

1                   **The utility of height for the Ediacaran organisms of Mistaken Point**

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11 **Ediacaran fossil communities consist of the oldest macroscopic eukaryotic organisms.**  
12 **Increased size (height) is hypothesized to be driven by competition for water-column**  
13 **resources, leading to vertical/epifaunal tiering and morphological innovations such as**  
14 **stems. Using spatial analyses, we find no correlation between tiering and resource**  
15 **competition, and that stemmed organisms are not tiered. Instead, we find height is**  
16 **correlated to greater offspring dispersal, demonstrating the importance of colonization**  
17 **potential over resource competition.**

18 Bedding-plane assemblages of Ediacaran fossils at Mistaken Point, Newfoundland (~566  
19 Ma)<sup>1</sup>, are among the oldest known eukaryotic microfossil communities<sup>2</sup>. In extant marine  
20 ecosystems, body size is key to structuring communities, due to size-structured predation  
21 dynamics<sup>3,4</sup>. However, the Mistaken Point communities pre-date macro-predation and  
22 (extensive) mobility<sup>5</sup>, and so body size must have played a different role. Instead, the driver of  
23 large size has been suggested to be competition for vertically distributed water-column  
24 resources, resulting in different taxa occupying different parts of the water column – a process  
25 known as tiering<sup>6</sup>. Consequently, tiering to avoid resource competition has been interpreted as  
26 the major driver in the diversification of Ediacaran body plans, most notably in the evolution of a  
27 non-branched (i.e. “naked”) stem<sup>7-9</sup>. Since Mistaken Point bedding planes consist of sessile  
28 organisms preserved *in-situ*, it is reasonable to assume that approximately all of the macroscopic  
29 organisms were preserved, so the bedding planes represent a near-census of the community at the  
30 time of burial<sup>2</sup>. Therefore, detailed statistical analyses of these populations and their spatial  
31 distributions can be used to determine the relationship between height and resource  
32 competition<sup>10,11</sup>.

33 We analysed the communities of three large bedding-plane assemblages in the Mistaken  
34 Point Ecological Reserve: the ‘D’, ‘E’, and Lower Mistaken Point (LMP) surfaces<sup>12</sup>, using the  
35 data from [11] (Supplementary Figure 1). These communities are dominated by rangeomorphs,  
36 a clade of “fractally-branching” organisms<sup>13,14,15</sup> with some taxa also possessing a naked stem<sup>7</sup>.  
37 These communities also include non-fractally branching frondose arboreomorphs<sup>16</sup>; the putative  
38 sponge *Thectardis*; fronds awaiting formal description (e.g. “Ostrich Feathers”)<sup>2</sup>; and irregular  
39 bedding-plane features referred to as ivesheadiomorphs and “Lobate Discs”<sup>2,10,17</sup> (see Methods  
40 for details). Community composition differs between the three communities, with the ‘D’ surface  
41 notably different due to exclusive population by rangeomorphs with no abundant stemmed taxa  
42 (Supplementary Table 2). All three assemblages occur within deep-marine turbidite sequences<sup>2</sup>,  
43 with fossils preserved as external moulds in siltstone hemipelagites, cast from above by  
44 volcanoclastic deposits<sup>18</sup> (Supplementary Figure 1). A volcanic tuff directly above the ‘E’ surface  
45 has been dated to 566.25±0.35Ma, which provides an upper age constraint on the underlying ‘D’  
46 and LMP surfaces<sup>19</sup>.

47 To quantify the extent of tiering, we calculated the percentage by which each taxon’s  
48 population exhibits distinct vertical stratification (*DVS*) with respect to the rest of the community  
49 (Supplementary Figure 2). The extent of community tiering is defined as the mean taxa *DVS*.  
50 Two different *DVS* metrics were calculated: height-based  $DVS^{height}$  and uptake-zone  $DVS^{uptake}$ .  
51 The taxon-specific  $DVS^{height}$  is defined as the percentage of specimens within the taxon  
52 population that are not matched in height by any specimen from a different taxonomic group  
53 (Supplementary Figure 2). Taxon-specific  $DVS^{uptake}$  is defined as the percentage of specimens  
54 within a taxon population whose “uptake-zone” (i.e. the branching organism part) is not in the  
55 same part of the water column as the uptake-zone of specimens from a different taxonomic group

56 (Supplementary Figure 2 and Methods). Consequently,  $DVS=0\%$  corresponds to no tiering while  
57  $DVS=100\%$  corresponds to a completed tiered community.

58 Competition was detected and quantified using spatial point process analyses, whereby  
59 pair correlation functions (PCFs) were calculated to describe the spatial distributions between  
60 pairs of taxa on each bedding plane<sup>20</sup>, with a  $PCF=1$  indicating a distribution that was  
61 completely spatially random (CSR),  $PCF>1$  indicating aggregation, and  $PCF<1$  indicating  
62 segregation<sup>20-22</sup>. Monte Carlo simulations and Diggle's goodness-of-fit test<sup>22</sup> ( $p_d$ ) were used to  
63 indicate significantly non-CSR distributions when the observed PCF deviated outside the  
64 simulation envelope coupled with a  $p_d \ll 1$ . Where bivariate spatial segregation was detected,  
65 partial PCF between size-classes (defined in Methods; Supplementary Figure 3) were calculated,  
66 and Diggle's segregation test<sup>23</sup> used to assess segregation of each size class. Identifying the  
67 processes behind spatial patterns is not straightforward<sup>22-27</sup>; however, inter-specific resource  
68 competition typically generates a segregated pattern, with segregated largest specimens and CSR  
69 or aggregated small specimens<sup>21</sup>. To further investigate the relationship of height with dispersal  
70 dynamics, the mean cluster radius was calculated by fitting univariate Thomas cluster models to  
71 the univariate PCFs<sup>27</sup> (Supplementary Table 4). Linear regressions of these radii were then fitted  
72 to mean height, maximum height and mean uptake-zone height for each frondose taxon  
73 (Supplementary Table 5).

74 Only the 'D' surface was found to exhibit high  $DVS$  (80.1% , Figure 1, Supplementary  
75 Table 1.  $DVS^{height}_D = DVS^{uptake}_D$ ). In contrast, the  $DVS^{height}$  for the 'E' surface community is only  
76 12.4%, and only 20.0% for the LMP community (Supplementary Table 1),  $DVS^{uptake}$  was larger  
77 than  $DVS^{height}$  ( $DVS^{uptake}_E = 44.9\%$ ;  $DVS^{uptake}_{LMP} = 40.9\%$ ), but still under 50%. Taxon  $DVS^{height}$   
78 and  $DVS^{uptake}$  are not significantly different between the LMP, 'D' or 'E' communities ( $p=0.10$

79 and  $p=0.37$ ) or  $DVS^{height}$  between ‘D’ and ‘E’ communities ( $p=0.03$ ;  $\alpha=0.016$ ). There are no  
80 instances of large spatial-scale bivariate segregation on the ‘D’ surface and two on the ‘E’  
81 surface (cf. [10]); Figure 2 and Supplementary Table 3). On the ‘E’ surface, spatial segregation  
82 is found between *Fractofusus* and Feather Dusters ( $PCF_{Min}=0.8852$ ;  $p=0.01$ ), and between  
83 Feather Dusters and *Charniodiscus* ( $PCF_{Min}=0.8972$ ;  $p=0.01$ ) with segregation detected between  
84 large specimens (both  $p=0.01$ ), but not between small specimens ( $p_{feaD-Fract}=0.25$  and  $p_{feaD-}$   
85  $Chard=0.14$ ; Supplementary Table 3, Figure 2a,b). Therefore, habitat segregation is excluded as  
86 the underlying cause of these spatial segregations, and so they most likely reflect resource  
87 competition. For LMP, segregation occurs between Charniid I and Ostrich Feather ( $PCF_{Min}=0.$   
88  $4932$ ;  $p=0.01$ ; Figure 2d), and between Charniid II and Ostrich Feather ( $PCF_{Min}=0.5346$ ;  
89  $p=0.01$ ; Figure 2c). The large specimens of Charniid II and Ostrich Feather were segregated  
90 ( $p=0.01$ ), while small specimens were aggregated ( $p=0.92$ ) thus resource competition is the most  
91 likely underlying process. However, the Charniid I – Ostrich Feather bivariate distribution was  
92 segregated across all size classes ( $p_{small}=0.02$  and  $p_{large}=0.01$ ; Supplementary Table 3), thus  
93 likely reflecting habitat segregation rather than competition.

94 If resource competition dominates community dynamics and leads to tiering, then the  
95 extent of  $DVS^{height}$  and/or  $DVS^{uptake-zone}$  should predict whether two taxa exhibit inter-specific  
96 competition, with high  $DVS$  taxon pairs not competing (as they occupy different parts of the  
97 water column). This resource competition-dominated community dynamic is consistent with the  
98 ‘D’ surface community, which exhibited high  $DVS$ , and had no instances of inter-specific  
99 resource competition (Figs. 2, 3; Supplementary Table 3). However, on the ‘E’ surface, the two  
100 instances of resource competition correspond to high levels of pairwise  $DVS^{uptake}$  with respect to  
101 both Feather Dusters – *Fractofusus* and Feather Dusters – *Charniodiscus* ( $DVS^{uptake}_{FeaD-}$

102  $Fract=75.1\%$ ;  $DVS^{uptake}_{FeaD-Chard}=60.3\%$ ; Supplementary Table 3). On the ‘E’ surface, Charniids  
103 and *Thectardis* both exhibit very low  $DVS^{uptake}$  levels ( $DVS^{uptake}_{Charniid}=10.4\%$  and  $DVS^{uptake}$   
104  $Thect=12.0\%$ ), but do not correspond to any of the instances of inter-specific competition  
105 identified; neither do the comparatively high levels of uptake-zone tiering correspond to the  
106 presence of resource competition (Figs. 2 and 3, Supplementary Table 1). The single LMP  
107 instance of resource competition, between Charniid II and Ostrich Feather, corresponded to a  
108 moderate level of pairwise  $DVS^{uptake}$  (38.8%), coupled to a very strong segregation  
109 ( $PCF_{Min}=0.4932$ ). A linear regression of the  $DVS^{uptake}$  with  $PCF_{Min}$  showed no significant  
110 relationship ( $p=0.283$ ), so our results from the ‘E’ and LMP surface provide no evidence that  
111 resource competition resulted in vertically tiered populations.

112 When the E’ surface taxa were subset into rangeomorphs/non-rangeomorphs, and  
113 stemmed/non-stemmed groups there were no significant differences in  $DVS^{uptake}$  or  $DVS^{height}$   
114 between rangeomorphs and non-rangeomorphs or stemmed and non-stemmed  $DVS^{uptake}$   
115 (Supplementary Table 2 ; all  $p \gg 0.1$ ). There was a significant difference in  $DVS^{height}$  between  
116 stemmed ( $DVS^{height}_{stem}=4.0\%$ ) and non-stemmed taxa ( $DVS^{height}_{non-stem}=19.9\%$ ;  $p=0.001$ ).

117 The development of stems has been hypothesized to enable organism uptake-zone to  
118 reach new water column heights, thus avoiding competition for resources<sup>7-9, 28</sup> such as oxygen, or  
119 the dissolved organic carbon which Mistaken Point organisms likely utilised<sup>28,29</sup> (see Ref [2] for  
120 further discussion). This hypothesis predicts that stemmed organisms should be more tiered (i.e.  
121 higher  $DVS$ ) than non-stemmed organisms, but our results disagree: non-stemmed taxa exhibit a  
122 significantly higher degree of  $DVS^{height}$  than stemmed taxa (Supplementary Table 3). Thus, naked  
123 stems likely had a different function, such as enabling greater offspring dispersal<sup>7</sup>. For dispersal-  
124 generated aggregations, cluster size (Supplementary Tables 4-5) was found to strongly correlate

125 with maximum height of ‘E’ surface organisms ( $R^2=0.997$ ,  $p=0.034$ ), but not with mean height  
126 or mean uptake-zone height (all  $p \gg 0.1$ ). This result demonstrates that maximum height  
127 directly resulted in greater offspring dispersal. Therefore, while stemmed organisms did not  
128 significantly benefit from the additional height for nutrient acquisition, they did gain increased  
129 offspring dispersal. While at least some Ediacaran species exhibited close-to-parent offspring  
130 dispersal due to non-waterborne, stolon dominated reproduction<sup>11</sup>, evidence of wide-spread  
131 dispersal<sup>30-34</sup> demonstrates the prevalence of Ediacaran waterborne propagation, and so the  
132 importance of colonization potential for Ediacaran macrofossils.

133 The lack of correlation between *DVS* and resource competition throughout Mistaken  
134 Point communities contradicts previous suggestions that competition for resources drove  
135 Ediacaran community ecology<sup>2-4,6,28</sup>. While increased height would have placed organisms in  
136 faster water flow<sup>8</sup>, increasing resource refresh rates, the lack of tiering within these communities  
137 demonstrates that these advantages were not significant. Additionally, we have shown that the  
138 advantage of height in these communities was a larger radius of offspring clusters – representing  
139 increased dispersal distances. Therefore, our results point to reproduction, not limited resources,  
140 as the principal driver of the dynamics of these oldest complex macro-communities.

## 141 **Methods**

142 **Data.** We used the data compiled by Clapham et al. (2003)<sup>11,35</sup> from the Lower Mistaken Point  
143 (LMP), ‘D’ surface and ‘E’ surface which recorded the spatial position, size measurements and  
144 orientation of each fossil. Specimens were recorded as one of fourteen taxonomic groups of  
145 macrofossils, including two ‘bin’ groups<sup>36</sup>: 1) *Bradgatia*, 2) *Pectinifrons*, 3) *Thectardis*, 4)  
146 *Fractofusus andersoni* + *F. misraii*, 5) *Charniodiscus spinosus* + *C. procerus*, 6) “Feather  
147 Dusters” which includes *Plumeropriscum* and *Primocandlebrum*, 7) *Hiemalora*, 8)

148 Ivesheadiomorphs<sup>37</sup>, 9) Lobate Discs, which are interpreted either as taphomorphs  
149 (dead/decaying remains) or as microbial colonies<sup>2,10,17</sup>, 10) *Charnia* ‘A’ which consists of  
150 *Beothukis mistakensis*<sup>38,39</sup> (which dominates the ‘E’ surface) and *Charnia masoni*. 11) *Charnia*  
151 ‘B’ now reassigned as *Trepassia wardae*<sup>39</sup>. Charniid populations on Mistaken Point are  
152 dominated by *Beothukis* (only four individuals on the ‘E’ surface are true *Charnia* species),  
153 therefore direct comparison of data from this grouping with those from other taxonomic groups  
154 should be undertaken with caution. 12) “Ostrich Feathers” 13) “Holdfast Discs”, being all  
155 discoidal specimens of uncertain affinity, with or without associated stems, which lack sufficient  
156 detail to identify the taxon, 14) “Other Species” being rare forms that do not fall into any of the  
157 other groups; e.g., *Hapsidophyllas*.

158 **Methods.** Differential erosion has the potential to distort spatial analyses<sup>40</sup> so this data has been  
159 tested for impact of differential erosion using heterogeneous Poisson models to model possible  
160 sources of erosion<sup>11</sup>, with no significant effects found on ‘D’ and ‘E’ surfaces. In this study we  
161 fit three heterogeneous Poisson models to the LMP data, with the models dependent on  $x$  is  
162 North to South (parallel to strike),  $y$  is East to West (parallel to dip),  $xy$  is the distance from the  
163 South - East corner finding no significant erosional effect (all  $p < 0.01$ , where  $p = 1$  corresponds to  
164 a perfect model fit – the spatial distributions depend exactly on the covariant). The tectonically  
165 distorted data was retrodeformed by returning elongated holdfast discs to a circular outline<sup>6,18</sup>.

166 **Tiering metric.** We defined two different metrics for quantifying tiering: height Distinct  
167 Vertical Stratification ( $DVS^{height}$ ) and uptake-zone  $DVS^{uptake}$ .  $DVS^{height}$  is calculated by 1)  
168 creating a frequency table in 1cm bins of the height of each specimen within that taxon  
169 population. 2) A similar frequency table is created using the rest of the community. 3) The two  
170 frequency tables are subtracted from each other and then 4)  $DVS^{height}$  for each taxon is calculated

171 as the percentage of specimens remaining divided by the total number of specimens of that  
172 taxon. Community  $DVS^{height}$  is the mean of all the taxa  $DVS^{height}$ .  $DVS^{uptake}$  is calculated  
173 similarly, but the frequency tables are created by filling in every 1cm that the specimen uptake-  
174 zone occupies. For example, a 4cm *Bradgatia* would be represented by a count in the 0cm –  
175 4cm bin, whereas a 4cm *Charniodiscus* with a 1cm stem would be represented by a count in the  
176 1cm – 4cm bin. For example,  $DVS=0\%$  corresponds to no taxa occupying a unique part of the  
177 water column, i.e. the height distribution of that population is totally overlapped by the  
178 populations of other taxa.  $DVS=100\%$  corresponds to each taxon occupying a distinct stratum of  
179 the water column, i.e. there is no overlap between specimens of any taxa.

180 Alternative metrics, such as overlap of a range (such as the interquartile range, or 95%  
181 standard deviations) were ruled out because such range comparisons 1) assume a distribution e.g.  
182 normal or log-normal, which isn't necessarily accurate; 2) outliers (such the giant *Fronndophyllas*  
183 found on Lower Mistaken Point) severely bias the data and 3) such range metrics do not take  
184 into account relatively frequency – many populations had relatively few specimens at the end of  
185 their height range biasing the analyses.

186 Specimen heights were defined as the specimen length for *Bradgatia*, Charniid I,  
187 *Thectardis*; specimen width for *Pectinifrons*; stem length plus frond length for Charniid II,  
188 Feather Dusters, *Charniodiscus* and Ostrich Feathers. *Fractofusus* height was calculated a  
189 quarter of its width, thus assuming the *Fractofusus* has two vanes. It has been suggested that  
190 *Fractofusus* had three vanes<sup>41</sup> which would increase its vertical height. Repeating our analyses  
191 with height assuming three vanes reduces overall  $DVS^{height}_D$  by 9.3% to 70.8%, by 1.9% to  
192  $DVS^{height}_E=10.9\%$  and by 4.9% to  $DVS^{uptake}_E=40.0\%$ , so did not significantly change our results.  
193 Comparisons between  $DVS$  on the 'D', 'E' and LMP surfaces, and between the 'E' surface

194 community rangeomorphs/non-rangeomorphs and the stemmed/non-stemmed, were performed  
195 using Mann-Whitney tests. To account for the non-independence of the shared-sites in the  
196 pairwise comparisons of *DVS* on the ‘D’, ‘E’ and LMP surfaces, the significance level was set  $\alpha$   
197 =  $0.05/3 = 0.017$ , but note that such adjustment is likely to be too conservative.

198 **Data availability.** Access to the fossil localities is by scientific research permit only. Natural  
199 Areas Program, Canada for further information. Data used is publicly available at  
200 [https://figshare.com/articles/Mistaken\\_Point\\_Ediacaran\\_count\\_data/1111665](https://figshare.com/articles/Mistaken_Point_Ediacaran_count_data/1111665)

201 **Code availability.** The code defining these tiering metrics has been uploaded as an R package  
202 (tiering) to <https://cran.r-project.org/>.

203 **Spatial analyses.** Initial data exploration, inhomogeneous Poisson modelling, residual analysis  
204 and segregation tests<sup>23</sup> were performed in R<sup>42</sup> using the package spatstat<sup>43-45</sup>. Programita<sup>46-50</sup> was  
205 used to find distance measures and to perform aggregation model fitting (described in detail in  
206 references<sup>44,46-50</sup>).

207 Bivariate PCFs were calculated from the population density using a grid of 10cm x 10cm  
208 cells on the ‘D’ and ‘E’ surfaces, and 1cm x 1cm on LMP. To minimize noise a smoothing was  
209 applied to the PCF dependent on specimen abundance: A three cell smoothing over this grid was  
210 applied to the ‘D’ and ‘E’ surfaces, with five cells for LMP.

211 To test whether the PCF exhibited complete spatial randomness (CSR), 999 simulations  
212 were run for each relationship on a homogeneous background to generate simulation envelopes  
213 around the completely spatially random (CSR) which is where the PCF=. The fit of the fossil  
214 data to CSR was tested using Diggle’s goodness-of-fit test<sup>22</sup>  $p_d$  (where  $p_d=1$  corresponds to CSR,  
215 and  $p_d=0$  corresponds to non-CSR) with PCF deviations outside the simulation envelope coupled  
216 to a  $p_d \ll 1$  taken to indicate significantly non-CSR distributions. Note that due to non-

217 independence of spatial data, Monte-Carlo generated simulation envelopes cannot be interpreted  
218 as confidence intervals<sup>47</sup>, and also run the risk of Type I errors if the observed PCF falls near the  
219 edge of the simulation envelope<sup>21</sup> so that hypothesis testing needs to be further supplemented.  
220 None-the-less, if the observed data fell below the Monte-Carlo simulations, the bivariate  
221 distribution was described as segregated, and above the Monte-Carlo simulations the bivariate  
222 distribution was described as aggregated. Non-CSR distributions were tested for statistical  
223 significance using Diggle's goodness-of-fit test<sup>22</sup>, with segregations further tested using Diggle's  
224 segregation test<sup>23</sup> (Supplementary Table 3). Diggle's goodness-of-fit test, is a single test  
225 statistic<sup>21</sup> ( $p_d$ ) representing the total squared deviation between the observed pattern and the  
226 theoretical result across the studied distances. This test statistic was used in conjunction with  
227 visual inspection of Monte Carlo simulations for two reasons. First,  $p_d$  does not strictly test  
228 whether a model should be accepted or rejected, but whether the PCFs for the observed data are  
229 within the range of the stochastic realization of the model<sup>26</sup>. Second,  $p_d$  depends on the range  
230 over which it is calculated. Diggle's segregation test<sup>23</sup>, detects where two types (taxa here) are  
231 spatially segregated by calculating the sum of the square of the probability that each data point is a  
232 given type (taxa) minus the average fraction of data points which are a given type (taxa).

233 If a taxon was not randomly distributed on a homogeneous background, and was aggregated  
234 (Figure 2, Supplementary Table 4), the random model on a heterogeneous background was tested  
235 by creating a heterogeneous background from the density map of the taxon under consideration,  
236 being defined by a circle of radius  $R$  over which the density is averaged throughout the sample  
237 area. Density maps were formed using estimators within the range of  $0.1m < R < 1m$ , and the  $R$   
238 corresponding to the best-fit model was used. If excursions outside the simulation envelopes for

239 both homogeneous and heterogeneous Poisson models remained, then Thomas cluster models  
240 were fitted to the data as follows:

241

242 1. The PCF and L function<sup>51</sup> of the observed data were found. Both measures were  
243 calculated to ensure that the best-fit model is not optimized towards only one distance measure,  
244 and thus encapsulates all spatial characteristics.

245 2. Best-fit Thomas cluster processes<sup>52</sup> were fitted to the two functions where  $PCF > 1$ . The  
246 best-fit lines were not fitted to fluctuations around the random line of  $PCF = 1$  in order to aid good  
247 fit about the actual aggregations, and to limit fitting of the model about random fluctuations.  
248 Programita used the minimal contrast method<sup>21-23</sup> to find the best-fit model.

249 3. If the model did not describe the observed data well, the lines were refitted using just the  
250 PCF. If that fit was also poor, then only the L-function was used.

251 4. 99 simulations of this model were generated to create simulation envelopes, and the fit  
252 checked using the O-ring statistic<sup>46</sup>.

253 5.  $p_d$  was calculated over the model range. Very small-scale segregations (under 2cm) were  
254 not included in the model fitting, since they likely represent the finite size of the specimens, and  
255 the lack of specimen overlap.

256 6. If there were no excursions outside the simulation envelope and the  $p_d$ -value was high,  
257 then a univariate homogeneous Thomas cluster model was interpreted as the best model.

258 The most objective way to resolve the number and range of size classes in a population is  
259 by fitting height-frequency distribution data to various models, followed by comparison of  
260 (logarithmically scaled) Bayesian information criterion (BIC) values<sup>55</sup>, which we performed in R  
261 using the package MCLUST<sup>56</sup>. The number of populations thus identified was then used to

262 define the most appropriate size classes. A BIC value difference of  $> 10$  corresponds to a  
263 “decisive” rejection of the hypothesis that two models are the same, whereas values  $< 6$  indicate  
264 only weakly reject similarity of the models<sup>55-57</sup>.

265 Once defined, the PCFs for each size class were calculated, and segregated tests performed.  
266 Although it was necessary to set firm boundaries for each size class, the populations are normally  
267 distributed and therefore overlap. As a result, the largest individuals of the small population are  
268 grouped within the middle size class, while some of the smallest of the medium population are  
269 included within the small size class. As such, the medium population was excluded from  
270 analyses.

271 For each bivariate distribution displaying segregation, the size-classes of each taxon were  
272 calculated, the bivariate PCFs of the smallest size-classes and largest size-classes were plotted  
273 with 99 Monte Carlo simulations of a complete spatially random distribution and segregation  
274 tests performed.

275 **Regression analyses.** In order to investigate the relationship between height and dispersal linear  
276 regressions were performed in R<sup>41</sup>. Programita<sup>46-50</sup> was used to find the taxa whose univariate  
277 distributions were best modelled by Thomas Cluster models (thus most likely to be dispersal  
278 induced) and the best-fit cluster radius was used to indicate dispersal range. Four different  
279 height variables were found for each taxon’s population 1) Mean height 2) Maximum height, 3)  
280 Mean mid-point of uptake-zone and 4) Maximum mid-point of uptake zone. The uptake-zone  
281 mid-point for each specimen was calculated as the half-way point between the top of the stem  
282 and the top of the frond and was a proxy for dispersal release throughout the entire uptake-zone.

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## References

1. Pu, J.P., Bowring, S.A., Ramezani, J., Myrow, P., Raub, T.D., Landing, E., Mills, A., Hodgkin, E. and Macdonald, F.A.. Dodging snowballs: Geochronology of the Gaskiers glaciation and the first appearance of the Ediacaran biota. *Geology*, **44**, 955-958 (2016).
2. Liu, A. G, Kenchington C. G. & Mitchell, E. G. Remarkable insights into the paleoecology of the Avalonian Ediacaran biota. *Gondwana Res.* **27**, 1355–1380 (2015).
3. Butterfield, N. J. Animals and the invention of the Phanerozoic Earth system. *Trends in Ecology & Evolution* **26**, 81-87 (2011).
4. Woodward, G., Ebenman, B., Emmerson, M., Montoya, J.M., Olesen, J.M., Valido, A. and Warren, P.H.. Body size in ecological networks. *Trends in ecology & evolution*, **20**, 402-409 (2005).
5. Liu, A. G., McIlroy, D., & Brasier, M. D. First evidence for locomotion in the Ediacara biota from the 565 Ma Mistaken Point Formation, Newfoundland. *Geology*, **38**, 123-126 (2010).
6. Clapham, M. E., & Narbonne G. M., Gehling, J. G. Ediacaran epifaunal tiering. *Geology*, **30**, 627-630 (2002).
7. Laflamme, M., Flude, L. I., & Narbonne, G. M. Ecological tiering and the evolution of a stem: the oldest stemmed frond from the Ediacaran of Newfoundland, Canada. *Journal of Palaeontology*, **86**, 193-200 (2012).
8. Ghisalberti, M., et al.. Canopy flow analysis reveals the advantage of size in the oldest communities of multicellular eukaryotes. *Current Biology*, **24** 305-309 (2014).
9. Laflamme, M., & Narbonne, G. M.. Ediacaran fronds. *Palaeogeography, Palaeoclimatology, Palaeoecology*, **258**, 162-179 (2008).

- 305 10. Mitchell, E. G & Butterfield, N. J. Spatial analyses of Ediacaran communities at Mistaken  
306 Point. *Paleobiology*, **44**, 40-57. (2018).
- 307 11. Mitchell, E. G., Kenchington, C. G., Liu, A. G., Matthews, J. J., & Butterfield, N. J.  
308 Reconstructing the reproductive mode of an Ediacaran macro-organism. *Nature*, **524**, 343-  
309 346 (2015).
- 310 12. Clapham, M. E., Narbonne, G.M. & Gehling, J. G. Paleocology of the oldest known animal  
311 communities: Ediacaran assemblages at Mistaken Point, Newfoundland. *Paleobiology* **29**,  
312 527–544 (2003).
- 313 13. Landing, E., Narbonne, G.M. & Myrow, P. (eds) Trace fossils, small shelly fossils and the  
314 Precambrian–Cambrian boundary. *Bull. NY State Mus.* **463**, 1–81 (1988).
- 315 14. Narbonne, G. M. Modular construction of early Ediacaran complex life forms. *Science* **305**,  
316 1141–1144 (2004).
- 317 15. Hoyal Cuthill, J. F. & Conway Morris, S. Fractal branching organizations of Ediacaran  
318 rangeomorph fronds reveal a lost Proterozoic body plan. *Proc. Natl Acad. Sci. USA* **111**,  
319 13122–13126 (2014).
- 320 16. Brasier, M. D., Antcliffe, J. B. & Liu, A. G. The architecture of Ediacaran fronds.  
321 *Palaeontology* **55**, 1105–1124 (2012).
- 322 17. Liu, A. G., McIlroy, D., Antcliffe, J. B. & Brasier, M. D. Effaced preservation in the  
323 Ediacara biota and its implications for the early microfossil record. *Palaeontology* **54**, 607–  
324 630 (2011).
- 325 18. Wood, D. A., Dalrymple, R. W., Narbonne, G. M., Gehling, J. G. & Clapham, M. E.  
326 Paleoenvironmental analysis of the late Neoproterozoic Mistaken Point and Trepassey  
327 formations, southeastern Newfoundland. *Can. J. Earth Sci.* **40**, 1375–1391 (2003).

- 328 19. Benus, A. P. Sedimentological context of a deep-water Ediacaran fauna (Mistaken Point  
329 Formation, Avalon zone, eastern Newfoundland). *Bull. NY State Mus.* **463**, 8–9 (1988).
- 330 20. Illian, J., Penttinen, A., Stoyan, H. & Stoyan, D. *Statistical Analysis and Modelling of*  
331 *Spatial Point Patterns* Vol. 70 (John Wiley, 2008).
- 332 21. Diggle, P. *Statistical Analysis of Spatial Point Patterns* 2nd edn (Arnold, 2003).
- 333 22. Diggle, P., Zheng, P. and Durr, P.. Nonparametric estimation of spatial segregation in a  
334 multivariate point process: bovine tuberculosis in Cornwall, UK. *Journal of the Royal*  
335 *Statistical Society: Series C (Applied Statistics)*, **54**, 645-658 (2005).
- 336 23. Wiegand, T., Gunatilleke, S., Gunatilleke, N. & Okuda, T. Analyzing the spatial structure of  
337 a Sri Lankan tree species with multiple scales of clustering. *Ecology* **88**, 3088–3102 (2007).
- 338 24. Levin, S. A. in *Ecological Time Series Vol.2* (eds Powell, T. M. & Steele, J. H.) 277–326  
339 (Springer, 1995).
- 340 25. McIntire, E. J. & Fajardo, A. Beyond description: the active and effective way to infer  
341 processes from spatial patterns. *Ecology* **90**, 46–56 (2009).
- 342 26. Wiegand, T. & Moloney, K. A. *Handbook of Spatial Point-Pattern Analysis in Ecology*  
343 (CRC, 2013).
- 344 27. Murrell, D.J. and Law, R.. Heteromyopia and the spatial coexistence of similar  
345 competitors. *Ecology letters*, **6**,48-59 (2003).
- 346 28. Hoyal Cuthill, J.F. and Conway Morris, S. Nutrient-dependent growth underpinned the  
347 Ediacaran transition to large body size. *Nature ecology & evolution*, **1**,1201 (2017).
- 348 29. Laflamme, M., Xiao, S., & Kowalewski, M. Osmotrophy in modular Ediacara organisms.  
349 *Proc. Natl Acad. Sci. USA*, **106**, 14438-14443 (2009).

- 350 30. Darroch, S. A. F., Laflamme, M. & Clapham, M. E. Population structure of the oldest known  
351 macroscopic communities from Mistaken Point, Newfoundland. *Paleobiology* **39**, 591–608  
352 (2013).
- 353 31. Droser, M. L. & Gehling, J. G. Synchronous aggregate growth in an abundant new Ediacaran  
354 tubular organism. *Science* **319**, 1660–1662 (2008).
- 355 32. Penny, A. M. et al. Ediacaran metazoan reefs from the Nama Group, Namibia. *Science* **344**,  
356 1504–1506 (2014).
- 357 33. Yuan, X. et al. The Lantian biota: a new window onto the origin and early evolution of  
358 multicellular organisms. *Chin. Sci. Bull.* **58**, 701–707 (2013).
- 359 34. Hua, H., Chen, Z., Yuan, X., Zhang, L. & Xiao, S. Skeletogenesis and asexual reproduction  
360 in the earliest biomineralizing animal Cloudina. *Geology* **33**, 277–280 (2005).
- 361 35. Clapham, M. E. Ordination methods and the evaluation of Ediacaran communities.  
362 In: *Quantifying the Evolution of Early Life* (pp. 3–21). Ed. Laflamme, Marc, Schiffbauer,  
363 James D., Dornbos, Stephen Q. Springer Netherlands (2011).
- 364 36. Shen, B., Dong, L., Xiao, S. & Kowalewski, M. The Avalon explosion: evolution of  
365 Ediacara morphospace. *Science* **319**, 81–84 (2008)
- 366 37. Liu, A. G., McIlroy, D., Antcliffe, J. B. & Brasier, M. D. Effaced preservation in the  
367 Ediacara biota and its implications for the early macrofossil record. *Palaeontology* **54**, 607–  
368 630 (2011)
- 369 38. Narbonne, G. M., Laflamme, M., Greentree, C. & Trusler, P. Reconstructing a lost world:  
370 Ediacaran rangeomorphs from Spaniard's Bay, Newfoundland. *J. of Paleontology* **83**, 503–  
371 523 (2009)

- 372 39. Brasier, M. D. & Antcliffe, J. B. Evolutionary relationships within the Avalonian Ediacara  
373 biota: new insights from laser analysis. *Journal of the Geological Society* **166**, 363-384  
374 (2009).
- 375 40. Matthews, J.J., Liu, A.G. and McIlroy, D.. Post-fossilization processes and their  
376 implications for understanding Ediacaran macrofossil assemblages. *Geological Society,  
377 London, Special Publications*, **448**, 251-269 (2017).
- 378 41. Narbonne, G.M., Laflamme, M., Trusler, P.W., Dalrymple, R.W. and Greentree, C.. Deep-  
379 water Ediacaran fossils from northwestern Canada: taphonomy, ecology, and  
380 evolution. *Journal of Paleontology*, **88**, 207-223 (2014).
- 381 42. R Core Team. *R: A Language and Environment for Statistical Computing*. R Foundation for  
382 Statistical Computing Vienna, Austria (2013)
- 383 43. Baddeley, A. & Turner, R. Spatstat: an R package for analyzing spatial point patterns. *J. of  
384 Statistical Software* **12**, 1–42 (2005)
- 385 44. Berman, M. Testing for spatial association between a point process and another stochastic  
386 process. *Applied Statistics* **35**, 54–62 (1986)
- 387 45. Baddeley, A., Rubak, E. & Møller, J. Score, pseudo-score and residual diagnostics for  
388 spatial point process models. *Statistical Science* **26**, 613–646 (2011)
- 389 46. Wiegand, T. & Moloney, K. Rings, circles, and null-models for point pattern analysis in  
390 ecology. *Oikos* **104**, 209–229 (2004)
- 391 47. Wiegand, T., Kissling, W., Cipriotti, P. & Aguiar, M. Extending point pattern analysis for  
392 objects of finite size and irregular shape. *J. of Ecology* **94**, 825–837 (2006)

- 393 48. Wiegand, T., Moloney, K., Naves, J. & Knauer, F. Finding the missing link between  
394 landscape structure and population dynamics: a spatially explicit perspective. *The American*  
395 *Naturalist* **154**, 605–627 (1999)
- 396 49. Loosmore, N. B. & Ford, E. D. Statistical inference using the G or K point pattern spatial  
397 statistics. *Ecology* **87**, 1925–1931 (2006)
- 398 50. Wiegand, T. & Moloney, K. A. *Handbook of Spatial Point-pattern Analysis in Ecology*. 538  
399 pages. CRC Press (2013)
- 400 51. Levin, S. A. *The problem of pattern and scale in ecology* (pp. 277–326). Springer U.S.  
401 (1995)
- 402 52. Besag, J. Spatial interaction and the statistical analysis of lattice systems. *J. of the Royal*  
403 *Statistical Society. Series B (Methodological)* **36**, 192–236 (1974)
- 404 53. Thomas, M. A generalization of Poisson’s binomial limit for use in ecology. *Biometrika* **36**,  
405 18–25 (1949)
- 406 54. Grabarnik, P., Myllymäki, M. & Stoyan, D. Correct testing of mark independence for  
407 marked point patterns. *Ecological Modelling* **222**, 3888–3894 (2011)
- 408 55. Fraley, C. & Raftery, A. E. *MCLUST version 3: an R package for normal mixture modeling*  
409 *and model-based clustering*. Washington University Seattle Dept of Statistics (2006)
- 410 56. Fraley, C. & Raftery, A. E. Bayesian regularization for normal mixture estimation and  
411 model-based clustering. *J. of Classification* **24**, 155–188 (2007)
- 412 57. Péliissier, R. & Goreaud, F. A practical approach to the study of spatial structure in simple  
413 cases of heterogeneous vegetation. *J. of Vegetation Science* **12**, 99–108 (2001)
- 414 58. Stoyan, D., Kendall, W. S. & Mecke, J. *Stochastic geometry and its applications*. 2nd  
415 edition. 458 pages. Springer Verlag (1995)

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425 prepared the manuscript. E. G. M. conceived and ran the analyses.

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427 The authors declare no competing interests.

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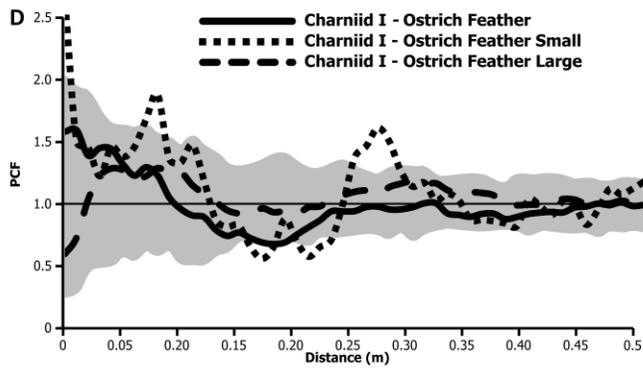
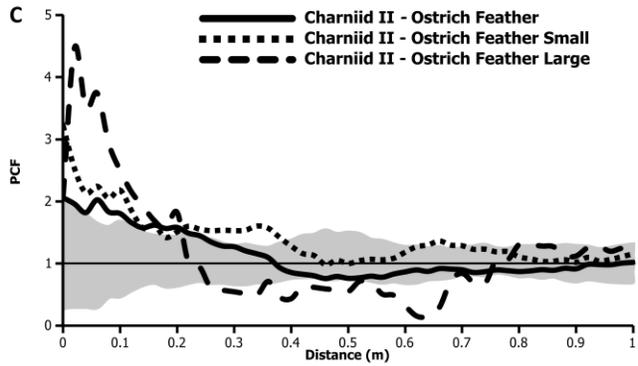
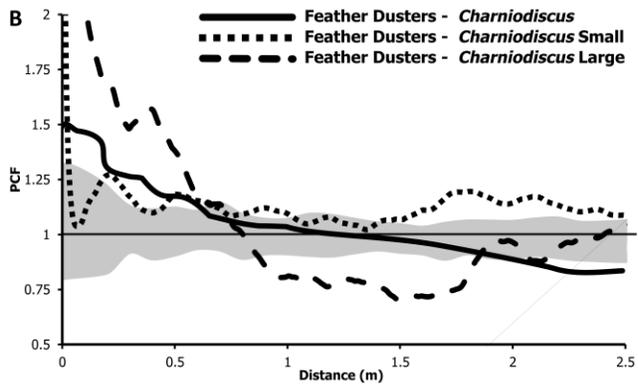
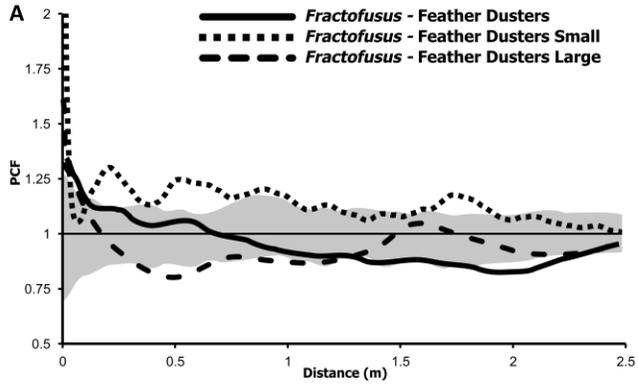
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435 **Figure 1. DVS for Mistaken Point communities.** Height distributions of the **a**, ‘D’ surface  
436 community **b**, ‘E’ surface community and the **c**, LMP surface community, and uptake-zone  
437 distributions of the **d**, ‘D’ surface community **e**, ‘E’ surface community and the **f**, LMP surface  
438 community. The taxonomic group is given on the *x*-axis, and the *y*-axis is the height above the  
439 substrate in millimetres. The shade of the bin is given by the scale to the right of each community  
440 plot, and represents the frequency of specimens at the given height (**a-c**) and the occupation  
441 frequency of specimen uptake-zone (**d-f**). For example, in the height frequency plots (**a-c**), a  
442 56mm tall specimen with or without a stem would feature in the 50-60mm box only. For the  
443 uptake-zone occupancy plots (**d-f**), a non-stemmed specimen 56mm tall would be shown in the  
444 10, 20, 30, 40, 50 and 60mm bins. A stemmed specimen 56mm tall with a 30mm stem would be  
445 shown in the 40, 50 and 60mm bins.

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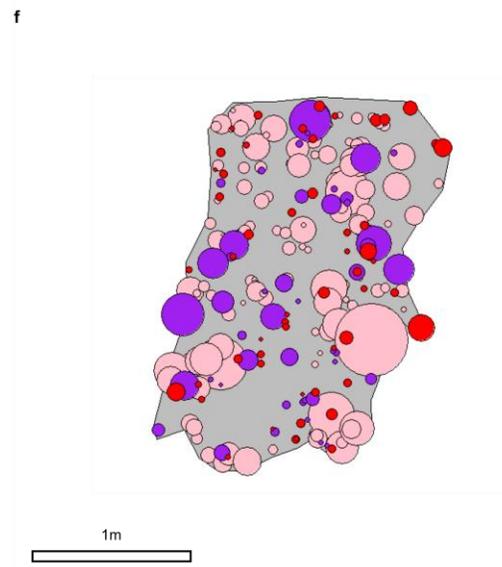
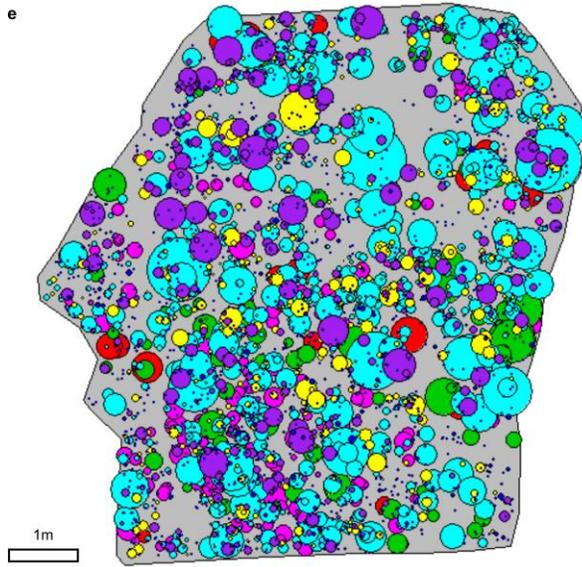
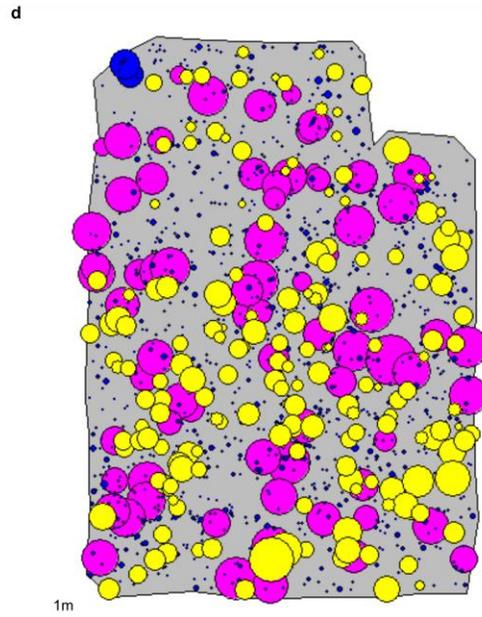
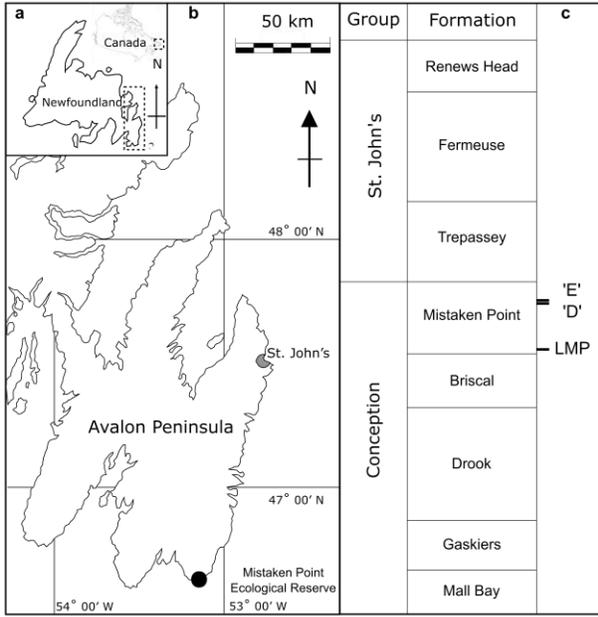
448 **Figure 2. PCF for resource competition interactions.** The x-axis is the inter-point distance  
449 between organisms in meters. On the y-axis, PCF=1 indicates CSR, <1 indicates segregation and  
450 >1 indicates aggregation. The grey shaded area denotes the boundaries of 99 Monte Carlo  
451 simulations of CSR. Since the PCF curves are not within these areas, the complete spatial  
452 randomness (CSR) hypotheses is rejected and one can assume that the distributions on both  
453 surfaces are aggregated at small spatial scales and segregated at large spatial scales. ( $p_d^{Fract-FeaD}$   
454  $< 0.01$ ,  $p_d^{Chard-FeaD} < 0.01$ ,  $p_d^{CharI-IOst} < 0.01$ ,  $p_d^{CharII-IOst} < 0.01$ ). **a**, PCFs for ‘E’ surface  
455 *Fractofusus* – Feather Dusters (1497 *Fractofusus* specimens of which 126 were small and 303  
456 were large and Feather Dusters 362 specimens of which 296 were small and 66 large). **b**, PCFs  
457 for ‘E’ surface *Charniodiscus* – Feather Dusters (*Charniodiscus* 825 specimens of which 489  
458 were small and 336 were large and Feather Dusters 362 specimens of which 296 were small and  
459 66 large). **c**, PCF for the segregated aggregation of the LMP surface (Charniid II 51 specimens of  
460 which 26 were small and 25 were large and Ostrich Feather 54 specimens of which 38 were  
461 small and 16 large). **d**, PCF for the segregated aggregation of the LMP surface (Charniid I 143  
462 specimens of which 47 were small and 25 were large and Ostrich Feather 54 specimens of which  
463 38 were small and 16 large).

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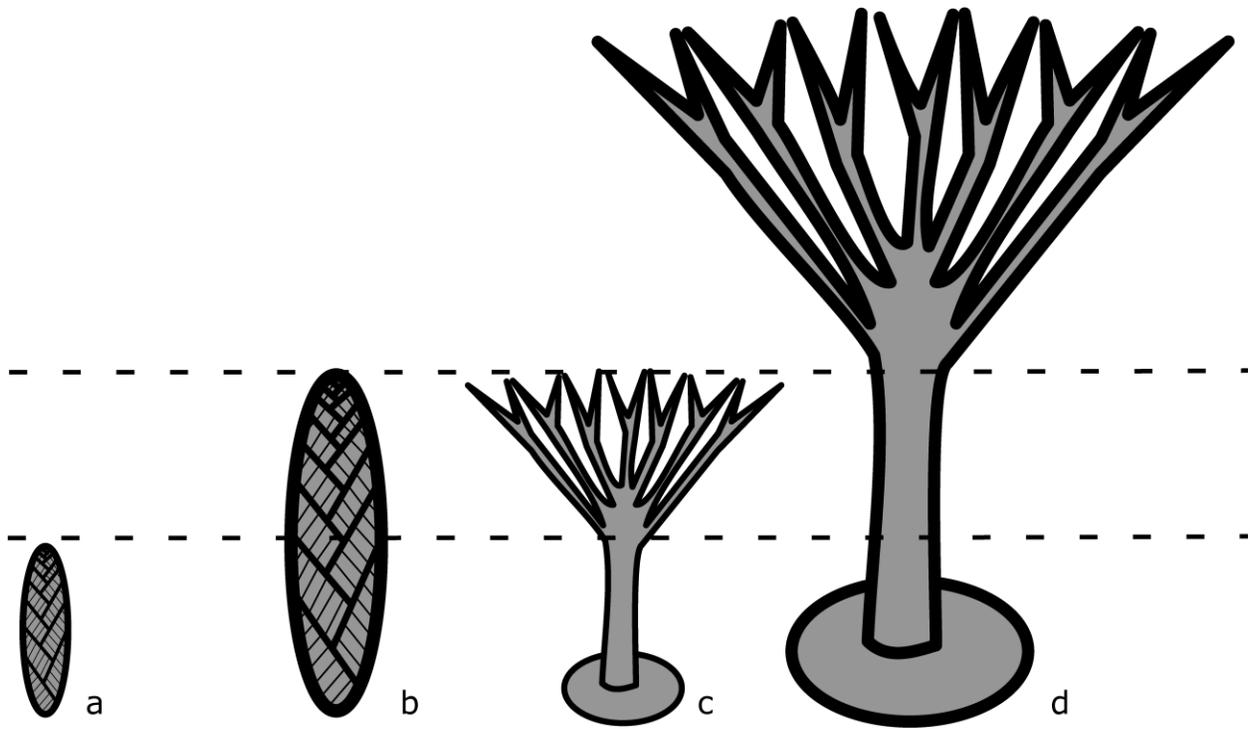
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**Supplementary Figure 1.**

**Map and simplified stratigraphic column showing the position of studied bedding planes with bedding plane maps. a,** Newfoundland, eastern Canada. Dashed area indicates region of interest in b. **b,** The Avalon Peninsula, eastern Newfoundland. Locations of the bedding planes are indicated. **c,** Stratigraphic column (not to scale) of the Avalon Peninsulas. The 'E' surface at Mistaken Point has been dated to  $566 \pm 0.3\text{Ma}$  (ref. 1). **d-e,** Maps of the 'D', 'E' and LMP surfaces showing specimen position and height (circle diameter). **d,** 'D' surface, showing *Fractofusus* (blue), *Pectinifrons* (yellow) and *Bradgatia* (Pink). **e,** 'E' surface with *Charniodiscus* (red), Holdfast discs with stems (orange), Charniid I (green), *Thectardis* (purple), *Fractofusus* (blue), *Bradgatia* (pink) and Feather Dusters (yellow) and **f,** Lower Mistaken Point showing Charniid A (I), Charniids II (purple) and Ostrich Feathers (red). Data from [12]. Scale bar 1m.



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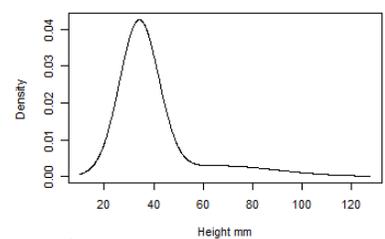
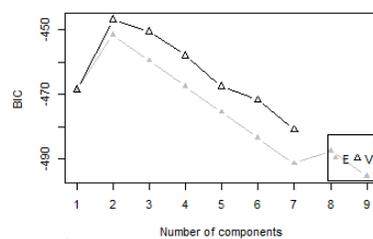
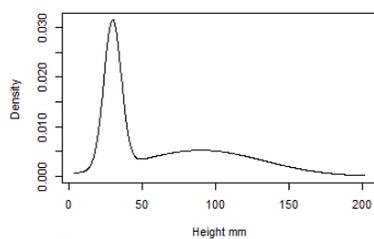
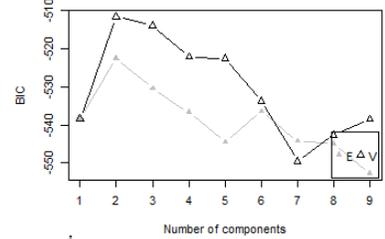
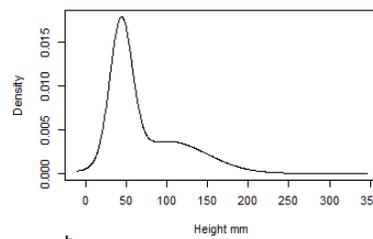
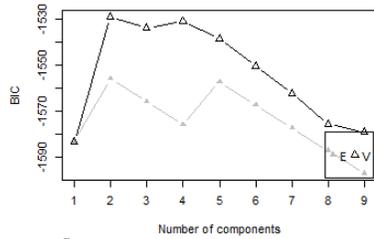
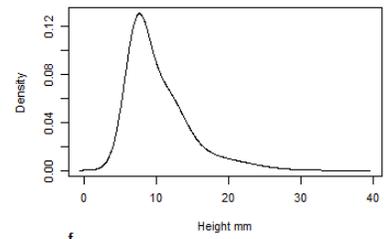
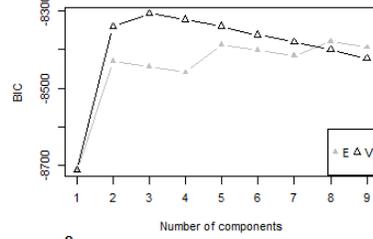
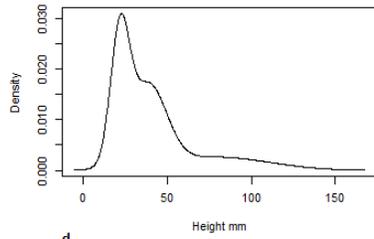
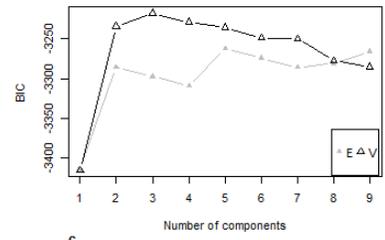
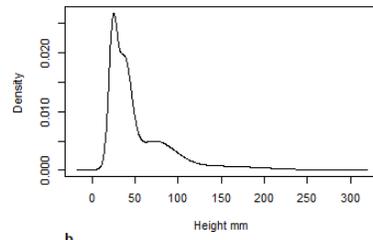
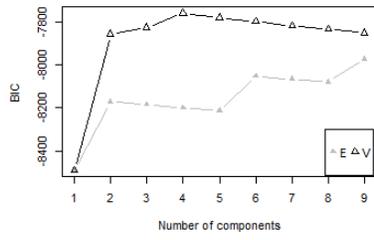
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**Supplementary Figure 2. Diagram illustrating DVS and uptake-zone quantification.** Uptake-zone was defined as the part of the organism which exhibited multiple scales of branching. In specimens i and ii, the uptake-zone consists of the entire height because they lack a naked stem. For specimens iii and iv, the uptake-zone is only the top 50% of the specimen, as the naked stem comprises the other 50%. To calculate *DVS*, the specimens within each taxon population were tabulated into 1cm height bins firstly using their height, and secondly their uptake-zone height ranges. For the above community (consisting of specimens i – iv), for the Charniid specimens (specimens i and ii), specimen i occupies a distinct stratum to the Feather Dusters (specimens iii and iv), while specimen ii height overlaps specimen iii in, and thus does not occupy a distinct stratum from Feather Dusters: consequently, the Charniids have a  $DVS^{height} = 50\%$ . For the Feather Dusters (specimens iii and iv), specimen iii overlaps with ii, so does not occupy a distinct stratum, but specimen iv is not overlapped by any Charniid specimens: so, Feather Duster  $DVS^{height} = 50\%$ . Community  $DVS^{height}$  is the mean of the values for all taxa in the community:  $DVS^{height}_{Community} =$

497 50%. The uptake-zone  $DVS^{uptake}_{Community} = 50\%$  as well, because the uptake-zones of specimens i  
498 and iv occupy distinct strata, but ii and iii do not.

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**Supplementary Figure 3.**

**Size distribution analysis of taxa with segregated bivariate PCFs.** Size distribution analysis of taxa with segregated bivariate PCFs. **a**, 'E' surface *Charniodiscus* height-frequency distributions, and **b**, the results of Bayesian Information Criterion<sup>54,55</sup> (BIC). Triangles and squares correspond to models assuming equal and unequal variance respectively. High BIC values correspond to a good model fit, so the best-fit model is a three component equal variance model. **c**, 'E' surface Feather Dusters height-frequency distributions and **d**, BIC. **e**, 'E' surface *Fractofusus* height-frequency distributions and **f**, BIC. **g**, LMP Charniid I height-frequency distributions, and **h**, BIC. **i** LMP Charniid II height-frequency distributions, and (J), BIC, (K), LMP Ostrich Feathers height-frequency distributions, and **j**, BIC.

Surface Taxon	Height DVS			Uptake-zone DVS		
	D	E	LMP	D	E	LMP
<i>Bradgatia</i>	0.6184	0.0000		0.6184	0.4204	
<b>Charniid</b>		0.0000	0.5232		0.1071	0.6821
<b>Charniid II</b>			0.0784			0.2549
<i>Charniodiscus</i>		0.0776			0.5806	
<b>Feather Dusters</b>		0.0000			0.0359	
<i>Fractofusus</i>	0.9957	0.7963		0.9957	0.8831	
<b>Ostrich Feather</b>			0.0000			0.2778
<i>Pectinifrons</i>	0.8057			0.8057		
<i>Thectardis</i>		0.0000			0.1200	

514 **Supplementary Table 1. Table of DVS values for Mistaken Point communities.** Table of height  
515 and uptake-zone DVS for each taxon population within each of D, E and LMP communities. *DVS*  
516 = 0% corresponds to no specimens occupying a unique part of the water column, i.e. the height  
517 distribution of that population is totally overlapped by other taxa populations. *DVS* =100%  
518 corresponds to no overlap between any specimens, so each taxon occupies a distinct strata.  
519

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Surface	D	E	Lower Mistaken Point
<b>Rangeomorph</b>	96.96%	55.15%	71.82%
<b>Stemmed</b>	0.54%	30.18%	42.27%
<b>Other</b>	2.5%	14.67%	14.09%

521 **Supplementary Table 2.**

522 **Community compositions.** Percentage of taxa from each surface that are rangeomorphs and have  
523 stemmed. The “Other category” refers to taxa which cannot be placed as either Rangeomorphs or  
524 stemmed taxa due to lack of taxonomic certainty.

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Surface	Taxon 1	Taxon 2	PCF <sub>min</sub>	Size class <i>p</i> value	
				Small	Large
E	<i>Fractofusus</i>	Feather Dusters	0.8852	0.25	0.01
E	Feather Dusters	<i>Charniodiscus</i>	0.8972	0.14	0.01
LMP	Charniid I	Ostrich Feather	0.4932	0.02	0.01
LMP	Charniid II	Ostrich Feather	0.5346	0.92	0.01

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**Supplementary Table 3.**

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**Segregation test for the different size-classes of segregated bivariate distributions.** A value of

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$p < 0.05$  is significantly segregated, while  $p > 0.05$  is not significantly segregated.

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Surface	Taxon	$\sigma$ (m)	Mean Height (mm)	Maximum Height (mm)	Mean mid-point of Uptake-zone (mm)	Maximum mid-point of Uptake-zone (mm)
E	<i>Charnidiscus</i>	0.07	54	291	30	58
E	Feather Duster	0.25	41	153	43	106
E	<i>Thectardis</i>	0.18	102	165	16	104
LMP	Charniid II	0.22	63	185	26	93
LMP	Ostrich Feather	0.18	39	118	14	34

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#### Supplementary Table 4.

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**Taxon height and cluster sizes.** The best-fit cluster size for the Thomas Cluster model of each

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frondose taxon exhibiting Thomas Cluster aggregation<sup>4,5</sup>. The mid-point of the active zone height

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is given by calculating the mid-point between the stem and the top of the frond for each specimen.

535

Surface	Top of Stem Height				Uptake-zone height				Top of frond Height			
	Mean		Max		Mean		Max		Mean		Max	
	<i>p</i>	R <sup>2</sup>	<i>p</i>	R <sup>2</sup>	<i>p</i>	R <sup>2</sup>	<i>p</i>	R <sup>2</sup>	<i>p</i>	R <sup>2</sup>	<i>p</i>	R <sup>2</sup>
<b>E</b>	<i>0.47</i>	0.54	<i>0.48</i>	0.54	<i>0.78</i>	0.12	<i>0.28</i>	0.82	<i>0.88</i>	0.04	<b>0.03</b>	<b>1.00</b>

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**Supplementary Table 5.**

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**Linear regression analyses.** Linear regressions of the fitted cluster sizes of Table S3 for frondose

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organisms showing a Thomas Cluster i.e. dispersal process aggregations. The regressions which

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are significant are given in bold. These analyses could not be repeated for LMP surface due to

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insufficient sample size.

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