}. These communities likewise grew under intermittently well-ventilated conditions, but with incursions of anoxia⁶⁴. Finally, the mixed siliciclastic and carbonate deposits of the uppermost Nama Group, southern Namibia (Fig. 3e), were deposited in a predominantly offshore, storm-dominated shelf environment from ~542–540 Ma⁶⁵. These contain the complex trace fossils *Streptichnus narbonnei* in addition to soft-bodied macrofossils^{46,66}. Thinly bedded limestone units recording deposition during highstands host skeletal *Namacalathus* and *Cloudina*. Palaeoredox conditions are interpreted as dominantly stable and oxic^{33,46}.

These analyses uncover an increase in morphological and ecological complexity through time, critically with an increase in the

co-existence of taxa, particularly of trace-makers. Organic tubular and trace fossil taxa co-occur locally and indeed over millions of years (supporting refs. 4,8, but contra ref. 17). However, where soft-bodied and skeletal biotas co-existed, they predominantly occupied different parts of basins. For example, taxa such as *Cloudina* and *Namacalathus* occupied carbonate sedimentary settings, while pre-existing clades (such as rangeomorphs) were generally found in siliciclastic settings. This is noteworthy since, in mixed sedimentary successions, clastics dominate transgressive systems tracts, but carbonates dominate highstand systems tracts.

Changes in seawater chemistry and the rise of biomineralisation.

The global onset of widespread calcareous biomineralisation at ~550 Ma, probably in low latitudes with supersaturated waters with respect to calcium carbonate⁶⁷, may suggest the operation of an extrinsic trigger. Terminal Ediacaran to Cambrian putative metazoan skeletal taxa are morphologically and mineralogically diverse, indicative of independent acquisition of skeletons in diverse taxonomic groups⁶⁸. Many, including the first putative poriferans⁶⁹, are inferred to have had a precursor organic skeleton and apparently simple microstructures⁶⁷. Some skeletal taxa known from carbonate successions appear to have non-skeletal, organic-walled counterparts in siliciclastic facies, for example *Cloudina/Conotubus*, *Sinotubulites/Corumbella* and *Protolagena/Sicylagena*⁷⁰. Early metazoan skeletal clades commonly co-opted carbonate minerals in concert with ambient ocean chemistry; this was potentially driven by inferred changing seawater magnesium/calcium (Mg/Ca) ratios⁷¹. Fluid inclusion data, models and early marine cements all suggest that seawater Mg/Ca progressively lowered during the Ediacaran to early Cambrian^{72,73}. The first skeletal macrofossils coincide with the appearance of widespread early marine cements of either aragonite or calcite high in magnesium, implying that calcareous biomineralisation may have been facilitated by increased relative calcium concentrations⁷².

Distinction between the Ediacaran and Cambrian biotas

Arguments for mass extinction and ecological replacement across the Ediacaran to Cambrian transition have emphasized taxonomic and ecological differences between the Ediacaran and Cambrian biotas⁴⁵. For example, typical Ediacaran and Cambrian trace fossils are claimed to be distinct in size and complexity. While most soft-bodied Ediacaran taxa and some skeletal taxa are lost, the gaps and biases in the record and the absence of sufficient chronostratigraphic control preclude definitive statements about gradual or rapid rates of decline, but do nonetheless support phases of faunal turnover (Fig. 4).

Potential evidence for a mass extinction at the Ediacaran–Cambrian boundary requires establishing the presence of a severe and global environmental perturbation that is synchronous with a rapid reduction in biodiversity⁸. The basal Cambrian negative carbon isotope excursion (BACE) has been suggested to mark a major perturbation coincident with the mass extinction of soft-bodied macrobiota, but an absence of dateable beds has hampered the construction of correlation schemes of sufficiently high resolution to establish global synchronicity of this event (see the review sections of refs. 8,9,45). Likewise, frequent unconformities across the Ediacaran–Cambrian boundary at many important fossiliferous sections make this hypothesis difficult to test (Supplementary Fig. 1). A literal reading of the record shows that the main decrease in soft-bodied benthic biodiversity occurred ~5–10 million years before the Ediacaran–Cambrian boundary^{45,3}. Indeed, despite the abundance of lagerstätten, the current inventory does not provide a continuous record through the interval, and there is also a notable dearth of precisely dated assemblages at and around the Ediacaran–Cambrian boundary. Widespread development of shallow marine anoxia coincident with the BACE would suggest a potential agent for

mass extinction, but the mechanisms driving the BACE are poorly known⁸. We suggest that such an anoxic episode would appear to be just one of several similar, both preceding and succeeding, redox perturbations.

A case for successive, transitional assemblages. At a local scale, Ediacaran biotas can be highly variable and can differ dramatically within successive beds^{74,75}. This community heterogeneity is suggestive of high beta (between-community) diversity, indicating that the representativeness of palaeobiological information may be restricted. In contrast, global, long-term biotic patterns in diversity across this interval have been considered to be tractable and have statistical support^{8,76}.

Age ranges of key fossil genera, and the minimum ages implied by higher phylogenetic relationships, indicate temporal overlap between taxa typically perceived as Ediacaran or Cambrian (Fig. 4). First, examples of organisms at least superficially similar to the Ediacaran macro-organism *Swartpuntia* are known to have existed in the Cambrian period^{77,78}, and putative links have been made between frondose Cambrian forms and Ediacaran rangeomorphs^{79–81}. In addition, there are documented occurrences of complex treptichnid trace fossils in the late Ediacaran, some of which have been convincingly attributed to bilaterian priapulid worms⁷⁷.

Second, high-resolution carbon isotope chemostratigraphy and biostratigraphy for a terminal Ediacaran to Cambrian succession on the eastern Siberian Platform show the presence of a succession of diverse fossil assemblages before the start of the BACE⁶. Here, a mixed Ediacaran and Cambrian skeletal biota (*Cloudina*, *Anabarites* and *Cambrotubulus*) appears in limestones within the late Ediacaran positive carbon isotope plateau (EPIP; Fig. 2d,e). The co-occurrence of cloudinids with various other skeletal species, representing a number of diverse clades of early Cambrian aspect, has also been documented elsewhere in Siberia, South China and Kazakhstan^{6,82,83}. The agglutinated fossils *Platysolenites* and *Spirosolenites*, which occur globally in the Cambrian⁸⁴, also co-occur with Ediacaran *Cloudina*, *Vendotaenia* and *Namacalathus* on the Western Siberian Platform⁸⁵. There is, therefore, evidence for considerable diversification of characteristic Cambrian-type skeletal taxa before the BACE.

Third, trace fossils made by mobile burrowing bilaterians first appear in the latest Ediacaran and continue as identical traces into the Cambrian (for example, *Helminthoidichnites* and *Archaeonassa*). Although their trace makers are unknown and may have changed over time, the continuity of these traces suggests that at least some behaviours of soft-bodied denizens of the latest Ediacaran, and therefore potentially some of the higher-level taxonomic groups to which they belong, continued across the boundary^{4,86} (Fig. 4).

These integrated data show that taxa attributed to so-called Ediacaran and earliest Cambrian skeletal biotas overlap in some localities, without notable biotic turnover. In addition, there may be a close relationship between organic-walled and skeletal tubular taxa⁷⁰, further supporting a transitional assemblage acquiring skeletonisation in permissive settings. Extrapolation of radiometric dating from South China⁸⁷, the northern Siberian Platform⁸⁸ and Oman⁸⁹ constrains this transitional skeletal biota to ~545–540 Ma.

Rather than advocating for distinct Ediacaran and Cambrian biotas, we here make a case for a succession of taxonomically distinct biotas or assemblages (Fig. 4b–d), each of which is marked by the appearance of new biological traits and ecological strategies and which were, to a greater or lesser extent, governed by facies. First was the Avalon (~571–557 Ma), followed by the White Sea (~560–551 Ma) and Nama (~555–541 Ma) assemblages^{76,90}, with frond-dominated Avalonian assemblages later embellished by a 'second wave'³ of Ediacaran diversification in the White Sea Assemblage. The White Sea Assemblage documents increased diversity and taxonomic disparity of macroscopic soft-bodied organisms, including

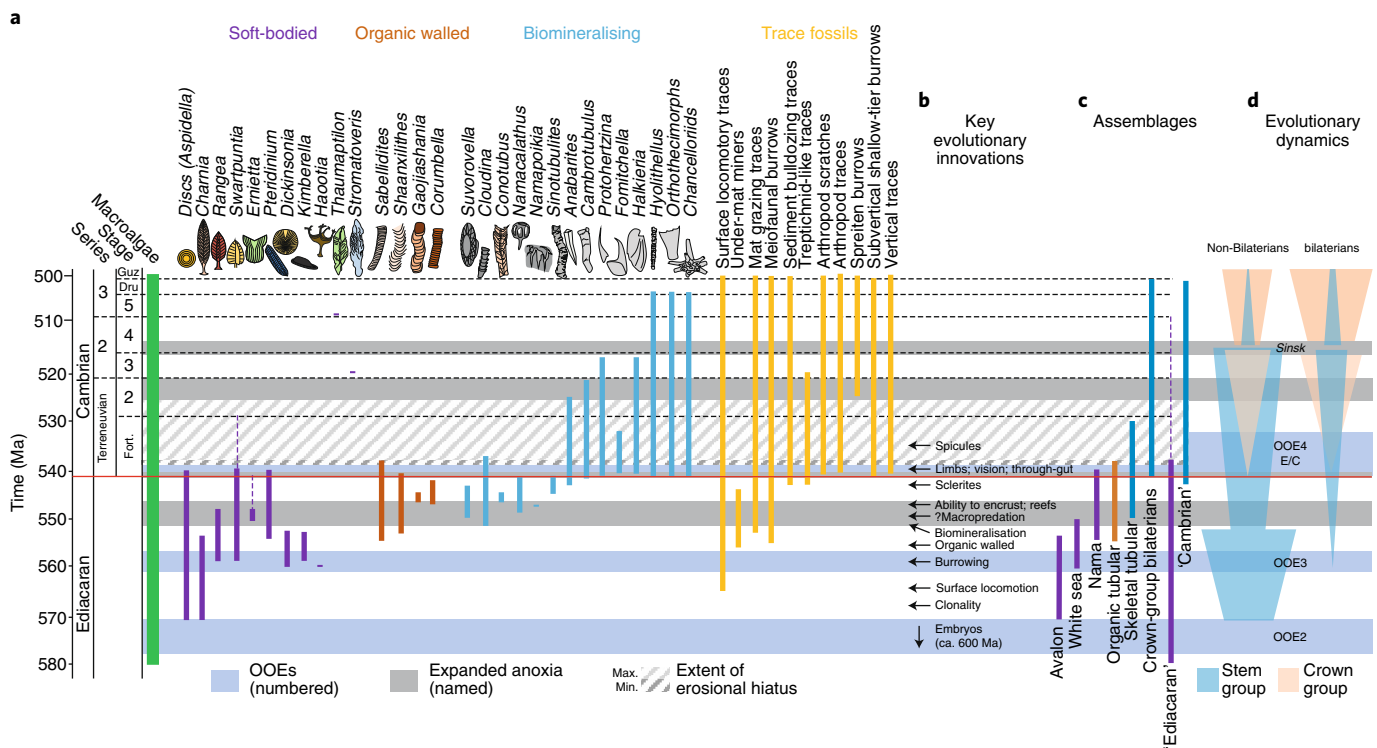


Fig. 4 | Biotic evolution across the Ediacaran–Cambrian. **a**, Temporal occurrence ranges for key soft-bodied, organic-walled, biomineralised, and trace fossil Ediacaran and transitional Cambrian taxa (references in Supplementary Information). Minimum and maximum duration of hiatus associated with the Ediacaran–Cambrian boundary is shown. **b**, Oldest fossil evidence for key evolutionary innovations. **c**, Distribution and succession of transitional assemblages. **d**, Schematic evolutionary dynamics showing the temporal distribution of stem and crown groups in non-bilaterians and bilaterians⁴², and major anoxic events and OOE^{54,106–109}. Full details are available in the Supplementary Information.

the first motile bilaterians and trace fossil makers (although we note the global dearth of shallow marine facies coeval with the Avalon Assemblage). By contrast, the latest Ediacaran successions from Namibia, Paraguay, Brazil, the United States, Siberia and China document lower-diversity assemblages of soft-bodied taxa, but two distinct new body plans also appear: organic-walled tubular taxa in shallow marine clastic settings and biomineralising tubular taxa in shallow marine carbonate environments that were previously largely unoccupied^{17,43,91}. After the Ediacaran–Cambrian boundary, there is a radiation of probable stem-group members of bilaterian phyla and non-bilaterians, and then a second radiation of bilaterian phyla inferred to be crown-group members, which continued to diversify in the GOBE⁴².

As noted in the distribution of Phanerozoic evolutionary faunas (EF; Fig. 4), the boundaries between these assemblages are not defined by complete replacement of one EF by the next, but rather by the rise to ecological dominance of groups whose origins predate that rise⁸.

Conclusions

The Ediacaran–Cambrian palaeontological and geochemical records reveal a progressive addition of biological novelty of form and process, and complexity within the Metazoa. Highly heterogeneous and fluctuating redox conditions throughout the late Ediacaran–early Palaeozoic interval, with successive but temporary expansions of oxic seafloor and possibly changing availability of phosphorous and nitrogen, facilitated the transition from low-oxygen Proterozoic oceans to more extensively oxygenated Phanerozoic oceans and the rise of modern biogeochemical cycles. This geochemical instability may have driven pulses of evolutionary innovation, but biotic feedbacks are poorly understood.

Ecological and evolutionary responses to this instability could have wide-reaching implications for discussions of gradualistic versus punctuated evolution.

We argue that the record can be considered to be a succession of assemblages, with the establishment of Cambrian crown-group animal ecosystems built on several successive Ediacaran advances as well as environmental and biotic feedbacks. The oldest record of Ediacara-type macrofossils appears to be dominated by probable non-bilaterian metazoans, with bilaterian metazoans appearing by ~560 Ma. A reduction in diversity occurs at ~551 Ma and is closely followed by the appearance of the first biomineralised taxa, but a well-documented expansion of seafloor anoxia postdates these events. Bilaterians, including predators, diversify after an episode of widespread anoxia at the Ediacaran–Cambrian boundary, immediately succeeded by an inferred ‘oceanic oxygenation event’ at ~540 Ma. Inferred stem-group poriferans, molluscs and brachiopods were seemingly devastated by the early Cambrian Sinsk anoxic event (~513 Ma), in contrast to inferred crown-group bilaterian phyla, whose diversification continues through to the GOBE.

There is currently no compelling evidence for either significant competitive replacement or biotic replacement from the latest Ediacaran to Cambrian. Indeed, we conclude that a discrete Cambrian Explosion event is difficult to temporally isolate, or indeed to define. The rise of early metazoans can be more simply and holistically recast as a series of successive, transitional radiation events, perhaps mediated via complex environmental change, which extended from the Ediacaran and continued to the early Palaeozoic.

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References

- Erwin, D.H. & Valentine, J.W. *The Cambrian Explosion: The Construction of Animal Biodiversity* (Roberts and Company Publishers Inc., Greenwood Village, CO, USA 2013).
- dos Reis, M. et al. Uncertainty in the timing of origin of animals and the limits of precision in molecular timescales. *Curr. Biol.* **25**, 2939–2950 (2015).
- Droser, M. L., Tarhan, L. G. & Gehling, J. G. The rise of animals in a changing environment: global ecological innovation in the late Ediacaran. *Annu. Rev. Earth Planet. Sci.* **45**, 593–617 (2017).
- Budd, G. E. & Jensen, S. The origin of the animals and a ‘Savannah’ hypothesis for early bilaterian evolution. *Biol. Rev. Camb. Philos. Soc.* **92**, 446–473 (2017).
- Budd, G. E. The Cambrian fossil record and the origin of the phyla. *Integr. Comp. Biol.* **43**, 157–165 (2003).
- Zhu, M., Zhuravlev, A., Yu, W., Wood, R., Zhao, F. & Sukhov, S. S. A deep root for the Cambrian Explosion: implications of new bio- and chemostratigraphy from the Siberian Platform. *Geology* **45**, 459–462 (2016).
- Darroch, S. A. F., Smith, E. F., Laflamme, M. & Erwin, D. H. Ediacaran extinction and Cambrian Explosion. *Trends Ecol. Evol.* **33**, 653–663 (2018).
- Tarhan, L. G., Droser, M. L., Cole, D. B. & Gehling, J. G. Ecological expansion and extinction in the late Ediacaran: weighing the evidence for environmental and biotic drivers. *Integr. Comp. Biol.* **58**, 688–702 (2018).
- Tarhan, L. G. The early Paleozoic development of bioturbation – evolutionary and geobiological consequences. *Earth Sci. Rev.* **178**, 177–207 (2018).
- Servais, T., Owen, A. W., Harper, D. A. T., Kröger, B. & Munnecke, A. The great Ordovician biodiversification event (GOBE): the palaeoecological dimension. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **294**, 99–119 (2010).
- Zhuravlev, A. Y. & Wood, R. A. Anoxia as the cause of the mid-Early Cambrian (Botomian) extinction event. *Geology* **24**, 311–314 (1996).
- Canfield, D. E. & Farquhar, J. Animal evolution, bioturbation, and the sulfate concentration of the oceans. *Proc. Natl Acad. Sci. USA* **106**, 8123–8127 (2009).
- Li, C. et al. Ediacaran marine redox heterogeneity and early animal ecosystems. *Sci. Rep.* **5**, 17097 (2015).
- Tostevin, R. et al. Low-oxygen waters limited habitable space for early animals. *Nat. Commun.* **7**, 12818 (2016).
- Lenton, T.M. & Daines, S.J. The effects of marine eukaryote evolution on phosphorus, carbon and oxygen cycling across the Proterozoic–Phanerozoic transition. *Emerg. Top. Life Sci.* ETL20170156 (2018).
- van de Velde, S., Mills, B. J. W., Meysman, F. J. R., Lenton, T. M. & Poulton, S. W. Early Palaeozoic ocean anoxia and global warming driven by the evolution of shallow burrowing. *Nat. Commun.* **9**, 2554 (2018).
- Schiffbauer, J. D. et al. The latest Ediacaran Wormworld fauna: setting the ecological stage for the Cambrian Explosion. *GSA Today* **26**, 4–11 (2016).
- Butterfield, N. J. Oxygen, animals and oceanic ventilation: an alternative view. *Geobiology* **7**, 1–7 (2009).
- Wood, R. & Erwin, D. H. Innovation not recovery: dynamic redox promotes metazoan radiations. *Biol. Rev. Camb. Philos. Soc.* **93**, 863–873 (2018).
- Sperling, E. A. & Stockey, R. G. The temporal and environmental context of early animal evolution: considering all the ingredients of an ‘explosion’. *Integr. Comp. Biol.* **58**, 605–622 (2018).
- Planavsky, N. J. et al. Late Proterozoic transitions in climate, oxygen, and tectonics, and the rise of complex life. *Paleontol. Soc. Papers (Earth-life transitions: paleobiology in the context of Earth system evolution)* **21**, 47–82 (2015).
- Grotzinger, J. P., Fike, D. A. & Fischer, W. W. Enigmatic origin of the largest-known carbon isotope excursion in Earth’s history. *Nat. Geosci.* **4**, 285–292 (2011).
- Sperling, E. A. et al. Oxygen, ecology, and the Cambrian radiation of animals. *Proc. Natl Acad. Sci. USA* **110**, 13446–13451 (2013).
- Cloud, P. E. Jr. Atmospheric and hydrospheric evolution on the primitive earth. Both secular accretion and biological and geochemical processes have affected earth’s volatile envelope. *Science* **160**, 729–736 (1968).
- Knoll, A. H. & Sperling, E. A. Oxygen and animals in Earth history. *Proc. Natl Acad. Sci. USA* **111**, 3907–3908 (2014).
- Lenton, T. M., Boyle, R. A., Poulton, S. W., Shields-Zhou, G. A. & Butterfield, N. J. Co-evolution of eukaryotes and ocean oxygenation in the Neoproterozoic era. *Nat. Geosci.* **7**, 257–265 (2014).
- Sperling, E. A. et al. Statistical analysis of iron geochemical data suggests limited late Proterozoic oxygenation. *Nature* **523**, 451–454 (2015).
- Guilbaud, R. et al. Oxygen minimum zones in the early Cambrian ocean. *Geochem. Perspect. Lett.* **6**, 33–38 (2018).
- Hammarlund, E. U. et al. Early Cambrian oxygen minimum zone-like conditions at Chengjiang. *Earth Planet. Sci. Lett.* **475**, 160–168 (2017).
- Bowyer, F., Wood, R. A. & Poulton, S. W. Controls on the evolution of Ediacaran metazoan ecosystems: a redox perspective. *Geobiology* **15**, 516–551 (2017).
- Mills, D. B. et al. Oxygen requirements of the earliest animals. *Proc. Natl Acad. Sci. USA* **111**, 4168–4172 (2014).
- Mills, D. B. et al. The last common ancestor of animals lacked the HIF pathway and respired in low-oxygen environments. *eLife* **7**, e31176 (2018).
- Wood, R. A. et al. Dynamic redox conditions control late Ediacaran ecosystems in the Nama Group, Namibia. *Precamb. Res.* **261**, 252–271 (2015).
- Tostevin, R. et al. Uranium isotope evidence for an expansion of anoxia in terminal Ediacaran oceans. *Earth Planet. Sci. Lett.* **506**, 104–112 (2018).
- Wang, D. et al. Coupling of ocean redox and animal evolution during the Ediacaran–Cambrian transition. *Nat. Commun.* **9**, 2575 (2018).
- Chen, X. et al. Rise to modern levels of ocean oxygenation coincided with the Cambrian radiation of animals. *Nat. Commun.* **6**, 7142 (2015).
- Sahoo, S. K. et al. Oceanic oxygenation events in the anoxic Ediacaran ocean. *Geobiology* **14**, 457–468 (2016).
- Li, C. et al. Coupled oceanic oxygenation and metazoan diversification during the early–middle Cambrian? *Geology* **45**, 743–746 (2017).
- Zhang, J. et al. Heterogeneous oceanic redox conditions through the Ediacaran–Cambrian boundary limited the metazoan zonation. *Sci. Rep.* **7**, 8550 (2017).
- Zhang, F. et al. Extensive marine anoxia during the terminal Ediacaran Period. *Sci. Adv.* **4**, ean8983 (2018).
- Wei, G.-Y. et al. Marine redox fluctuation as a potential trigger for the Cambrian Explosion. *Geology* **46**, 587–590 (2018).
- Zhuravlev, A. Y. & Wood, R. A. The two phases of the Cambrian Explosion. *Sci. Rep.* **8**, 16656 (2018).
- Woods, M. A., Wilby, P. R., Leng, M. J., Rushton, A. W. & Williams, M. The Furongian (late Cambrian) Steptoean positive carbon isotope excursion (SPICE) in Avalonia. *J. Geol. Soc. Lond.* **164**, 851–862 (2011).
- Muscante, A. D., Boag, T. H., Bykova, N. & Schiffbauer, J. D. Environmental disturbance, resource availability, and biologic turnover at the dawn of animal life. *Earth Sci. Rev.* **177**, 248–264 (2017).
- Laflamme, M., Darroch, S. A. F., Tweedt, S. M., Peterson, K. J. & Erwin, D. H. The end of the Ediacara biota: extinction, biotic replacement, or Cheshire Cat? *Gondwana Res.* **23**, 558–573 (2013).
- Darroch, S. A. F. et al. Biotic replacement and mass extinction of the Ediacara biota. *Proc. Biol. Sci.* **282**, 20151003 (2015).
- Chen, Z. et al. Trace fossil evidence for Ediacaran bilaterian animals with complex behaviors. *Precamb. Res.* **224**, 690–701 (2013).
- Chen, Z., Chen, X., Zhou, C., Yuan, X. & Xiao, S. Late Ediacaran trackways produced by bilaterian animals with paired appendages. *Sci. Adv.* **4**, eaa06691 (2018).
- Gehling, J. G. & Droser, M. Ediacaran scavenging as a prelude to predation. *Emerg. Top. Life Sci.* **2**, 213–222 (2018).
- Wilby, P. R., Carney, J. N. & Howe, M. P. A rich Ediacaran assemblage from eastern Avalonia: evidence of early widespread diversity in the deep ocean. *Geology* **39**, 655–658 (2011).
- Pu, J. P. et al. Dodging snowballs: geochronology of the Gaskiers glaciation and the first appearance of the Ediacaran biota. *Geology* **44**, 955–958 (2016).
- Canfield, D. E. & Poulton, S. W. & Narbonne, G.M. Late-Neoproterozoic deep-ocean oxygenation and the rise of animal life. *Science* **315**, 92–95 (2007).
- Gehling, J. G. & Droser, M. How well do fossil assemblages of the Ediacara Biota tell time? *Geology* **41**, 447–450 (2013).
- Jensen, S., Droser, M. L. & Gehling, J. G. in *Neoproterozoic Geobiology and Paleobiology* (eds Xiao, S. and Kaufman) 115–157 (Springer, New York, 2006).
- Grazhdankin, D. Patterns of distribution in the Ediacaran biotas: facies versus biogeography and evolution. *Paleobiology* **30**, 203–221 (2004).
- Evans, S. D., Diamond, C. W., Droser, M. L. & Lyons, T. W. Dynamic oxygen and coupled biological and ecological innovation during the second wave of the Ediacara biota. *Emerg. Top. Life Sci.* **2**, 223–233 (2018).
- Chen, Z. et al. New Ediacara fossils preserved in marine limestone and their ecological implications. *Sci. Rep.* **4**, 4180 (2014).
- Ling, H.-F. et al. Cerium anomaly variations in Ediacaran-earliest Cambrian carbonates from the Yangtze Gorges area, South China: implications for oxygenation of coeval shallow seawater. *Precamb. Res.* **225**, 110–127 (2013).
- Duda, J.-P. et al. Geobiology of a palaeoecosystem with Ediacara-type fossils: the Shibantan Member (Dengying Formation, South China). *Precamb. Res.* **255**, 48–62 (2014).
- Zhang, L. Y. A discovery and preliminary study of the late stage of late Gaojiashan biota from Sinian in Ningqiang County, Shaanxi. *Northwest Geoscience* **13**, 67–88 (1986).
- Hua, H., Chen, Z. & Yuan, X. The advent of mineralized skeletons in Neoproterozoic Metazoa — new fossil evidence from the Gaojiashan Fauna. *Geol. J.* **42**, 263–279 (2007).
- Xing, Y.-S., Ding, Q.-X., Luo, H.-L., He, T.-G. & Wang, Y.-G. The Sinian–Cambrian boundary of China. *Bulletin of the Institute of Geology of the Chinese Academy Special Issue* **10**, 182–183 (1984).

63. Cai, Y., Schiffbauer, J. D., Hua, H. & Xiao, S. Morphology and paleoecology of the late Ediacaran tubular fossil *Conotubus hemiannulatus* from the Gaojiashan Lagerstätte of southern Shaanxi Province. *South China. Precamb. Res.* **191**, 46–57 (2011).
64. Chen, L. Y., Chu, X. L., Zhang, X. L. & Zhai, M. G. Carbon isotopes, sulfur isotopes, and trace elements of the dolomites from the Dengying Formation in Zhenba area southern Shaanxi: implications for shallow water redox conditions during the terminal Ediacaran. *Sci. China Earth Sci.* **58**, 1107–1122 (2015).
65. Saylor, B. Z. Sequence stratigraphy and carbonate–siliciclastic mixing in a terminal Proterozoic foreland basin, Urusis Formation, Nama Group, Namibia. *J. Sediment. Res.* **73**, 264–279 (2003).
66. Jensen, S. M. & Runnegar, B. N. A complex trace fossil from the Spitskop Member (terminal Ediacaran–? Lower Cambrian) of southern Namibia. *Geol. Mag.* **142**, 561–569 (2005).
67. Wood, R. A. Paleocology of the earliest skeletal metazoan communities: implications for early biomineralization. *Earth Sci. Rev.* **106**, 184–190 (2011).
68. Murdock, D. J. E. & Donoghue, P. C. J. Evolutionary origins of animal skeletal biomineralization. *Cells Tissues Organs* **194**, 98–102 (2011).
69. Wood, R. & Penny, A. Substrate growth dynamics and biomineralization of an Ediacaran encrusting poriferan. *Proc. Biol. Sci.* **285**, 20171938 (2018).
70. Wood, R., Ivantsov, A. Y. & Zhuravlev, A. Y. First macrobiota biomineralization was environmentally triggered. *Proc. Biol. Sci.* **284**, 20170059 (2017).
71. Porter, S. M. Seawater chemistry and early carbonate biomineralization. *Science* **316**, 1302 (2007).
72. Brennan, S. T., Lowenstein, T. K. & Horita, J. Seawater chemistry and the advent of biocalcification. *Geology* **32**, 473–476 (2004).
73. Wood, R., Zhuravlev, A. Yu, Sukhov, S. S., Zhu, M. & Zhao, F. Demise of Ediacaran dolomitic seas marks widespread biomineralization on the Siberian Platform. *Geology* **45**, 27–30 (2017).
74. Clapham, M. E., Narbonne, G. M. & Gehling, J. G. Paleocology of the oldest known animal communities: Ediacaran assemblages at Mistaken Point, Newfoundland. *Paleobiology* **29**, 527–544 (2003).
75. Droser, M., Gehling, J. & Jensen, S. Assemblage palaeoecology of the Ediacara biota: the unabridged edition? *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **232**, 131–147 (2006).
76. Boag, T. H., Darroch, S. A. F. & Laflamme, M. Ediacaran distributions in space and time: testing assemblage concepts of earliest macroscopic body fossils. *Paleobiology* **42**, 574–594 (2016).
77. Jensen, S., Gehling, J. G. & Droser, M. L. Ediacara-type fossils in Cambrian sediments. *Nature* **393**, 567 (1998).
78. Hagadorn, J. W., Fedo, C. M. & Waggoner, B. M. Early Cambrian Ediacaran-type fossils from California. *J. Paleontol.* **74**, 731–740 (2000).
79. Shu, D.-G. et al. Lower Cambrian vendobionts from China and early diploblast evolution. *Science* **312**, 731–734 (2006).
80. Hoyal Cuthill, J. F. & Han, J. Cambrian petalonamid *Stromatoveris* phylogenetically links Ediacaran biota to later animals. *Palaeontology* **61**, 813–823 (2018).
81. Conway Morris, S. Ediacaran-like fossils in Cambrian Burgess Shale-type faunas of North America. *Palaeontology* **36**, 593–635 (1993).
82. Zhuravlev, A., Yu, Linan, E., Vintaned, J. A. G., Debrenne, F. & Fedorov, A. B. New finds of skeletal fossils in the terminal Neoproterozoic of the Siberian Platform and Spain. *Acta Palaeontol. Pol.* **57**, 205–224 (2012).
83. Yang, B. et al. Transitional Ediacaran–Cambrian small skeletal fossil assemblages from South China and Kazakhstan: implications for chronostratigraphy and metazoan evolution. *Precamb. Res.* **285**, 202–215 (2016).
84. McLroy, D., Green, O. R. & Brasier, M. D. Palaeobiology and evolution of the earliest agglutinated Foraminifera: *Platysolenites*, *Spirosolenites* and related forms. *Lethaia* **34**, 13–29 (2001).
85. Kontorovich, A. E. et al. A section of Vendian in the east of West Siberian Plate (based on data from the Borehole Vostok 3). *Russ. Geol. Geophys.* **49**, 932–939 (2008).
86. Budd, G. E. Early animal evolution and the origins of nervous systems. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **370**, 20150037 (2015).
87. Yang, C., Li, X. H., Zhu, M. & Condon, D. J. SIMS U–Pb zircon geochronological constraints on upper Ediacaran stratigraphic correlations. *South China. Geol. Mag.* **154**, 1202–1216 (2017).
88. Martin, M. W. et al. Age of Neoproterozoic bilaterian body and trace fossils, White Sea, Russia: implications for metazoan evolution. *Science* **288**, 841–845 (2000).
89. Bowring, S. A. et al. Geochronologic constraints on the chronostratigraphic framework of the Neoproterozoic Huqf Supergroup, Sultanate of Oman. *Am. J. Sci.* **307**, 1097–1145 (2007).
90. Waggoner, B. Biogeographic analyses of the Ediacara biota: a conflict with paleotectonic reconstructions. *Paleobiology* **25**, 440–458 (1999).
91. Smith, E. F. et al. The end of the Ediacaran: two new exceptionally preserved body fossil assemblages from Mount Dunfee, Nevada, USA. *Geology* **44**, 911–914 (2016).
92. Love, G. D. et al. Fossil steroids record the appearance of Demospongiae during the Cryogenian period. *Nature* **457**, 718–721 (2009).
93. Brocks, J. J. et al. The rise of algae in Cryogenian oceans and the emergence of animals. *Nature* **548**, 578–581 (2017).
94. Yuan, X., Chen, Z., Xiao, S., Zhou, C. & Hua, H. An early Ediacaran assemblage of macroscopic and morphologically differentiated eukaryotes. *Nature* **470**, 390–393 (2011).
95. Xiao, S., Zhang, Y. & Knoll, A. H. Three-dimensional preservation of algae and animal embryos in a Neoproterozoic phosphorite. *Nature* **391**, 553–558 (1998).
96. Liu, A. G., McLroy, D., Matthews, J. J. & Brasier, M. D. A new assemblage of juvenile Ediacaran fronds from the Drook Formation, Newfoundland. *J. Geol. Soc. Lond.* **169**, 395–340 (2012).
97. Liu, A. G., McLroy, D. & Brasier, M. D. First evidence for locomotion in the Ediacara biota from the 565 Ma Mistaken Point Formation, Newfoundland. *Geology* **38**, 123–126 (2010).
98. Germs, G. J. B. New shelly fossils from the Nama Group, South West Africa. *Am. J. Sci.* **272**, 752–761 (1972).
99. Bengtson, S. & Zhao, Y. Predatorial borings in late Precambrian mineralized exoskeletons. *Science* **257**, 367–369 (1992).
100. Landing, E. Precambrian–Cambrian boundary global stratotype ratified and a new perspective of Cambrian time. *Geology* **22**, 179–182 (1994).
101. Macdonald, F. A. et al. Calibrating the Cryogenian. *Science* **327**, 1241–1243 (2010).
102. Macdonald, F. A. et al. The stratigraphic relationship between the Shuram carbon isotope excursion, the oxygenation of Neoproterozoic oceans, and the first appearance of the Ediacara biota and bilaterian trace fossils in northwestern Canada. *Chem. Geol.* **362**, 250–272 (2013).
103. Zhu, M., Babcock, L. E. & Peng, S.-C. Advances in Cambrian stratigraphy and paleontology: Integrating correlation techniques, paleobiology, taphonomy and paleoenvironmental reconstruction. *Palaeoworld* **15**, 217–222 (2006).
104. Canfield, D. E. et al. Ferruginous conditions dominated later Neoproterozoic deep-water chemistry. *Science* **321**, 949–952 (2008).
105. Kendall, B. et al. Uranium and molybdenum isotope evidence for an episode of widespread ocean oxygenation during the late Ediacaran Period. *Geochim. Cosmochim. Acta* **156**, 173–193 (2015).
106. Dahl, T. W. et al. Devonian rise in atmospheric oxygen correlated to the radiations of terrestrial plants and large predatory fish. *Proc. Natl Acad. Sci. USA* **107**, 17911–17915 (2010).
107. Narbonne, G. M. & Gehling, J. G. Life after snowball: the oldest complex Ediacaran fossils. *Geology* **31**, 27–30 (2003).
108. Dunn, F. S., Liu, A. G. & Donoghue, P. C. J. Ediacaran developmental biology. *Biol. Rev. Camb. Philos. Soc.* **93**, 914–932 (2018).
109. Liu, A. G., Matthews, J. J., Menon, L. R., McLroy, D. & Brasier, M. D. *Haootia quadriformis* n. gen., n. sp., interpreted as a muscular cnidarian impression from the Late Ediacaran period (approx. 560 Ma). *Proc. Biol. Sci.* **281**, 20141202 (2014).
110. Amthor, J. E. et al. Extinction of *Cloudina* and *Namacalathus* at the Precambrian–Cambrian boundary in Oman. *Geology* **31**, 431–434 (2003).
111. Zhu, M., Zhang, J. & Yang, A. Integrated Ediacaran (Sinian) chronostratigraphy of South China. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **254**, 7–61 (2007).
112. Zhu, M. et al. Early Cambrian stratigraphy of east Yunnan, southwestern China: a synthesis. *Acta Palaeontologica Sin.* **40**, 4–39 (2001).
113. Zhu, M. & Li, X.-H. Introduction: from Snowball Earth to Cambrian explosion — evidence from China. *Geol. Mag.* **154**, 1187–1192 (2017).
114. Ahn, S. Y. & Zhu, M. Lowermost Cambrian acritarchs from the Yanjiabe Formation, South China: implication for defining the base of the Cambrian in the Yangtze Platform. *Geol. Mag.* **154**, 1217–1231 (2017).
115. Saltzman, M. R., Edwards, C. T., Adrain, J. M. & Westrop, S. R. Persistent oceanic anoxia and elevated extinction rates separate the Cambrian and Ordovician radiations. *Geology* **43**, 807–810 (2015).
116. Boyle, R. A., Dahl, T. W., Bjerrum, C. J. & Canfield, D. E. Bioturbation and directionality in Earth's carbon isotope record across the Neoproterozoic–Cambrian transition. *Geobiology* **16**, 252–278 (2018).
117. Shields, G. A. & Mills, B. J. W. Tectonic controls on the long-term carbon isotope mass balance. *Proc. Natl Acad. Sci. USA* **114**, 4318–4323 (2017).
118. Pogge von Strandmann, P. A. E. et al. Selenium isotope evidence for progressive oxidation of the Neoproterozoic biosphere. *Nat. Commun.* **6**, 10157 (2015).
119. Wen, H. et al. Molybdenum isotopic records across the Precambrian–Cambrian boundary. *Geology* **39**, 775–778 (2011).

120. Kimura, H. & Watanabe, Y. Oceanic anoxia at the Precambrian–Cambrian boundary. *Geology* **29**, 995–998 (2001).
121. Wille, M., Nägler, T. F., Lehmann, B., Schröder, S. & Kramers, J. D. Hydrogen sulphide release to surface waters at the Precambrian/Cambrian boundary. *Nature* **453**, 767–769 (2008).
122. Tostevin, R. et al. Constraints on the late Ediacaran sulfur cycle from carbonate associated sulfate. *Precamb. Res.* **230**, 113–125 (2017).
123. Decechi, T. A., Narbonne, G. M., Greentree, C. & Laflamme, M. Relating Ediacaran fronds. *Paleobiology* **43**, 171–180 (2017).
124. Ivantsov, A. Y. Feeding traces of Proarticulata – the Vendian Metazoa. *Paleontol. J.* **45**, 237–248 (2011).
125. Bobrovskiy, I. et al. Ancient steroids establish the Ediacaran fossil *Dickinsonia* as one of the earliest animals. *Science* **361**, 1246–1249 (2018).
126. Carbone, C. A. & Narbonne, G. M. When life got smart: the evolution of behavioral complexity through the Ediacaran and Early Cambrian of NW Canada. *J. Paleontol.* **88**, 309–330 (2014).
127. Buatois, L.A. & Mángano, M.G. In *The Trace-Fossil Record of Major Evolutionary Events, Topics in Geobiology* Vol. 39 (eds. Mángano, M.G. & Buatois, L.A.) 27–72 (Springer, 2016).
128. Penny, A. M. et al. Ediacaran metazoan reefs from the Nama Group, Namibia. *Namibia. Science* **344**, 1504–1506 (2014).
129. Mehra, A. & Maloof, A. Multiscale approach reveals that *Cloudina* aggregates are detritus and not in situ reef constructions. *Proc. Natl Acad. Sci. USA* **115**, E2519–E2527 (2018).
130. Bengtson, S. Origins and early evolution of predation. *Paleontological Society Papers* **8**, 289–318 (2002).

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Author contributions

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Competing interests

The authors declare no competing financial interests.

Additional information

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